

## 5. MAGNETOSTRATIGRAPHIC AND BIOSTRATIGRAPHIC SYNTHESIS, LEG 117, ARABIAN SEA<sup>1</sup>

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### ABSTRACT

During the late early Miocene to early middle Miocene, the Owen Ridge was uplifted to a sufficient height as to be above the realm of turbidite deposition. Monsoonal-induced upwelling appears to have been initiated during the Miocene. On the Oman Margin, the effect of upwelling on the microplankton was established by the middle Miocene. However, the effects of upwelling on the Owen Ridge region were not realized until later, in the early late Miocene. A transition in the upwelling regime took place between the Pliocene and Pleistocene. While the Miocene and Pliocene sediments are dominated by the siliceous component, the Pleistocene sediments seem to be dominated by the calcareous component.

### INTRODUCTION

During Leg 117, 25 holes were drilled at 12 sites in the western portion of the Arabian Sea (Fig. 1). Over 4300 m of high-quality core was recovered from a continuum of water depths ranging from above to below the points where the oxygen-minimum zone (OMZ) impinges on the seafloor. Because this portion of the Arabian Sea experiences upwelling of colder, nutrient-enriched waters during the months coincident with the summer monsoon, an examination of the microfossil content of the sediments of different ages and different locations within the region provides important information about the Neogene tectonic history, paleoceanography, and paleoproductivity in this portion of the Indian Ocean. The magnetostratigraphic and biostratigraphic assignments for Leg 117 sediments are shown in Figure 2. In several intervals, it was impossible to make specific zonal assignments due to the lack of some typical tropical zonal markers. These marker species will be discussed in the following section.

### PALEOMAGNETIC AND MICROFOSSIL SUMMARY

#### Magnetostratigraphy

The magnetostratigraphy of Leg 117 sediments was determined using inclination data of discrete samples (Hayashida and Bloemendal, this volume). Usually two specimens per section (1.5 m) were obtained from the sediments recovered by the Advanced Hydraulic Piston Corer (APC) or Extended Core Barrel (XCB) technique at Sites 720 through 731. Because the sedi-

ments at Site 729 were drilled with the Rotary Core Barrel (RCB), no magnetic measurements were made. Magnetic measurements were made on a shipboard flux-gate magnetometer and a shore-based cryogenic magnetometer. The weak and relatively soft magnetic character of the sediments partially restricted the quality and resolution of the magnetostratigraphy, especially of the intervals below approximately 200 mbsf. Gas expansion observed at some Oman Margin sites significantly affected the paleomagnetic record.

The polarity zones, usually identified in the uppermost 200 m of sediment, were assigned to the polarity time scale of Berggren et al. (1985) by referring to the biostratigraphic data. The magnetic record of the Owen Ridge sites (721 and 722) was correlated from the Brunhes (C1) to Chronozone 5 (C3A) or 6 (C3Ar). At these sites, however, polarity intervals of approximately, or less than, 10<sup>5</sup> yr in duration, such as the Olduvai (C2), Kaena (C2A-1), and Mammoth (C2A-2) Subchronozones, were not detected. On the Oman Margin, clear reversal records were observed from the Brunhes (C1) to the Olduvai Subchronozone (C2) at Site 724, and to the Gauss Chronozone (C2A) at Site 727. The Olduvai Subchronozone (C2) was missing at Site 728. Nevertheless, the interval from 50 to 200 mbsf at this site provided the best reversal stratigraphy from the Gauss (C2A) to the Gilbert (C3) Chronozones.

#### Calcareous Nannofossils

The calcareous nannofossil zonation of Martini (1971) was used during Leg 117 for the Miocene and Pliocene sediments. A modified Pleistocene zonation (Takayama and Sato, 1987) was used for the Quaternary sediments (Spaulding, this volume) and further modified by Sato et al. (this volume). Based upon the re-examination of calcareous nannofossils from the Pliocene-Pleistocene boundary stratotype at Le Castella, Italy, the FAD (first appearance datum) of *Gephyrocapsa caribbeanica* is used as a micropaleontological approximation of the Pliocene-Pleistocene boundary for this leg (T. Takayama, pers. comm., 1988).

Calcareous nannofossils are common to abundant in nearly every sample studied. A few samples had rare or few specimens and a few others were entirely barren of nannofossils. Preservation ranges from poor to good with most samples falling somewhere in the moderate category. In some of the lower Miocene sediments asteroliths and sphenoliths are heavily overgrown. Some of the typical tropical Miocene and Pliocene marker species are missing or too rare to be stratigraphically useful. These include representatives of the ceratoliths, amauroliths, astero-

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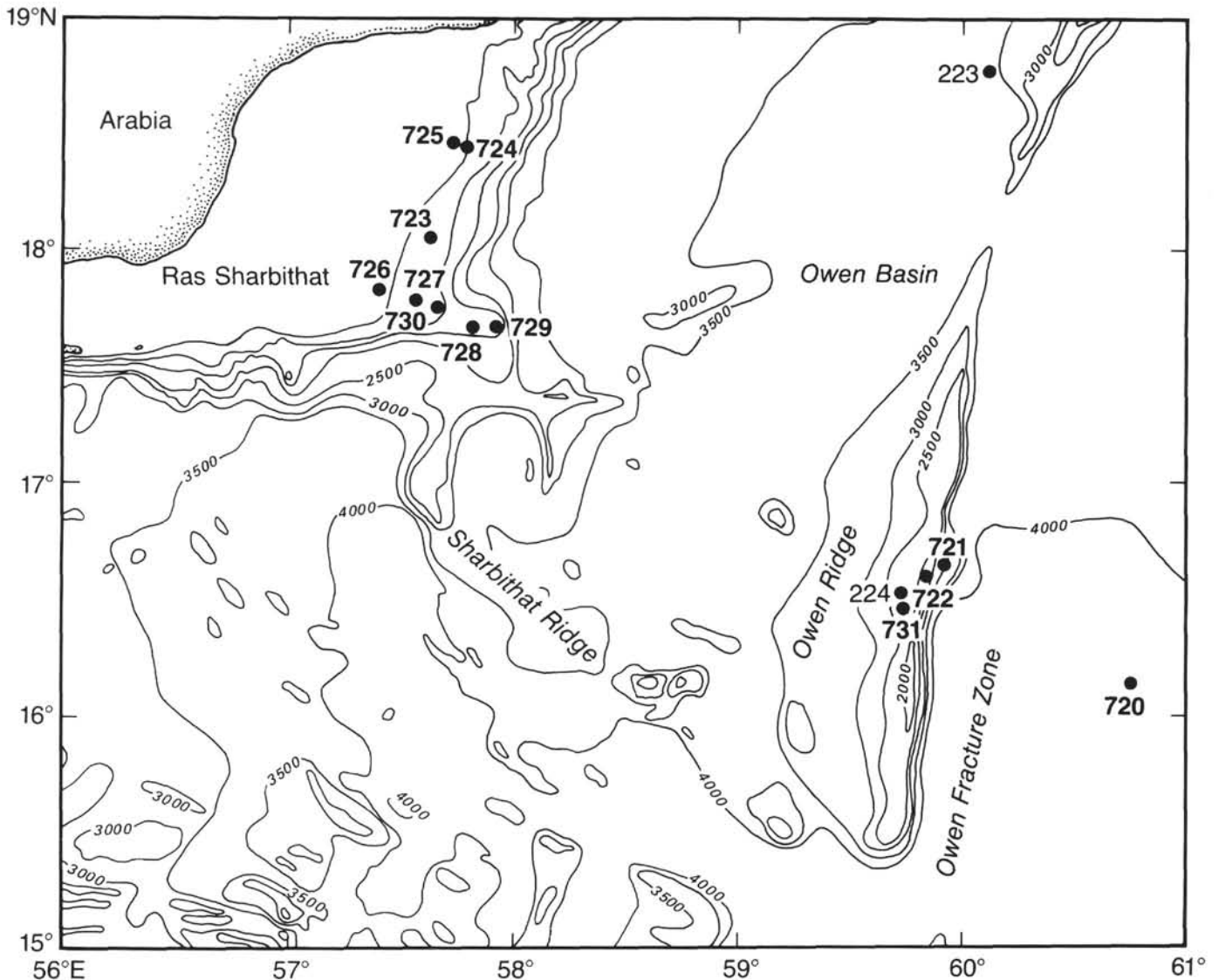


Figure 1. Locations of sites drilled during Leg 117.

liths, and helicosphaerids. The diversity of the nannofossils seems to be somewhat lower than one would expect from a tropical region located from 16°N to 18°N. There seems to be no doubt that the upwelling waters in this part of the Indian Ocean noticeably affect the composition of the nannofossil assemblages. A few reworked Cretaceous and Tertiary specimens were found in over 60% of the samples studied from the Oman Margin. Because the oldest sediments recovered are early Miocene in age, it is suggested that these Cretaceous and Tertiary specimens were carried by the southwest monsoon winds that are known to supply quite significant amounts of clay to the region.

Overall, the calcareous nannofossils provide the most consistently reliable biostratigraphic zonations for Leg 117. Thirty-four nannofossil datums are recognized in the Quaternary, Neogene, and uppermost Paleogene sediments. Six of these datums are new and can be found in Sato et al. (this volume).

#### Planktonic Foraminifers

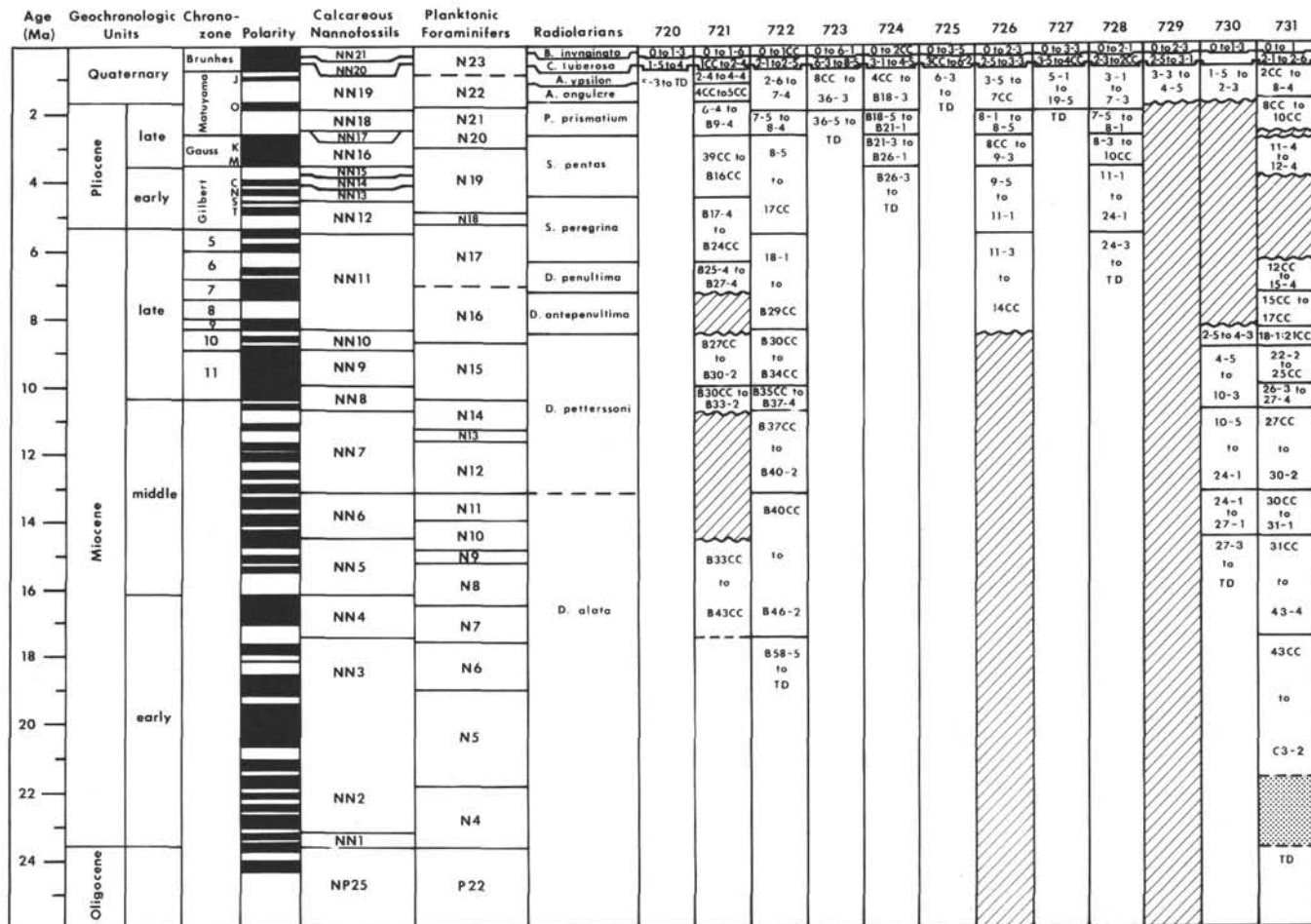
Planktonic foraminifers range from barren or few to abundant at each site. The zonation of Blow (1969) was adhered to as closely as possible during this leg. Some zonal marker species for the Pliocene and Pleistocene are missing or are very rare. Be-

cause the evolutionary series of *Globorotalia tosaensis* to *G. truncatulinooides* is missing in the upper Pliocene to lower Pleistocene sediments, the N21/N22 zonal boundary was approximated by using the LAD (last appearance datum) of *Globigerinoides obliquus*. Also, because *Globorotalia margaritae* and *Globigerinoides fistulosus* are rare the zonation of the Pliocene sediments is hindered. Thirty-two planktonic foraminiferal datums are recognized during this leg.

The planktonic foraminifers from Owen Ridge Sites 721 and 722 provide a complete biostratigraphy. At the other Owen Ridge site (731), the planktonic foraminifers are rare and show signs of substantial dissolution. The sites on the Oman Margin (723–730), in general, contain planktonic foraminifers which have been reduced in numbers as a result of dissolution, diagenesis, or a reduction in their ecological niches.

#### Benthic Foraminifers

Shipboard analyses showed that benthic foraminifers ranged from barren to abundant at each site. Benthic foraminifers occur sporadically and range from poorly- to well-preserved. The foraminifers in the limestone recovered at Sites 726 and 729 are recrystallized but their presence provides the only age control for



fossil stratigraphy. Below a depth of 300 mbsf the absence of biostratigraphic constraints coupled with the lack of sufficient numbers of stable samples prohibits the correlation of the recovered sediments with the polarity reversal time scale.

The turbiditic sediments contain lower concentrations of calcareous nannofossils, planktonic and benthic foraminifers, and radiolarians. Quaternary calcareous nannofossil Zones NN19 to NN21 and planktonic foraminiferal Zones N22 and N23 are recognized. No biostratigraphic zonation based on radiolarians is possible because the assemblage is composed primarily of long-ranging *Spumellaria*. However, robust specimens and diverse assemblages from two pelagic layers within the turbidite sequences in Core 117-720A-30X suggest a cool-water affinity based on the presence of *Eucyrtidium acuminatum* and *Pterocanium praetextum eucolpum*.

*Braarudosphaera bigelowii*, a typical shallow-water calcareous nannofossil (Martini, 1967; Takayama, 1972), and *Ammonia beccarii*, a shallow-water benthic foraminifer (Murray, 1976), are found in samples from the turbiditic sediments.

Reworked Cretaceous and Tertiary calcareous nannofossils and *Nuttaloides truempyi*, a Late Cretaceous to Eocene benthic foraminifer (van Morkhoven et al., 1986), are also found in several turbiditic samples.

The Quaternary sedimentary sequence at Site 720 records fan deposition with some intercalated pelagic oozes (less than 10 m thick). These thin oozes are reflective of changes in turbidite flux which may be related to changes in sea level.

### Owen Ridge (Sites 721, 722, and 731)

#### Quaternary

The pelagic Quaternary sequences at each of these three sites are comprised primarily of calcareous nannofossil ooze. The thickness of the Quaternary sediments is relatively consistent over the length of the Owen Ridge. Pleistocene to Holocene sediments range from 63.50 m thick at Sites 721 and 722 to 84.20 m thick at Site 731. Calcareous nannofossils are abundant with moderate to good preservation in the Quaternary oozes. Planktonic and benthic foraminifers are common to abundant, diverse, and well-preserved. Radiolarians are well-preserved, robust, and abundant at Sites 721 and 722 and are few to common with moderate preservation at Site 731.

#### Neogene

Like the Quaternary sediments, the Pliocene sequences on the Owen Ridge also consist primarily of nannofossil ooze. Complete Pliocene sequences were recovered from Sites 721 and 722. However, based on the radiolarian zonation at Site 731, an unconformity is recognized in the upper Pliocene sediments. This resulted in the elimination of all of the lower Pliocene and a portion of the lowermost upper Pliocene sediments. Calcareous nannofossils are abundant, and planktonic and benthic foraminifers are common to abundant. Radiolarians are abundant in nearly every Pliocene sample.

During the late Miocene, nannofossil ooze was the dominant lithology deposited over the Owen Ridge. Increased amounts of biogenic silica are present in the middle to upper Miocene nannofossil oozes and chalks at all three sites. Benthic foraminifers at Site 722 are commonly pyritized at levels coincident with increased amounts of biogenic silica.

An extensive middle Miocene hiatus is present at Site 721. Uppermost lower Miocene deposition at Sites 721 and 731 and lower to middle Miocene sediments at Site 722 consist of clastic turbidites of silty clays and muds. Preservation of microfossils in the turbiditic sediments is significantly poorer than in the overlying pelagic sediments. Planktonic foraminifers and radio-

larians are absent from the lower Miocene and most of the middle Miocene sediments at Site 721. Few benthic foraminifers with poor preservation are found in the turbiditic sequences. Calcareous nannofossils in the lower Miocene and lowermost middle Miocene sediments are abundant but have poor preservation which is manifested as heavy calcitic overgrowths on the sphenoliths, asteroliths, and *Cyclicargolithus floridanus*. Lowermost Miocene sediments at Site 731 are barren of all microfossils.

### Oman Margin (Sites 723–730)

#### Quaternary

Holes drilled at Sites 723, 724, 726, 727, and 728 recovered complete Quaternary sequences. The Pleistocene sediments at Sites 723, 727, 728, 729, and 730 are nannofossil oozes while the Pleistocene sediments from Sites 724, 725, and 726 are more hemipelagic in nature (nannofossil ooze to sand-silt-clay to calcareous clayey silts to nannofossil foraminifer-rich mud). Sites 724, 725, and 726 are closer to land, and therefore, it is not surprising that they have a more dominant terrigenous fraction. Abundant calcareous nannofossils with moderate to good preservation are present in all of the Pleistocene to Holocene sediments on the Oman Margin. *Coccolithus pelagicus* and *C. crassipons*, typical cold-water taxa, are present in large numbers in the lower Pleistocene sediments from Sites 723, 724, 726, 727, and 728.

*Hyalinea balthica*, a cool-water benthic foraminifer, is present throughout the Quaternary sediments at Sites 723, 724, and 725. High relative abundances of the planktonic foraminifers *Globigerina bulloides*, *Neogloboquadrina dutertrei*, *Globorotalia menardii*, and *Globigerinita glutinata* indicate that upwelling conditions existed at Site 723 during the Pleistocene. The high P/B (planktonic/benthic) foraminiferal ratio and the stable assemblage composition of benthic foraminifers suggest that no significant changes in the water depth have occurred during the Pleistocene at Site 727. The P/B ratio together with the benthic foraminiferal faunal composition suggest that during the Pleistocene Site 729 was located in the upper middle bathyal zone.

Radiolarians have the most sporadic occurrence of any of the microfossils studied in detail. At Site 723 all of the typical tropical Pleistocene radiolarian zonal markers are missing. However, the assemblage in Sample 117-723A-5H-CC contains the Arabian Upwelling Assemblage of Johnson and Nigrini (1982). At Site 725 the uppermost 10 cores are barren of radiolarians, but samples below 95.3 mbsf have a significant siliceous fauna and flora which include radiolarians, sponge spicules, and diatoms. Again, some tropical marker species are missing. However, a more diverse fauna is found at Site 725 than at Site 724. *Dictyophimus infabricatus*, an upwelling form found in the eastern equatorial Pacific, is relatively abundant at Site 725.

#### Neogene

Upper Pliocene sediments at Site 723 have high relative abundances of planktonic foraminifers *Globigerina bulloides*, *Neogloboquadrina dutertrei*, *Globorotalia menardii*, and *Globigerinita glutinata* which indicate upwelling conditions existed at this site during the late Pliocene.

Few calcareous nannofossils with poor preservation are present throughout much of the upper Pliocene calcareous clayey silts at Site 724. Zones NN17 and NN18 have been combined due to the absence of the low-latitude marker species, *Discoaster pentaradiatus*. Low-diversity radiolarian assemblages are characteristic of most of the upper Pliocene sediments at Site 724.

During the late Pliocene silty clays to nannofossil ooze to chalk alternating with foraminifer-rich clayey silts were deposited at Site 726. Calcareous nannofossil Zones NN16 and NN17

were combined in the absence of the low-latitude marker species, *Discoaster surculus*. Planktonic foraminifers are rare and exhibit poor preservation.

Biogenic opal is a common component of the marly nannofossil ooze which was deposited at Site 728 during the late Pliocene. This is indicative of increased paleoproductivity during these times. Hiatuses at Sites 729 and 730 preclude the recognition of upper Pliocene sediments.

*Coccolithus pelagicus* and *C. crassipons*, typical cold-water taxa, are abundant in the upper Pliocene sediments at Sites 723, 724, 726, 727, and 728. *Hyalinea balthica*, a benthic foraminifer found in cold water masses, is present in the upper Pliocene sediments at this Sites 724 and 728. The genus *Helicosphaera* is abundant in upper Pliocene samples from Sites 723, 724, 725, and 728. This genus is thought to have preferred regions of upwelling (Perch-Nielsen, 1985).

Calcareous clayey silts were also deposited at Site 724 during the early Pliocene. These contain abundant calcareous nannofossils and common planktonic foraminifers. Fewer radiolarians are present than in the upper Pliocene sediments from this site. *Ammonia beccarii*, a benthic foraminifer which today is found in waters less than 350 m (Murray, 1976) is found in the lower Pliocene sediments at Sites 724 and 728.

The lower Pliocene sediments at Site 726 are similar to those deposited during the late Pliocene. Calcareous nannofossil Zones NN12-NN15 were combined at Sites 726 and 728 because the marker species are missing. Planktonic foraminiferal Zones N19 through N21 were also combined for the same reason. The lower Pliocene sediments at Site 728 contain common biogenic opal. *Hyalinea balthica*, a cold-water benthic foraminifer, is present in the uppermost lower Pliocene sediments at Site 728.

Lower Pliocene sediments are not present at Sites 729 and 730 due to hiatuses at these sites.

During the latter part of the late Miocene, rare to abundant calcareous nannofossils were deposited at Site 726. The benthic foraminifer, *Bolivina seminuda*, comprises up to 95% of the assemblage. A hiatus at Site 726 precludes the recognition of lowest upper Miocene sediments.

Common biogenic opal is present in upper Miocene sediments at Site 728. Planktonic foraminifers are rare to few, which results in the combination of Zones N16 and N17. The dominance of thick-walled *Globorotalia tumida tumida* indicates that dissolution has affected the planktonic foraminiferal assemblage. The dissolution process may have been affected by an intensified or expanded oxygen-minimum zone during this time.

A hiatus at Site 730 removed the uppermost upper Miocene sediments. Siliceous microfossils are found in the upper Miocene sediments that are present at this site.

Site 730 is the only site on the Oman Margin which provided a record of the deposition which occurred during the early and middle Miocene. Chalks which contain a relatively high carbonate and foraminiferal content make up the lower and middle Miocene sediments. Calcareous nannofossils *Reticulofenestra pseudumbilica* and *R. gelida* occur together in nearly every sample in Zones NN11 through NN15. Backman (1980) has suggested that the presence of both of these forms of *Reticulofenestra* in the same samples suggests that there was sufficient seasonality to allow the growth of both of these forms. The colder water associated with the annual upwelling is probably responsible for the presence of *R. gelida* in samples from Hole 728A.

Early Miocene nannofossils possess poor preservation which is manifested by heavy overgrowths on sphenoliths, discoasters, and *Cyclicargolithus floridanus*. Calcareous nannofossil Zones NN4 and NN5 are combined due to the absence of the tropical marker species, *Helicosphaera ampliapertura*. Planktonic foraminiferal Zones N13 and N14 are combined due to the absence of the marker species which delineates the top of Zone N13. Plank-

tonic foraminifers are common to abundant in lower Miocene sediments.

Hiatuses at Sites 726 and 729 prevent the recognition of Oligocene to middle Miocene sediments.

Drilling at Sites 726 and 729 recovered large-foraminifer- to algae-bearing carbonates. The presence of abundant benthic foraminifers including *Nummulites* cf. *planulatus*, *N.* cf. *pratti*, and *Actinocyclus* sp. provide an Eocene age assignment for this unit. Amphistegid and nummulitid type larger foraminifers suggest that this unit was deposited in shallow water (less than 130 m).

Based on the benthic foraminiferal faunas, it can be deduced that the seafloor at Site 728 has subsided more than 1000 m since the early Pliocene. The subsidence at Site 724 has been less drastic, probably a deepening on the order of 250 m. Conversely, Site 725 appears to have undergone some uplift during the middle Pleistocene. The shallow-water benthic foraminifer, *Ammonia beccarii*, is present in only the upper 85 m of sediment at Hole 725C.

## MAGNETOSTRATIGRAPHY

### Quaternary

When compared to the calcareous nannofossil datums, the Brunhes/Matuyama Chronozone boundary should be observed between 100 and 150 mbsf in Hole 723A. A few reversed samples occur over this interval, but this boundary reversal is not clearly recorded in the sediments at this site. In the lower Pleistocene sediments some reversely magnetized samples exist, but most of the Matuyama Chronozone cannot be recognized. Overall, the Pleistocene sediments at Site 723 are poor recorders of geomagnetic field reversals, and minimal magnetostratigraphic data can be derived from them.

The Brunhes/Matuyama Chronozone boundary and the Jaramillo Subchronozone are tentatively identified at Site 724. The top of the Olduvai Subchronozone is also recognized.

The remanent magnetization of sediments at Site 725 is low, as at Sites 723 and 724. Comparison with calcareous nannofossil stratigraphy indicates that the Brunhes/Matuyama Chronozone boundary may be present at 79 mbsf. The Jaramillo Subchronozone is considered to be present between 100 and 121 mbsf at Site 725.

At Site 726, the base of the Jaramillo Subchronozone is tentatively recognized at 39 mbsf.

At Hole 727A the inclination data indicates the presence of at least four polarity intervals. The Brunhes Chronozone is present between 0 and 76 mbsf. The sediments below 76 mbsf are assigned to the Matuyama Chronozone. The Jaramillo Subchronozone (C1r-1) is recognized between 95.40-95.83 mbsf and 104.14-104.69 mbsf. The Olduvai Subchronozone (C2) is present between approximately 153.0 and 182.4 mbsf.

A comparatively complete magnetostratigraphic record is present at Site 728. The Brunhes/Matuyama Chronozone boundary and Jaramillo Subchronozone are recognized. However, the Olduvai Subchronozone appears to be missing.

Because the RCB system was used at Site 729, no attempt was made to measure the magnetic signal of these sediments.

### Neogene

The bottom of the Olduvai Subchronozone and the Matuyama/Gauss Chronozone boundaries are recognized at Site 724. The Matuyama/Gauss and Gauss/Gilbert Chronozone boundaries, the top of the Kaena, and the base of the Mammoth Subchronozones are recorded at Site 728.

A reversed interval below 75 mbsf at Site 726 corresponds in time to several reversed and normal intervals. No correlations

were attempted with the geomagnetic time scale because of this anomalous relationship.

The Cochiti, Nunivak, Sidufjall, and Thvera Subchronozones are recorded in sediments at Site 728. The interval between 40 and 110 mbsf is characterized by higher intensities than those of the sediments above and below. A long normal interval is present between 50 and 110 mbsf. The upper part of this normally polarized interval may correlate with Chron C5 (Chronozone 11).

### CHRONOSTRATIGRAPHY

The magnetic anomaly time scale of Berggren et al. (1985) was utilized for assigning ages to the polarity reversals recorded in the Leg 117 sediments. These ages were obtained through the correlation of Anomaly 5 with Chron 11. Ages for some calcareous nannofossil and planktonic foraminifer datums have been calculated based on the sediments recovered on this leg. These new ages (Table 1) can be found in Kroon et al. (this volume), Sato et al. (this volume), and Spaulding (this volume). Radiolarian event ages are taken from Johnson and Nigrini (1985) and Johnson et al. (1989).

The top of the acme of the calcareous nannofossil, *Reticulofenestra asanoi*, occurs within or just below the Brunhes/Matuyama reversal boundary on the Oman Margin. Also on the margin, the LAD of *Gephyrocapsa parallela* occurs just above the top of the Jaramillo Subchronozone. However, on the Owen Ridge the FAD of *G. parallela* occurs just above the bottom of the Jaramillo Subchronozone. The LAD of large *Gephyrocapsa* occurs just below the Jaramillo Subchronozone on the Owen Ridge and the Oman Margin. However, at another site on the margin, this datum occurs at levels coincident with the bottom of the Jaramillo Subchronozone. *Helicosphaera sellii* has its LAD between the Jaramillo and Olduvai Subchronozones on the Oman Margin.

Many of the ages of Neogene planktonic foraminifer events (Kroon et al., this volume) appear to be diachronous. This is consistent with the conclusions reached by Dowsett (1989). Obviously anomalous planktonic foraminifer ages include those in the *Neogloboquadrina-Pulleniatina* lineages. The FAD of *Globorotalia truncatulinoides*, *G. tosaensis*, and *G. crassaformis* occur in the Arabian Sea (Kroon et al., this volume) much later than in other regions (Hills and Thierstein, 1989).

As can be seen in Table 3, 4, 5, 11, and 13, the order of radiolarian events is quite consistent in sediments younger than 4.5 Ma. In older sediments, however, the order in which these events occur becomes much more unstable. This is probably due to the fact that the ages younger than 4.5 Ma have been derived paleomagnetically (Johnson et al., 1989) from piston cores.

### SEDIMENTATION RATES

#### Indus Fan

The sedimentation rate at Site 720 (Fig. 3) is based solely on calcareous nannofossil datums (Table 2). The uppermost 5 m show a sedimentation rate of 21 m/m.y. This corresponds to an interval of nannofossil ooze deposition on the Indus Fan. In the turbiditic sequence the sedimentation rate ranges between 133 and 600 m/m.y.

#### Owen Ridge

The sedimentation rate curve for Site 721 (Fig. 4) is based on calcareous nannofossils, planktonic foraminifers, radiolarians, and magnetostratigraphy (Table 3). The sedimentation rate at this site is rather uniform in the uppermost 247 m, ranging between 25 and 59 m/m.y. Between 247 and 261 mbsf the sedimentation rate drops to 7 m/m.y. From 261 mbsf to 320 mbsf the sedimentation rate ranges between 30 and 37 m/m.y. The basal part of this site (347–424.2 mbsf) consists of turbiditic sequences of clayey siltstones to mudstones. No datums are avail-

**Table 1. Ages of polarity reversal boundaries and biostratigraphic events for Leg 117. (T = upper limit, B = lower limit, / = evolutionary transition).<sup>a</sup>**

|     | Event                                     | Age (Ma)   | Source of age <sup>a</sup> |
|-----|---|------------|----------------------------|
| P1  | B <i>Globorotalia theyeri</i>             | 0.12; 0.36 | 1                          |
| N1  | T <i>Helicosphaera inversa</i>            | 0.15       | 2                          |
| N2  | B <i>Emiliana huxleyi</i>                 | 0.23; 0.24 | 3; 2                       |
| R1  | T <i>Stylatractus universus</i>           | 0.37–0.47  | 4                          |
| R2  | B <i>Collosphaera tuberosa</i>            | 0.40–0.59  | 4                          |
| N3  | T <i>Pseudoemiliana lacunosa</i>          | 0.38; 0.39 | 3; 2                       |
| N4  | B <i>Helicosphaera inversa</i>            | 0.48       | 2                          |
| P2  | T <i>Globorotalia tosaensis</i>           | 0.51; 0.64 | 1                          |
| P3  | B <i>Globorotalia crassaformis</i>        | 0.60; 0.74 | 1                          |
| R3  | T <i>Anthocyrtidium nosicae</i>           | 0.66–0.78  | 4                          |
| R4  | T <i>Pterocorys campanula</i>             | 0.66–0.78  | 4                          |
| P4  | B <i>Globigerinella calida</i>            | 0.69; 0.74 | 1                          |
| P5  | B <i>Globigerinoides tenellus</i>         | 0.69; 0.89 | 1                          |
| M1  | Brunhes/Matuyama                          | 0.73       | 5                          |
| R5  | B <i>Pterocorys hertwigii</i>             | 0.76–0.84  | 4                          |
| N5  | T <i>Reticulofenestra asanoi</i>          | 0.70; 0.83 | 3; 2                       |
| N6  | B <i>Gephyrocapsa parallela</i>           | 0.85; 0.89 | 3; 2                       |
| M2  | T Jaramillo                               | 0.91       | 5                          |
| R6  | T <i>Anthocyrtidium angulare</i>          | 0.94–1.04  | 4                          |
| M3  | B Jaramillo                               | 0.98       | 5                          |
| P6  | T <i>Neogloboquadrina acostaensis</i>     | 1.01; 1.10 | 1                          |
| R7  | B <i>Lamprocyrtis nigriniae</i>           | 1.02–1.07  | 4                          |
| N7  | B Acme <i>Reticulofenestra asanoi</i>     | 1.06       | 2                          |
| R8  | T <i>Lamprocyrtis neoheteroporos</i>      | 1.09–1.13  | 4                          |
| P7  | B <i>Globorotalia tosaensis</i>           | 1.10; 1.40 | 1                          |
| N8  | T <i>Gephyrocapsa</i> (large)             | 1.07; 1.10 | 3; 2                       |
| N9  | T <i>Helicosphaera sellii</i>             | 1.19; 1.34 | 2; 3                       |
| N10 | B <i>Gephyrocapsa</i> (large)             | 1.36       | 2                          |
| R9  | T <i>Pterocanium prismatium</i>           | 1.52–1.56  | 4                          |
| R10 | B <i>Anthocyrtidium angulare</i>          | 1.52–1.56  | 4                          |
| N11 | T <i>Calcidiscus macintyreii</i>          | 1.47; 1.57 | 3; 2                       |
| N12 | B <i>Gephyrocapsa oceanica</i>            | 1.53; 1.57 | 3; 2                       |
| M4  | T Olduvai                                 | 1.66       | 5                          |
| N13 | B <i>Gephyrocapsa caribbeanica</i>        | 1.66; 1.80 | 2; 3                       |
| P8  | B <i>Globorotalia truncatulinoides</i>    | 1.64; 1.85 | 1                          |
| M5  | B Olduvai                                 | 1.88       | 5                          |
| N14 | T <i>Discoaster brouweri</i>              | 1.91; 2.03 | 2; 3                       |
| N15 | B acme <i>Gephyrocapsa</i>                | 2.13       | 2                          |
| P9  | T <i>Globigerinoides obliquus</i>         | 2.20; 2.39 | 1                          |
| N16 | T acme <i>Crenalithus doronicoideus</i>   | 2.25       | 2                          |
| N17 | T <i>Discoaster pentaradiatus</i>         | 2.27; 2.31 | 2; 3                       |
| N18 | T <i>Discoaster surculus</i>              | 2.42; 2.49 | 3; 2                       |
| R11 | B <i>Cycladophora davisiana</i>           | 2.42–2.44  | 4                          |
| R12 | B <i>Theocorythium trachelium</i>         | 2.40–2.50  | 6                          |
| M6  | Matuyama/Gauss                            | 2.47       | 5                          |
| R13 | B <i>Lamprocyrtis neoheteroporos</i>      | 2.51–2.53  | 4                          |
| P10 | T <i>Globoquadrina altispira</i>          | 2.57; 2.66 | 1                          |
| N19 | T <i>Discoaster tamalis</i>               | 2.62; 2.77 | 2; 3                       |
| N20 | T <i>Reticulofenestra ampla</i>           | 2.62       | 2                          |
| P11 | T <i>Neogloboquadrina humerosa</i>        | 2.66; 2.81 | 1                          |
| P12 | B <i>Neogloboquadrina dutertrei</i>       | 2.91; 3.30 | 1                          |
| M7  | T Kaena                                   | 2.92       | 5                          |
| N21 | T lower acme <i>C. doronicoideus</i>      | 2.97       | 2                          |
| P13 | T <i>Globorotalia limbata</i>             | 3.11; 3.30 | 1                          |
| P14 | coiling change <i>Pulleniatina</i> (?)    | 3.11; 3.69 | 1                          |
| M8  | B Mammoth                                 | 3.18       | 5                          |
| R15 | T <i>Phormostichoartus fistula</i>        | 3.26–3.28  | 4                          |
| R16 | T <i>Lychnodictyum auidax</i>             | 3.33–3.35  | 4                          |
| M9  | Gauss/Gilbert                             | 3.40       | 5                          |
| P15 | B <i>Globigerinoides ruber</i>            | 3.42; 3.65 | 1                          |
| N22 | T <i>Sphenolithus abies</i>               | 3.44       | 3                          |
| P16 | T <i>Sphaeroidinellopsis</i> spp.         | 3.47; 3.57 | 1                          |
| R17 | T <i>Phormostichoartus doliolum</i>       | 3.53–3.55  | 4                          |
| N23 | T <i>Reticulofenestra pseudoubilica</i>   | 3.44; 3.56 | 3; 2                       |
| P17 | B <i>Pulleniatina obliquiloculata</i>     | 3.65; 3.80 | 1                          |
| R18 | B <i>Amphirhopalum ypsilon</i>            | 3.77–3.79  | 4                          |
| R20 | B <i>Spongaster tetras tetras</i>         | 3.83–3.85  | 4                          |
| M10 | T Cochiti                                 | 3.88       | 5                          |
| M11 | B Cochiti                                 | 3.97       | 5                          |
| M12 | T Nunivak                                 | 4.10       | 5                          |
| P18 | B <i>Neogloboquadrina nigriniae</i>       | 4.18; 4.45 | 1                          |
| M13 | B Nunivak                                 | 4.24       | 5                          |
| R25 | <i>S. berminghamsi</i> → <i>S. pentas</i> | 4.3–4.4    | 6                          |
| M14 | T Sidufjall                               | 4.40       | 5                          |
| M15 | B Sidufjall                               | 4.47       | 5                          |
| P19 | B <i>Sphaeroidinella dehiscentis</i>      | 4.54; 4.55 | 1                          |
| M16 | T Thvera                                  | 4.57       | 5                          |

Table 1 (continued).

|     | Event  | Age (Ma)   | Source of age <sup>a</sup> |
|-----|--|------------|----------------------------|
| R27 | T <i>Solenosphaera omnitubus</i>             | 4.7–4.8    | 6                          |
| M17 | B Thvera                                     | 4.77       | 5                          |
| R28 | T <i>Botryostrobus bramletti</i>             | 4.9–5.0    | 6                          |
| R39 | T <i>Dendrospyrus bursa</i>                  | 5.0–5.1    | 6                          |
| P20 | B <i>Pulleniatina primalis</i>               | 5.18; 5.28 | 1                          |
| P21 | B <i>Neogloboquadrina tegillata</i>          | 5.18; 5.28 | 1                          |
| P22 | B <i>Globorotalia tumida tumida</i>          | 5.28; 5.34 | 1                          |
| R31 | T <i>Acrobotrys tritubus</i>                 | 5.3–5.4    | 6                          |
| M18 | Gilbert/Chronozone 5                         | 5.35       | 5                          |
| P23 | B <i>Globigerinoides sacculifer</i>          | 5.43; 5.68 | 1                          |
| P24 | N. <i>acostaensis</i> → <i>Pulleniatina</i>  | 5.45; 5.55 | 1                          |
| P25 | B <i>Globorotalia margaritae</i>             | 5.50; 5.68 | 1                          |
| N24 | T <i>Discoaster quinqueringus</i>            | 5.6        | 5                          |
| P26 | B <i>Neogloboquadrina humerosa</i>           | 5.55; 5.67 | 1                          |
| N25 | T <i>Discoaster berggrenii</i>               | 5.6        | 5                          |
| R33 | T <i>Stichocorys johnsonii</i>               | 5.7–5.8    | 6                          |
| M19 | Chronozone 5/6                               | 5.89       | 5                          |
| P27 | B <i>Globorotalia plesiotumida</i>           | 5.98; 5.99 | 1                          |
| R35 | S. <i>delmontensis</i> → <i>S. peregrina</i> | 6.1–6.7    | 6                          |
| R37 | T <i>Calocycletta caepa</i>                  | 6.2–6.6    | 6                          |
| R40 | B <i>Solenosphaera omnitubus</i>             | 6.3–6.5    | 6                          |
| R42 | T <i>Diartus hughesi</i>                     | 7.1–7.2    | 6                          |
| R43 | B <i>Acrobotrys tritubus</i>                 | 7.7–7.8    | 6                          |
| R47 | B <i>Spongaster berminghami</i>              | 7.9–8.0    | 6                          |
| R48 | T <i>Stichocorys wolfii</i>                  | 8.0–8.2    | 6                          |
| R49 | T <i>Botryostrobus miralestensis</i>         | 8.1–8.2    | 6                          |
| R50 | T <i>Diartus petterssoni</i>                 | 8.1–8.2    | 6                          |
| N26 | B <i>Discoaster quinqueringus</i>            | 8.2        | 5                          |
| R51 | D. <i>petterssoni</i> → <i>D. hughesi</i>    | 8.3–8.5    | 6                          |
| R52 | B <i>Diartus hughesi</i>                     | 8.7–8.8    | 6                          |
| R53 | B <i>Botryostrobus bramletti</i>             | 8.8–9.0    | 6                          |
| N28 | T <i>Discoaster hamatus</i>                  | 8.85       | 5                          |
| N29 | T <i>Catinaster coalitus</i>                 | 9.0        | 5                          |
| N30 | B <i>Discoaster hamatus</i>                  | 10.0       | 5                          |
| R55 | T <i>Cyrtocapsella japonica</i>              | 10.0–10.3  | 6                          |
| P28 | T <i>Globorotalia mayeri</i>                 | 10.4       | 5                          |
| R56 | B <i>Diartus petterssoni</i>                 | 10.6–10.8  | 6                          |
| N31 | B <i>Catinaster coalitus</i>                 | 10.8       | 5                          |
| P29 | B <i>Globigerina nepenthes</i>               | 11.3       | 5                          |
| P30 | T <i>Globorotalia fohsi</i>                  | 11.5       | 5                          |
| R57 | B <i>Phormostichoartus doliolum</i>          | 11.1–11.9  | 6                          |
| N33 | T <i>Cyclicargolithus floridanus</i>         | 11.6       | 6                          |
| R59 | T <i>Cyrtocapsella cornuta</i>               | 11.6–11.9  | 6                          |
| R60 | T <i>Lithopera renzae</i>                    | 12.1–12.3  | 6                          |
| N32 | B <i>Discoaster kugleri</i>                  | 13.1       | 5                          |
| N34 | T <i>Sphenolithus heteromorphus</i>          | 14.4       | 5                          |
| P31 | T <i>Globorotalia peripheronda</i>           | 14.6       | 5                          |
| P32 | B <i>Orbulina universa</i>                   | 15.2       | 5                          |
| N35 | T <i>Sphenolithus belemnus</i>               | 17.1       | 5                          |

<sup>a</sup> Sources of ages are as follows: 1 = Kroon et al. (this volume); 2 = Sato et al. (this volume); 3 = Spaulding (this volume); 4 = Johnson and Nigrini (1985); 5 = Berggren et al. (1985); 6 = Johnson et al. (1989).

able for this part of the sequence, and therefore, no accumulation rate can be calculated.

The sedimentation rate curve for Site 722 (Fig. 5) is based on calcareous nannofossils, planktonic foraminifers, radiolarians, and magnetostratigraphy (Table 4). The sedimentation rate at this site, like Site 721, is rather consistent, ranging from 23 to 51 m/m.y. A relatively uniform sedimentation rate is not unexpected for the accumulation of pelagic nannofossil oozes and chalks. The lower sequence of sediments at this site (411.1–565.6 mbsf) consists of interbedded fine-grained turbidites and marly nannofossil chalks. Because no stratigraphic datums are recognized within this interval, it is not included in Figure 5.

The sedimentation rate curve for Site 731 (Fig. 6) is based on calcareous nannofossils, planktonic foraminifers, radiolarians, and magnetostratigraphy (Table 5). The sedimentation rate at this site on the Owen Ridge has fluctuated since the early Miocene. The accumulation rate ranges from 18 to 49 m/m.y. A hiatus, of up to 2 m.y. in duration, is present at approximately

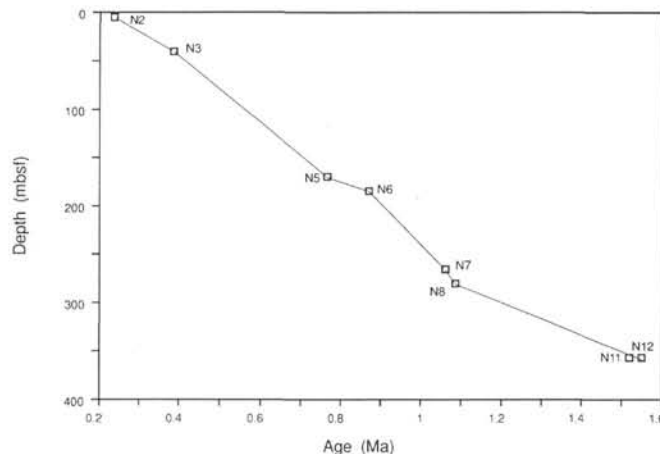


Figure 3. Sedimentation rate curve for Site 720.

Table 2. Biostratigraphic events at Site 720.

| Event                                    | Hole 720A depth (mbsf) |
|--|------------------------|
| Calcareous nannofossils                  |                        |
| N2 B <i>Emiliania huxleyi</i>            | 4.00–6.00              |
| N3 T <i>Pseudoemiliania lacunosa</i>     | 38.20–42.64            |
| N5 T <i>Reticulofenestra asanoi</i>      | 164.00–175.53          |
| N6 B <i>Gephyrocapsa parallela</i>       | 183.40–185.60          |
| N7 B acme <i>Reticulofenestra asanoi</i> | 260.50–270.10          |
| N8 T <i>Gephyrocapsa</i> (large)         | 279.80–280.80          |
| N11 T <i>Calcidiscus macintyreii</i>     | 347.14–366.24          |
| N12 B <i>Gephyrocapsa oceanica</i>       | 347.14–366.24          |

100 mbsf. The Pleistocene sedimentation rate is comparable, but a bit higher than the rates at Sites 721 and 722.

### Oman Margin

The sedimentation rate curve for Site 723 (Fig. 7) is based primarily on calcareous nannofossils with one planktonic foraminifer datum (Table 6). The accumulation rate ranges between 122 and 194 m/m.y. This site accumulated sediments at a rate faster than any of the other sites of this leg, with the exception of the Indus Fan site (720). These rates which are typical of hemipelagic sedimentation are, on the average, about five times higher than the rate of pelagic accumulation at the Owen Ridge sites.

The sedimentation rate curve for Site 724 (Fig. 8) is based primarily on calcareous nannofossils (Table 7). The rates of accumulation range between 75 and 94 m/m.y. for the uppermost 192.55 m of sediment. From 192.55 to 240 mbsf the rate is lower, approximately 40 m/m.y. The sedimentation rate at this site is about one-half that of Site 723. This is probably due to the fact that Site 723 is located in the center of a large slope basin while this site is located in the center of a smaller and shallower slope basin.

The sedimentation rate curve for Site 725 (Fig. 9) is based solely on calcareous nannofossil datums (Table 8). The sedimentation rate ranges from 98 to 160 m/m.y. Between 100 and 103 mbsf, the sedimentation rate is only 29 m/m.y.; it is possible that a hiatus is present within this interval.

The sedimentation rate curve for Site 726 (Fig. 10) is based on calcareous nannofossils and planktonic foraminifers (Table 9). The sedimentation rate for the upper 71 m of sediment ranges from 28 to 63 m/m.y. This higher sedimentation rate is for silty clays and calcareous oozes with varying amounts of foraminifers. In the lower half of Hole 726A (71–118 mbsf) the sedimen-

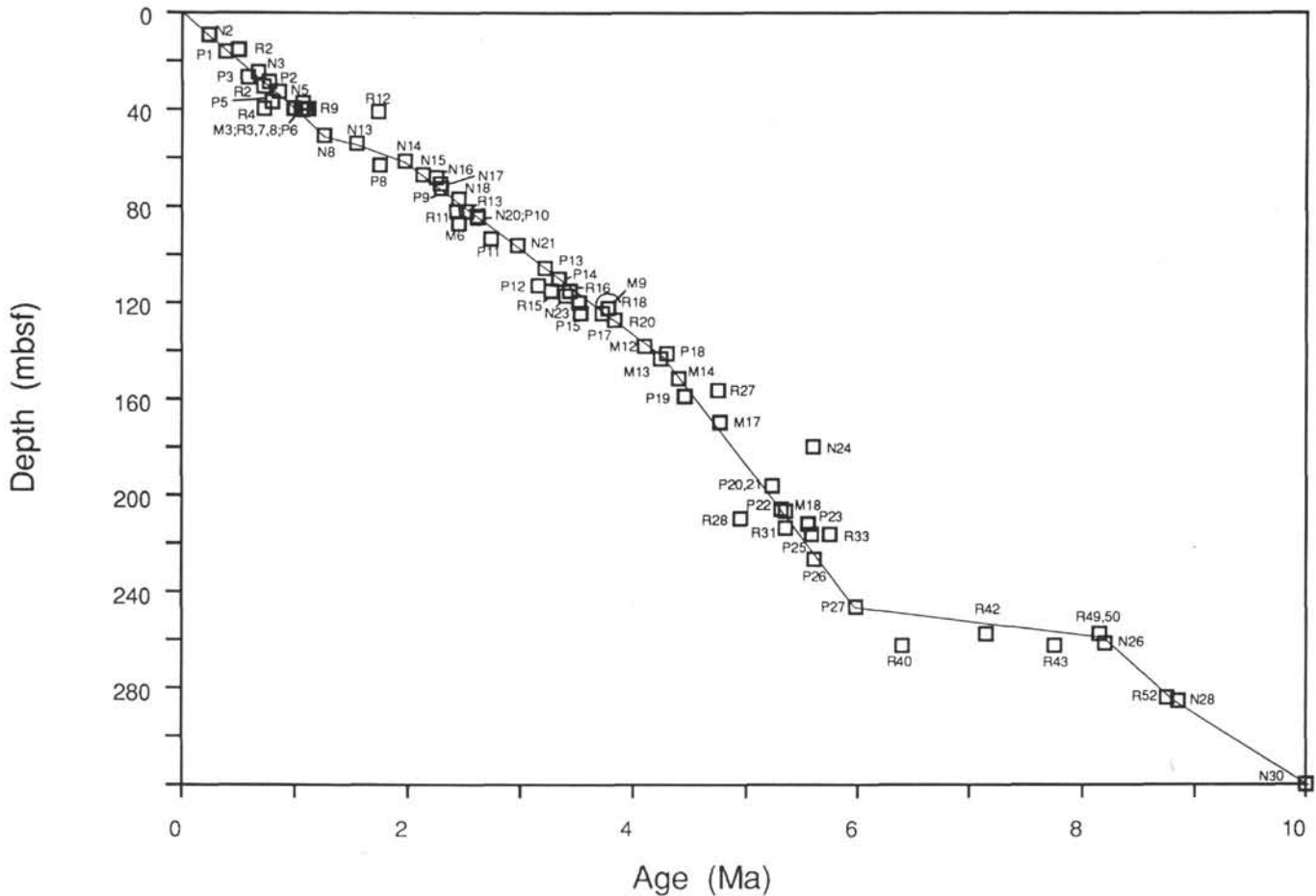


Figure 4. Sedimentation rate curve for Site 721.

tation rate is much lower, about 4–9 m/m.y. This much lower accumulation rate is for lithologies which contain lag deposits of foraminifers, fish teeth, and fish bones which are interbedded with  $C_{org}$ -rich nannofossil-foraminifer oozes. This lower rate of sedimentation is not unexpected for sediments which possess features that are diagnostic of non-deposition and winnowing.

The sedimentation rate curve for Site 727 (Fig. 11) is based on calcareous nannofossils, planktonic foraminifers, and magnetostratigraphy (Table 10). The accumulation rates range from 53 to 130 m/m.y. Site 723, located nearby, accumulated sediments at a rate two times that of this site, which is located near the edge of the same basin.

The sedimentation rate curve for Site 728 (Fig. 12) is based on calcareous nannofossils, planktonic foraminifers, radiolarians, and magnetostratigraphy (Table 11). The accumulation rate for the last 3.56 m.y. ranges between 13 and 52 m/m.y. A higher sedimentation rate (84 m/m.y.) existed between 3.56 and 5.75 Ma. This higher sedimentation rate during the late Miocene and early Pliocene and the concomitant deposition of sediments with accumulations of  $C_{org}$  and siliceous microfossils may be the result of increased primary productivity during this time. The sedimentation rate between 5.75 and 8.15 Ma ranged from 18 to 48 m/m.y. This site, which is located in a lower slope basin, accumulates sediments of a more strictly pelagic nature than the other sites on the Oman Margin. This accounts for its lower overall sedimentation rate.

The sedimentation rate curve for Site 729 (Fig. 13) is based solely on calcareous nannofossil datums (Table 12). The curve represents the sedimentation rate of Lithologic Unit I (nanno-

fossil-foraminifer-rich mud to marly nannofossil ooze) only. The sedimentation rate at this site ranges from 8 to 48 m/m.y. The lower hemipelagic sedimentation rate is probably due to the effects of winnowing. Evidence for this are silty foraminiferal sand layers which range from 0.2 to 1.5 m in thickness.

The sedimentation rate curve for Site 730 (Fig. 14) is based on calcareous nannofossils, planktonic foraminifers, and radiolarians (Table 13). The upper Pleistocene sediments at this site were deposited at a rate of 15 m/m.y. A hiatus is present at approximately 14 mbsf which separates the Pleistocene and upper Miocene sediments. The Miocene units have sedimentation rates which range between 22 and 45 m/m.y. These rates are similar to those calculated for pelagic sedimentation on the Owen Ridge.

#### SUMMARY AND CONCLUSIONS

Very limited data are available regarding calcareous nannofossils in upwelling regions. However, an examination of the species lists of taxa from upwelling regions in the Arabian Sea (DSDP Leg 23) and the Gulf of Aden (DSDP Leg 24) indicates that ceratoliths, amauroliths, and discoasters are less abundant in sediments deposited in upwelling regions. Conversely, *Coccolithus pelagicus* is a more dominant component of the same assemblages. Cold water related to upwelling along the western edge of the Arabian Sea most likely accounts for the general scarcity of ceratoliths and amauroliths in sediments from Sites 720 through 731. The abundance of *Coccolithus pelagicus* in the upper Pliocene to lower Pleistocene sediments at Sites 723, 724, 726, 727, and 728 may be explained by the occurrences of glaciations between 2.5 and 1.8 m.y. ago, which were at least



Table 3. Biostratigraphic and paleomagnetic events at Site 721.

| Event                   |  | Hole 721A depth<br>(mbsf) | Hole 721B depth<br>(mbsf) |
|-------------------------|--|---------------------------|---------------------------|
| Calcareous nannofossils |  |                           |                           |
| N2                      | B <i>Emiliana huxleyi</i>                    | 8.65–9.80                 |                           |
| N3                      | T <i>Pseudoemiliana lacunosa</i>             | 15.45–16.95               |                           |
| N5                      | T <i>Reticulofenestra asanoi</i>             | 28.12–29.20               |                           |
| N6                      | B <i>Gephyrocapsa parallela</i>              | 31.85–33.35               |                           |
| N7                      | B acme <i>Reticulofenestra asanoi</i>        | 36.54–38.04               |                           |
| N8                      | T <i>Gephyrocapsa</i> (large)                | 38.90–40.05               |                           |
| N13                     | B <i>Gephyrocapsa caribbeanica</i>           | 40.05–41.55               |                           |
| N14                     | T <i>Discoaster brouweri</i>                 | 60.45–61.95               |                           |
| N15                     | B acme <i>Gephyrocapsa</i>                   | 66.45–67.33               |                           |
| N16                     | T acme <i>Crenalithus doronicoides</i>       | 67.33–68.45               |                           |
| N17                     | T <i>Discoaster pentaradiatus</i>            | 69.95–71.45               |                           |
| N18                     | T <i>Discoaster surculus</i>                 | 74.45–79.45               |                           |
| N20                     | T <i>Reticulofenestra ampla</i>              | 83.95–85.45               |                           |
| N21                     | T lower acme <i>Crenalithus doronicoides</i> |                           | 95.60–96.75               |
| N23                     | T <i>Reticulofenestra pseudoumbilica</i>     |                           | 114.80–115.95             |
| N24                     | T <i>Discoaster quinqueramus</i>             |                           | 178.85–180.05             |
| N26                     | B <i>Discoaster quinqueramus</i>             |                           | 260.65–262.17             |
| N28                     | T <i>Discoaster hamatus</i>                  |                           | 281.53–289.00             |
| N30                     | B <i>Discoaster hamatus</i>                  |                           | 319.15–320.65             |
| Planktonic foraminifers |  |                           |                           |
| P1                      | B <i>Globorotalia theyeri</i>                | 14.96                     |                           |
| P3                      | B <i>Globorotalia crassaformis</i>           | 24.66                     |                           |
| P2                      | T <i>Globorotalia tosaensis</i>              | 26.62                     |                           |
| P4                      | B <i>Globigerinella calida</i>               | 30.47                     |                           |
| P5                      | B <i>Globigerinoides tenellus</i>            | 36.81                     |                           |
| P6                      | T <i>Neogloboquadrina acostaensis</i>        | 40.16                     |                           |
| P7                      | B <i>Globorotalia tosaensis</i>              | 50.66                     |                           |
| P8                      | B <i>Globorotalia truncatulinoides</i>       | 62.96                     |                           |
| P9                      | T <i>Globigerinoides obliquus</i>            | 72.46                     |                           |
| P10                     | T <i>Globoquadrina altispira</i>             | 83.91                     |                           |
| P11                     | T <i>Neogloboquadrina humerosa</i>           |                           | 93.44                     |
| P14                     | coiling change <i>Pulleniatina</i> (?)       |                           | 105.50                    |
| P13                     | T <i>Globorotalia limbata</i>                |                           | 105.50                    |
| P12                     | B <i>Neogloboquadrina dutertrei</i>          |                           | 112.93                    |
| P16                     | T <i>Sphaeroidinellopsis</i> spp.            |                           | 119.96                    |
| P15                     | B <i>Globigerinoides ruber</i>               |                           | 124.47                    |
| P17                     | B <i>Pulleniatina obliquiloculata</i>        |                           | 124.47                    |
| P18                     | B <i>Neogloboquadrina nigriniae</i>          |                           | 140.96                    |
| P19                     | B <i>Sphaeroidinella dehiscens</i>           |                           | 158.66                    |
| P20                     | B <i>Pulleniatina primalis</i>               |                           | 195.89                    |
| P21                     | B <i>Neogloboquadrina tegillata</i>          |                           | 195.89                    |
| P22                     | B <i>Globorotalia tumida tumida</i>          |                           | 205.66                    |
| P23                     | B <i>Globigerinoides sacculifer</i>          |                           | 206.85                    |
| P25                     | B <i>Globorotalia margaritae</i>             |                           | 216.04                    |
| P24                     | N. <i>acostaensis</i> → <i>Pulleniatina</i>  |                           | 219.37                    |
| P26                     | B <i>Neogloboquadrina humerosa</i>           |                           | 226.55                    |
| P27                     | B <i>Globorotalia plesiotumida</i>           |                           | 246.30                    |
| Radiolarians            |  |                           |                           |
| R2                      | B <i>Collosphaera tuberosa</i>               | 12.55–17.90               |                           |
| R1                      | T <i>Stylatractus universus</i>              | 22.25–31.95               |                           |
| R3                      | T <i>Anthocyrtidium nosicae</i>              | 37.30–41.65               |                           |
| R4                      | T <i>Pterocorys campanula</i>                | 37.30–41.65               |                           |
| R6                      | T <i>Anthocyrtidium angulare</i>             | 37.30–41.65               |                           |
| R7                      | B <i>Lamprocyrtis nigriniae</i>              | 37.30–41.65               |                           |
| R8                      | T <i>Lamprocyrtis neoheteroporos</i>         | 37.30–41.65               |                           |
| R9                      | T <i>Pterocanium prismatium</i>              | 51.25–56.60               |                           |
| R10                     | B <i>Anthocyrtidium angulare</i>             | 51.25–56.60               |                           |
| R12                     | B <i>Theocorythium trachelium</i>            | 51.25–56.60               |                           |
| R11                     | B <i>Cycladophora davisiana</i>              | 79.55–84.90               |                           |
| R13                     | B <i>Lamprocyrtis neoheteroporos</i>         | 79.55–84.90               |                           |
| R14                     | T <i>Stichocorys peregrina</i>               | 84.90–89.15               |                           |
| R15                     | T <i>Phormostichoartus fistula</i>           |                           | 105.20–114.80             |
| R16                     | T <i>Lychnodictyum audax</i>                 |                           | 105.20–114.80             |
| R17                     | T <i>Phormostichoartus doliolum</i>          |                           | 120.15–124.40             |
| R18                     | B <i>Amphirhopalum ypsilon</i>               |                           | 120.15–124.40             |
| R19                     | T <i>Spongodiscus ambus</i>                  |                           | 120.15–124.40             |
| R20                     | B <i>Spongaster tetras tetras</i>            |                           | 124.40–129.75             |
| R26                     | T <i>Didymocyrtis penultima</i>              |                           | 129.75–134.10             |
| R23                     | T <i>Dictyophimus splendens</i>              |                           | 153.50–158.85             |
| R27                     | T <i>Solenosphaera omnitubus</i>             |                           | 153.50–158.85             |
| R30                     | T <i>Stichocorys delmontensis</i>            |                           | 163.20–168.55             |
| R29                     | T <i>Siphostichartus corona</i>              |                           | 202.22–207.35             |

Table 3 (continued).

| Event  | Hole 721A depth (mbsf) | Hole 721B depth (mbsf) |
|--|------------------------|------------------------|
| Radiolarians (cont.)                             |                        |                        |
| R28 T <i>Botryostrobus bramlettei</i>            |                        | 207.35–211.70          |
| R31 T <i>Acrobotrys tritubus</i>                 |                        | 211.70–215.55          |
| R33 T <i>Stichocorys johnsoni</i>                |                        | 215.55–216.79          |
| R34 T <i>Didymocyrtis antepenultima</i>          |                        | 225.15–226.56          |
| R35 <i>S. delmontensis</i> → <i>S. peregrina</i> |                        | 226.56–235.85          |
| R36 B <i>Spondiscus ambus</i>                    |                        | 240.10–245.45          |
| R42 T <i>Diartus hughesi</i>                     |                        | 255.15–259.60          |
| R49 T <i>Botryostrobus miralestensis</i>         |                        | 255.15–259.60          |
| R50 B <i>Diartus petterssoni</i>                 |                        | 255.15–259.60          |
| R39 T <i>Dendrosphyris bursa</i>                 |                        | 259.60–264.95          |
| R40 B <i>Solenosphaera omnitubus</i>             |                        | 259.60–264.95          |
| R41 B <i>Stichocorys peregrina</i>               |                        | 259.60–264.95          |
| R43 B <i>Acrobotrys tritubus</i>                 |                        | 259.60–264.95          |
| R46 T <i>Didymocyrtis laticonus</i>              |                        | 264.95–269.40          |
| R52 B <i>Diartus hughesi</i>                     |                        | 279.20–289.00          |
| Polarity reversals                               |                        |                        |
| M3 B Jaramillo                                   | 38.55–40.05            |                        |
| M6 Matuyama/Gauss                                | 79.76–80.15            | 85.05–86.35            |
| M9 Gauss/Gilbert                                 |                        | 116.65–117.45          |
| M12 T Nunivak(?)                                 |                        | 137.45–138.25          |
| M13 B Nunivak(?)                                 |                        | 141.95–144.14          |
| M14 T Sidufjall                                  |                        | 150.95–151.66          |
| M17 B Thvera                                     |                        | 167.35–171.85          |
| M18 Gilbert/5                                    |                        | 206.15–106.85          |

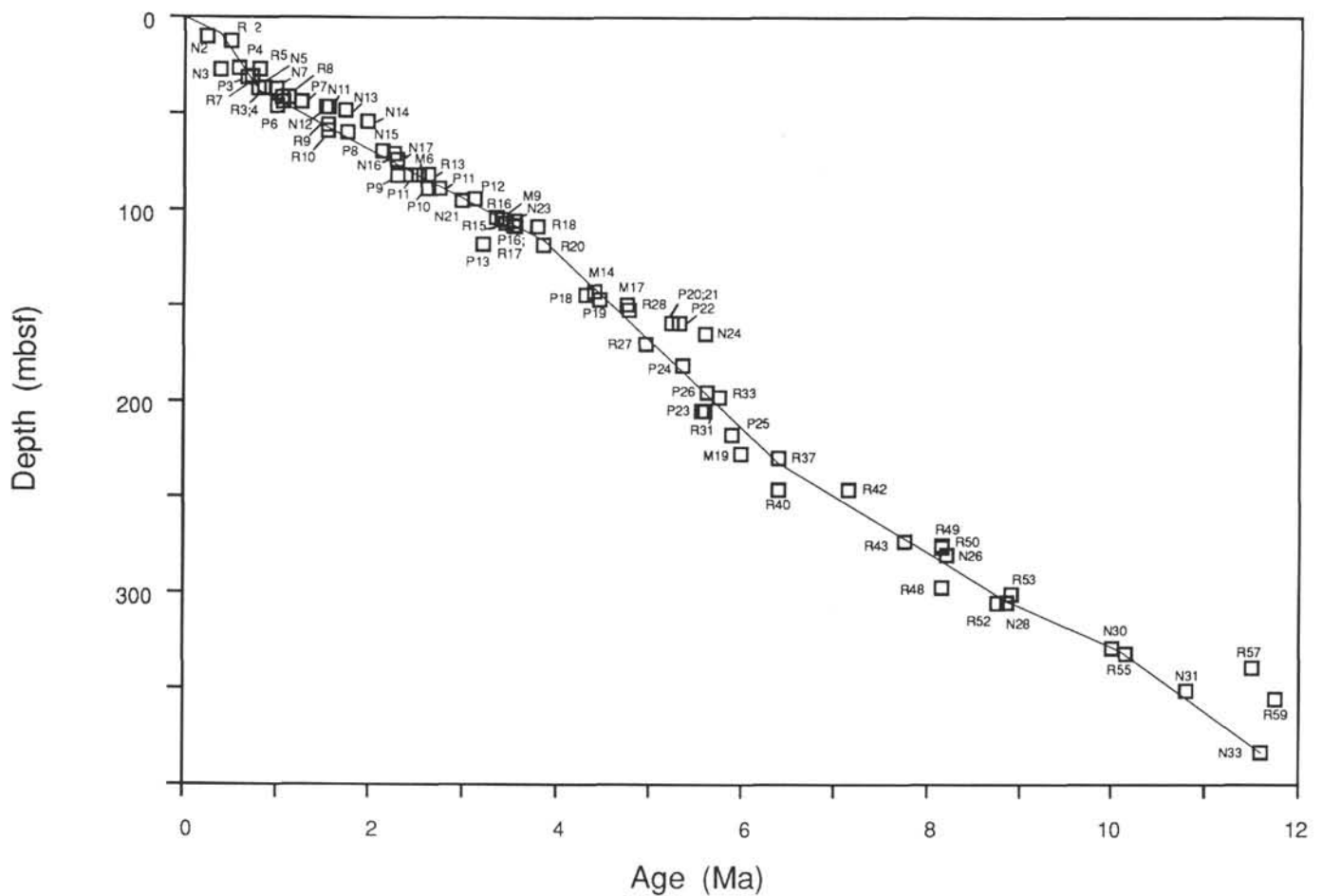


Figure 5. Sedimentation rate curve for Site 722.

Table 4. Biostratigraphic and paleomagnetic events at Site 722.

| Event                   |  | Hole 722A depth<br>(mbsf) | Hole 722B depth<br>(mbsf) |
|-------------------------|--|---------------------------|---------------------------|
| Calcareous nannofossils |  |                           |                           |
| N2                      | B <i>Emiliana huxleyi</i>                    | 9.80–10.95                |                           |
| N3                      | T <i>Pseudoemiliana lacunosa</i>             | 26.55–27.70               |                           |
| N5                      | T <i>Reticulofenestra asanoi</i>             | 31.68–33.15               |                           |
| N6                      | B <i>Gephyrocapsa parallela</i>              | 36.15–37.30               |                           |
| N7                      | B acme <i>Reticulofenestra asanoi</i>        | 39.75–41.25               |                           |
| N8                      | T <i>Gephyrocapsa</i> (large)                | 42.75–44.25               |                           |
| N11                     | T <i>Calcidiscus macintyreii</i>             | 45.75–47.25               |                           |
| N12                     | B <i>Gephyrocapsa oceanica</i>               | 45.75–47.25               |                           |
| N13                     | B <i>Gephyrocapsa caribbeanica</i>           | 47.25–48.75               |                           |
| N14                     | T <i>Discoaster brouweri</i>                 | 53.14–54.66               |                           |
| N15                     | B acme <i>Gephyrocapsa</i>                   | 68.35–69.85               |                           |
| N16                     | T acme <i>Crenalithus daronicoides</i>       | 69.85–71.35               |                           |
| N17                     | T <i>Discoaster pentaradiatus</i>            | 72.84–74.35               |                           |
| N20                     | T <i>Reticulofenestra ampla</i>              | 80.83–81.95               |                           |
| N21                     | T lower acme <i>Crenalithus daronicoides</i> | 93.73–96.20               |                           |
| N23                     | T <i>Reticulofenestra pseudumbilica</i>      | 105.90–107.06             |                           |
| N24                     | T <i>Discoaster quinquerramus</i>            | 163.90–165.05             |                           |
| N26                     | B <i>Discoaster quinquerramus</i>            |                           | 275.60–285.30             |
| N28                     | T <i>Discoaster hamatus</i>                  |                           | 304.70–305.85             |
| N30                     | B <i>Discoaster hamatus</i>                  |                           | 324.00–333.70             |
| N31                     | B <i>Catinaster coalitus</i>                 |                           | 349.05–353.10             |
| N32                     | B <i>Discoaster kugleri</i>                  |                           | 375.10–382.20             |
| N33                     | T <i>Cyclicargolithus floridanus</i>         |                           | 382.20–384.02             |
| N35                     | T <i>Sphenolithus belemnus</i>               |                           | 433.15–552.28             |
| Planktonic foraminifers |  |                           |                           |
| P1                      | B <i>Globorotalia theyeri</i>                | 5.16                      |                           |
| P2                      | B <i>Globorotalia tosaensis</i>              | 21.56                     |                           |
| P4                      | B <i>Globigerinella calida</i>               | 29.01                     |                           |
| P5                      | B <i>Globigerinoides tenellus</i>            | 29.01                     |                           |
| P3                      | B <i>Globorotalia crassaformis</i>           | 31.16                     |                           |
| P6                      | T <i>Neogloboquadrina acostaensis</i>        | 43.76                     |                           |
| P7                      | B <i>Globorotalia tosaensis</i>              | 43.76                     |                           |
| P8                      | B <i>Globorotalia truncatulinoidea</i>       | 59.66                     |                           |
| P9                      | T <i>Globigerinoides obliquus</i>            | 81.96                     |                           |
| P10                     | T <i>Globoquadrina altispira</i>             | 88.66                     |                           |
| P11                     | T <i>Neogloboquadrina humerosa</i>           | 88.66                     |                           |
| P12                     | B <i>Neogloboquadrina dutertrei</i>          | 94.18                     |                           |
| P15                     | B <i>Globigerinoides ruber</i>               | 105.66                    |                           |
| P16                     | T <i>Sphaeroidinellopsis</i> spp.            | 108.06                    |                           |
| P14                     | coiling change <i>Pulleniatina</i> (?)       | 111.05                    |                           |
| P13                     | T <i>Globorotalia limbata</i>                | 117.76                    |                           |
| P17                     | B <i>Pulleniatina obliquiloculata</i>        | 120.76                    |                           |
| P18                     | B <i>Neogloboquadrina nigrinia</i>           | 144.33                    |                           |
| P19                     | B <i>Sphaeroidinella dehiscens</i>           | 146.73                    |                           |
| P20                     | B <i>Pulleniatina primalis</i>               | 158.80                    |                           |
| P21                     | B <i>Neogloboquadrina tegillata</i>          | 158.80                    |                           |
| P22                     | B <i>Globorotalia tumida tumida</i>          | 158.80                    |                           |
| P24                     | <i>N. acostaensis</i> → <i>Pulleniatina</i>  | 188.46                    |                           |
| P26                     | B <i>Neogloboquadrina humerosa</i>           | 195.16                    |                           |
| P23                     | B <i>Globigerinoides sacculifer</i>          | 204.86                    |                           |
| P25                     | B <i>Globorotalia margaritae</i>             | 227.16                    |                           |
| Radiolarians            |  |                           |                           |
| R2                      | B <i>Collosphaera tuberosa</i>               | 9.80–15.15                |                           |
| R5                      | B <i>Pterocorys hertwigii</i>                | 24.75–29.00               |                           |
| R7                      | B <i>Lamprocyrtis nigrinia</i>               | 38.60–43.95               |                           |
| R8                      | T <i>Lamprocyrtis neoheteroporos</i>         | 38.60–43.95               |                           |
| R3                      | T <i>Anthocyrtidium nosicace</i>             | 43.95–48.20               |                           |
| R4                      | T <i>Pterocorys campanula</i>                | 43.95–48.20               |                           |
| R6                      | T <i>Anthocyrtidium angulare</i>             | 43.95–48.20               |                           |
| R12                     | B <i>Theocorythium trachelium</i>            | 48.20–50.55               |                           |
| R9                      | T <i>Pterocanium prismatium</i>              | 53.55–57.50               |                           |
| R10                     | B <i>Anthocyrtidium angulare</i>             | 57.50–59.85               |                           |
| R11                     | B <i>Cycladophora davisiana</i>              | 76.80–86.50               |                           |
| R13                     | B <i>Lamprocyrtis neoheteroporos</i>         | 76.80–86.50               |                           |
| R14                     | T <i>Stichocorys peregrina</i>               | 76.80–86.50               |                           |
| R15                     | T <i>Phormostichoartus fistula</i>           | 101.55–105.90             |                           |
| R16                     | T <i>Lychnodictyum audax</i>                 | 101.55–105.90             |                           |
| R17                     | T <i>Phormostichoartus doliolum</i>          | 105.90–111.25             |                           |
| R18                     | B <i>Amphirhopalum ypsilon</i>               | 105.90–111.25             |                           |
| R20                     | B <i>Spongaster tetras tetras</i>            | 115.60–120.95             |                           |
| R25                     | <i>S. berminghamsi</i> → <i>S. pentas</i>    | 115.60–120.95             |                           |
| R23                     | T <i>Dictyophimus splendens</i>              | 125.30–130.65             |                           |
| R24                     | T <i>Spongodiscus ambus</i>                  | 130.65–134.90             |                           |
| R26                     | T <i>Didymocyrtis penultima</i>              | 134.90–140.25             |                           |

Table 4 (continued).

| Event                |  | Hole 722A depth<br>(mbsf) | Hole 722B depth<br>(mbsf) |
|----------------------|--|---------------------------|---------------------------|
| Radiolarians (cont.) |  |                           |                           |
| R27                  | T <i>Solenosphaera omnitubus</i>             | 144.60–154.30             |                           |
| R28                  | T <i>Botryostrobus bramletti</i>             | 166.25–173.60             |                           |
| R29                  | T <i>Siphostichoartus corona</i>             | 178.95–183.30             |                           |
| R30                  | T <i>Stichocorys delmontensis</i>            | 183.30–188.65             |                           |
| R33                  | T <i>Stichocorys johnsonii</i>               | 193.00–202.70             |                           |
| R31                  | T <i>Acrobotrys tritubus</i>                 | 208.05–212.40             |                           |
| R36                  | B <i>Spondiscus ambus</i>                    | 208.05–212.40             |                           |
| R35                  | <i>S. delmontensis</i> → <i>S. peregrina</i> | 212.40–217.75             |                           |
| R37                  | T <i>Calocyclus caepa</i>                    | 227.35–231.70             |                           |
| R38                  | T <i>Phormostichoartus marylandicus</i>      | 231.70–241.70             |                           |
| R39                  | T <i>Dendrospyrus bursa</i>                  | 231.70–241.70             |                           |
| R40                  | B <i>Solenosphaera omnitubus</i>             | 241.70–251.00             |                           |
| R41                  | B <i>Stichocorys peregrina</i>               | 241.70–251.00             |                           |
| R42                  | T <i>Diartus hughesi</i>                     | 241.70–251.00             |                           |
| R34                  | T <i>Didymocorytis antepenultima</i>         | 256.30–260.70             |                           |
| R45                  | T <i>Dictyocoryne ontongensis</i>            | 270.40–280.00             |                           |
| R46                  | T <i>Didymocorytis laticonus</i>             | 270.40–280.00             |                           |
| R49                  | T <i>Botryostrobus miralestensis</i>         | 270.40–280.00             |                           |
| R43                  | B <i>Acrobotrys tritubus</i>                 |                           | 271.25–275.16             |
| R50                  | T <i>Diartus petterssoni</i>                 |                           | 275.16–277.71             |
| R51                  | <i>D. petterssoni</i> → <i>D. hughesi</i>    |                           | 277.71–301.50             |
| R48                  | T <i>Stichocorys wolffii</i>                 |                           | 294.36–300.35             |
| R53                  | B <i>Botryostrobus bramletti</i>             |                           | 300.35–301.50             |
| R47                  | B <i>Spongaster berminghami</i>              |                           | 301.50–309.29             |
| R52                  | B <i>Diartus hughesi</i>                     |                           | 301.50–309.29             |
| R54                  | B <i>Dictyocoryne ontongensis</i>            |                           | 301.50–309.29             |
| R55                  | T <i>Cyrtocapsella japonica</i>              |                           | 329.35–333.80             |
| R57                  | B <i>Phormostichoartus doliolum</i>          |                           | 339.05–339.31             |
| R58                  | T <i>Lithopera thornburgi</i>                |                           | 348.75–352.93             |
| R59                  | T <i>Cyrtocapsella cornuta</i>               |                           | 352.75–352.93             |
| Polarity reversals   |  |                           |                           |
| M3                   | B Jaramillo                                  | 37.65–38.40               | 39.80–40.60               |
| M6                   | Matuyama/Gauss                               | 82.45–83.25               | 83.80–85.20               |
| M9                   | Gauss/Gilbert                                | 103.95–105.35             | 104.00–105.58             |
| M14                  | T Sidufjall                                  | 142.05–142.85             |                           |
| M17                  | B Thvera                                     | 151.00–153.30             | 152.40–155.40             |
| M18                  | Gilbert/Chronozone 5                         | 180.73–181.55             | 181.39–189.60             |
| M19                  | Chronozone 5/6                               | 214.34–220.37             | 218.50–221.50             |

two-thirds the magnitude of late Pleistocene glacial maxima (Shackleton and Opdyke, 1977). According to Perch-Nielsen (1985), the genus *Helicosphaera* seems to be more abundant in hemipelagic sediments and may prefer areas of upwelling. This may account for the large numbers of this genus found in the Pliocene and Pleistocene sediments at some of the sites (723, 724, 725, and 728).

A graph of the species diversity of the calcareous nannofossils for Leg 117 when compared to a typical tropical region (Shipboard Scientific Party, 1989, p. 603) shows that the nannofloras in the Arabian Sea may have first been affected by the upwelling waters during the early late Miocene (approximately 9.5 Ma).

An examination of the living planktonic foraminifers present in the surface waters of the Arabian and Red Seas during the summer of 1984 and 1985 (Kroon, 1988; Auras-Schudnagies et al., in press) indicates that the "upwelling assemblages" are dominated by *Globigerina bulloides* and *Neogloboquadrina dutertrei*. Other species present in lesser quantities include *Globorotalia menardii* and *Pulleniatina obliquiloculata*.

The presence of the cold-water benthic foraminifer, *Hyalinea balthica*, in the Pliocene and Pleistocene sediments of Sites 723 and 724 may be correlative with the increased numbers of the nannofossil, *Coccolithus pelagicus*, at the same time intervals at the same sites. The above discussion about the late Pliocene to early Pleistocene glaciations may well be applicable to the presence of *H. balthica* in the sediments as well. It cannot be determined to what extent the combination of the glaciation with the

upwelling in the Arabian Sea may have affected the presence of cold-water taxa.

The comparison of the radiolarian assemblages from the Peru Margin (ODP Leg 112) with those from this leg indicate that a specific assemblage may be unique to upwelling areas. Three categories of radiolarians can be recognized in upwelling assemblages. These are:

1. Taxa endemic to upwelling,
2. Taxa abundant in middle latitudes, but also present in areas of upwelling, and
3. Tropical taxa which are more robust and/or more abundant in regions of upwelling.

During the late early Miocene to early middle Miocene, the Owen Ridge was uplifted to a sufficient height as to be above the realm of turbidite deposition. Monsoonal-induced upwelling appears to have been initiated during the early late Miocene as reflected by increased amounts of biogenic silica preserved in the sediments. Sudden increases in the abundances of the planktonic foraminifers, *Globigerina bulloides* and *Globorotalia menardii*, and the appearance of the *Neogloboquadrina acostaensis-N. humerosa-N. dutertrei* group occur between Zones N15 and N16 at Site 721. This change in the fauna may represent the initiation of the monsoonal-related intensity in the earliest late Miocene. According to Bé (1977), the modern representatives of these taxa are indicative of upwelling. The upwelling is probably also responsible for the comparative scarcity of warm-water Pli-

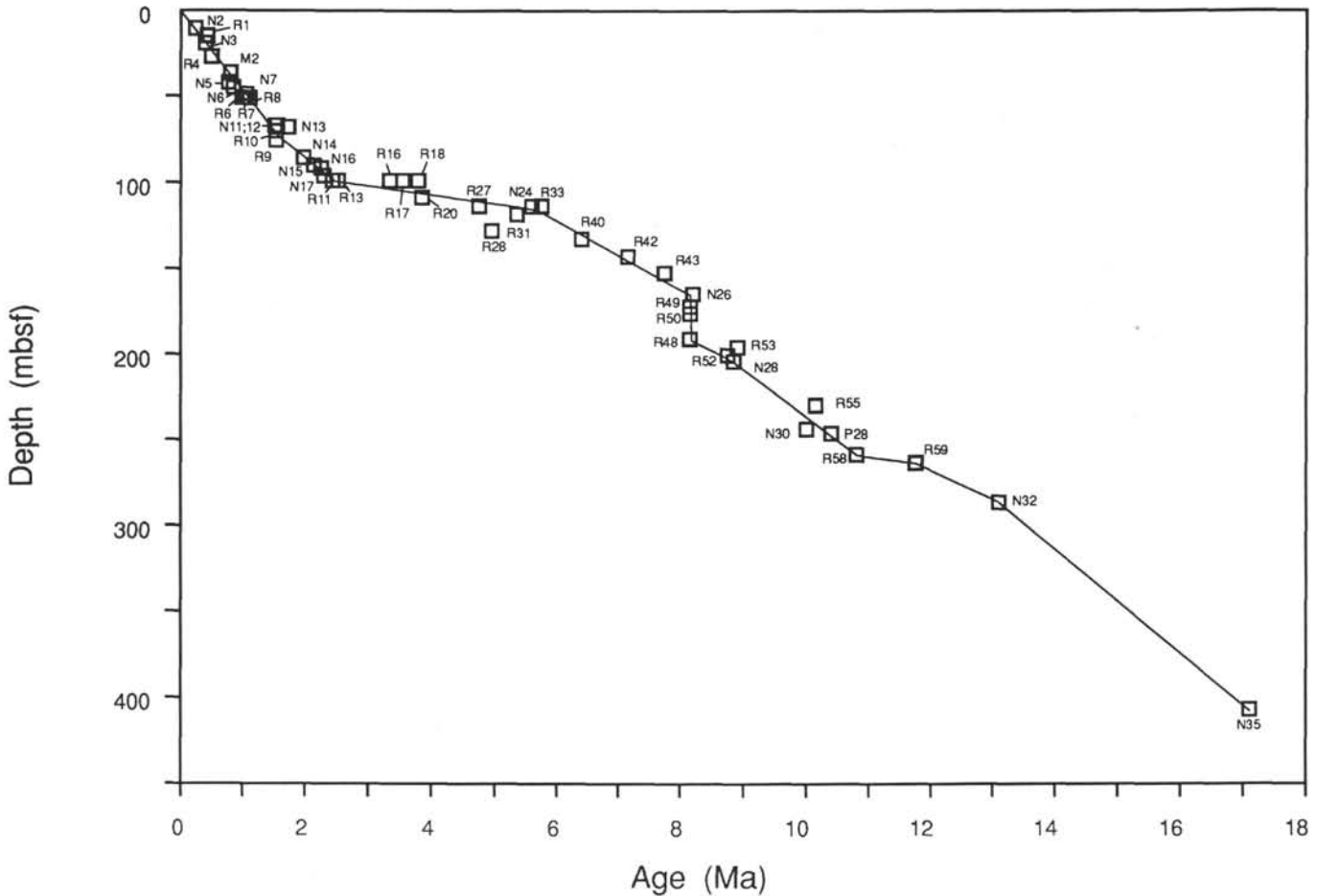


Figure 6. Sedimentation rate curve for Site 731.

ocene calcareous nannofossils at Sites 721 and 722. Upper Miocene and Pliocene nannofloras are less diverse at Site 731 than the same age assemblages from Sites 721 and 722. Also, at all three Owen Ridge sites, *Coccolithus pelagicus*, a typical cold-water nannofossil is dominant in the upper Pliocene and lower Pleistocene sediments.

Sites 726 and 729 were located in basins that were probably less than 130 m deep during the Eocene. Unfortunately, the Oligocene history of the Oman Margin remains unknown. The first occurrence of siliceous microfossils, presumably the result of the initiation of monsoonal-induced upwelling, in the middle Miocene sediments on the Oman Margin is earlier than on the Owen Ridge.

The presence (or absence) of certain microfossils in the Neogene and Quaternary sediments on the Oman Margin reflects the continued influence of the upwelling on the Neogene and Quaternary microfossil assemblages in the western Arabian Sea. These include the presence of both *Reticulofenestra pseudumbilica* and *R. gelida* in sediments from calcareous nannofossil Zones NN11 through NN15. Common biogenic opal in upper Miocene sediments at Site 728 also are indicators of increased paleoproductivity resulting from the upwelling of nutrient-enriched waters.

Planktonic foraminifers indicative of upwelling, *Hyalinea balthica*, a cold-water benthic foraminifer, and *Coccolithus pelagicus* and *Helicosphaera* spp., and biogenic opal are all common components of Pliocene sediments. *Coccolithus pelagicus* continued to be abundant into the early Pleistocene.

Radiolarians which are characteristic of the Arabian Upwelling Assemblage, the benthic foraminifer *Hyalinea balthica*, and planktonic foraminifers indicative of upwelling also occur in Pleistocene sediments on the Oman Margin.

Sites 724 and 728 have undergone subsidence of 250 m and 1000 m, respectively, since the early Pliocene. Site 725 has been uplifted during the latest Pleistocene to Holocene. The high P/B ratio and the uniform benthic foraminiferal assemblages indicate that there have been no significant changes in the water depth at Site 727 during the Neogene or Quaternary.

An interesting transition in the upwelling regime took place between the Pliocene and the Pleistocene. While the Miocene and Pliocene sediments are dominated by the siliceous component, the Pleistocene sediments seem to be dominated by the calcareous component. Sites 721 and 731 are the only ones which contain large amounts of siliceous microfossils in the Pleistocene deposits.

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**Table 5. Biostratigraphic and paleomagnetic events at Site 731.**

| Event                   |  | Hole 731A depth (mbsf) |
|-------------------------|--|------------------------|
| Calcareous nannofossils |  |                        |
| N2                      | T <i>Emiliania huxleyi</i>                   | 9.80–10.95             |
| N3                      | T <i>Pseudoemiliania lacunosa</i>            | 18.45–19.30            |
| N5                      | T <i>Reticulofenestra asanoi</i>             | 40.95–42.45            |
| N6                      | B <i>Gephyrocapsa parallela</i>              | 43.95–45.45            |
| N7                      | B acme <i>Reticulofenestra asanoi</i>        | 47.80–48.95            |
| N8                      | T <i>Gephyrocapsa (large)</i>                | 51.95–53.45            |
| N11                     | T <i>Calcidiscus macintyreii</i>             | 65.95–67.00            |
| N12                     | B <i>Gephyrocapsa oceanica</i>               | 65.95–67.00            |
| N13                     | B <i>Gephyrocapsa caribbeanica</i>           | 163.70–165.15          |
| N14                     | T <i>Discoaster brouweri</i>                 | 83.85–86.40            |
| N15                     | B acme <i>Gephyrocapsa</i>                   | 89.05–90.55            |
| N16                     | T acme <i>Crenolithus doronicoides</i>       | 90.55–92.05            |
| N17                     | T <i>Discoaster pentaradiatus</i>            | 95.05–96.00            |
| N24                     | T <i>Discoaster quinqueramus</i>             | 112.85–114.35          |
| N26                     | B <i>Discoaster quinqueramus</i>             | 163.70–165.05          |
| N28                     | T <i>Discoaster hamatus</i>                  | 202.50–205.11          |
| N30                     | B <i>Discoaster hamatus</i>                  | 241.30–245.47          |
| N31                     | B <i>Catinaster coalitus</i>                 | 256.10–260.60          |
| N32                     | B <i>Discoaster kugleri</i>                  | 282.55–289.60          |
| N33                     | B <i>Cyclicargolithus floridanus</i>         | 289.60–291.42          |
| N35                     | T <i>Sphenolithus belemnus</i>               | 404.95–409.00          |
| Planktonic foraminifer  |  |                        |
| P28                     | T <i>Globorotalia mayeri</i>                 | 241.30–250.90          |
| Radiolarians            |  |                        |
| R1                      | T <i>Stylatractus universus</i>              | 9.80–19.30             |
| R3                      | B <i>Collosphaera tuberosa</i>               | 24.65–28.80            |
| R4                      | T <i>Pterocorys camapanula</i>               | 28.80–34.15            |
| R5                      | B <i>Pterocorys hertwigii</i>                | 34.15–38.30            |
| R6                      | T <i>Anthocyrtidium angulare</i>             | 47.80–53.15            |
| R7                      | B <i>Lamprocyrtis nigrinae</i>               | 47.80–53.15            |
| R8                      | T <i>Lamprocyrtis neoheteroporos</i>         | 47.80–53.15            |
| R10                     | B <i>Anthocyrtidium angulare</i>             | 67.00–72.35            |
| R12                     | B <i>Theocorythium trachelium</i>            | 67.00–72.35            |
| R9                      | T <i>Pterocanium prismatum</i>               | 72.35–76.70            |
| R15                     | T <i>Phormostichoartus fistula</i>           | 82.05–86.40            |
| R11                     | B <i>Cycladophora davisiana</i>              | 96.00–101.35           |
| R13                     | B <i>Lamprocyrtis neoheteroporos</i>         | 96.00–101.35           |
| R14                     | T <i>Stichocorys peregrina</i>               | 96.00–101.35           |
| R16                     | T <i>Lychnodictyum audax</i>                 | 96.00–101.35           |
| R17                     | T <i>Phormostichoartus doliolum</i>          | 96.00–101.35           |
| R18                     | B <i>Amphirhopalum ypsilon</i>               | 96.00–101.35           |
| R24                     | T <i>Spongodiscus ambus</i>                  | 101.35–105.70          |
| R20                     | B <i>Spongaster tetras tetras</i>            | 105.70–111.05          |
| R21                     | T <i>Spongaster berminghamsi</i>             | 105.70–111.05          |
| R26                     | T <i>Didymocyrtis penultima</i>              | 105.70–111.05          |
| R23                     | T <i>Dictyophimus splendens</i>              | 111.05–115.40          |
| R27                     | T <i>Solenosphaera omnitubus</i>             | 111.05–115.40          |
| R29                     | T <i>Siphostichartus corona</i>              | 111.05–115.40          |
| R30                     | T <i>Stichocorys delmontensis</i>            | 111.05–115.40          |
| R33                     | T <i>Stichocorys johnsoni</i>                | 111.05–115.40          |
| R31                     | T <i>Acrobotrys tritubus</i>                 | 115.40–120.70          |
| R36                     | B <i>Spongodiscus ambus</i>                  | 115.40–120.70          |
| R35                     | <i>S. delmontensis</i> → <i>S. peregrina</i> | 115.40–125.00          |
| R37                     | T <i>Calocyrtella caepa</i>                  | 120.70–125.00          |
| R28                     | T <i>Botryostrobus bramlettei</i>            | 125.00–130.35          |
| R41                     | B <i>Stichocorys peregrina</i>               | 125.00–130.35          |
| R39                     | T <i>Dendrospyris bursa</i>                  | 125.00–130.35          |
| R34                     | T <i>Didymocyrtis antepenultima</i>          | 125.00–134.70          |
| R40                     | B <i>Solenosphaera omnitubus</i>             | 130.35–134.70          |
| R42                     | T <i>Diartus hughesi</i>                     | 140.95–144.40          |
| R43                     | B <i>Acrobotrys tritubus</i>                 | 149.75–154.00          |
| R45                     | T <i>Dictyocoryne ontongensis</i>            | 149.75–154.00          |
| R49                     | T <i>Botryostrobus miralestensis</i>         | 169.05–173.40          |
| R50                     | T <i>Diartus petterssoni</i>                 | 173.40–178.75          |
| R46                     | T <i>Didymocyrtis laticonus</i>              | 178.75–183.10          |
| R51                     | <i>D. petterssoni</i> → <i>D. hughesi</i>    | 178.75–198.15          |
| R48                     | T <i>Stichocorys wolffii</i>                 | 188.45–192.80          |
| R53                     | B <i>Botryostrobus bramlettei</i>            | 192.80–198.15          |
| R52                     | B <i>Diartus hughesi</i>                     | 198.15–202.50          |
| R54                     | B <i>Dictyocoryne ontongensis</i>            | 198.15–202.50          |
| R55                     | T <i>Cyrtocapsella japonica</i>              | 227.25–231.60          |
| R58                     | T <i>Lithopera thornburgi</i>                | 256.27–260.60          |
| R59                     | T <i>Cyrtocapsella cornuta</i>               | 260.60–265.96          |
| Polarity reversals      |  |                        |
| M1                      | Brunhes/Matuyama(?)                          | 35.25–36.75            |
| M2                      | T Jaramillo(?)                               | 38.23–38.75            |
| M3                      | B Jaramillo                                  | 45.45–46.25            |

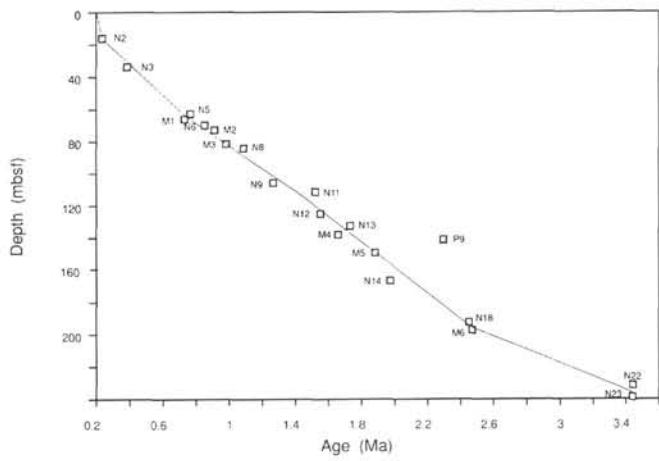
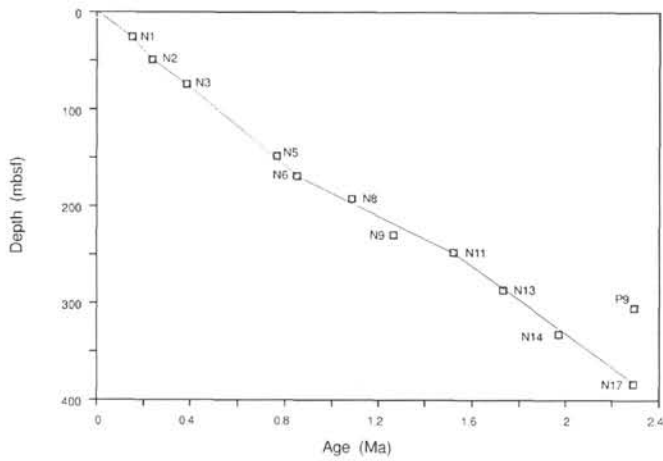


Figure 7. Sedimentation rate curve for Site 723.

Figure 8. Sedimentation rate curve for Site 724.

Table 6. Biostratigraphic events at Site 723.

| Event                                  | Hole 723A depth (mbsf) |
|--|------------------------|
| Calcareous nannofossils                |                        |
| N1 T <i>Helicosphaera inversa</i>      | 24.70–27.10            |
| N2 B <i>Emiliana huxleyi</i>           | 47.60–50.60            |
| N3 T <i>Pseudoemiliana lacunosa</i>    | 73.00–75.40            |
| N5 T <i>Reticulofenestra asanoi</i>    | 143.10–153.70          |
| N6 B <i>Gephyrocapsa parallela</i>     | 162.40–173.30          |
| N8 T <i>Gephyrocapsa (large)</i>       | 191.40–192.74          |
| N9 T <i>Helicosphaera sellii</i>       | 227.58–231.27          |
| N11 T <i>Calcidiscus macintyreii</i>   | 245.87–249.40          |
| N13 B <i>Gephyrocapsa caribbeanica</i> | 285.19–287.90          |
| N14 T <i>Discoaster brouweri</i>       | 330.36–333.53          |
| N17 T <i>Discoaster pentaradiatus</i>  | 381.50–384.68          |
| Planktonic foraminifers                |                        |
| P9 T <i>Globigerinoides obliquus</i>   | 302.65–307.10          |

Table 7. Biostratigraphic and paleomagnetic events at Site 724.

| Event  | Hole 724A depth (mbsf) | Hole 724B depth (mbsf) |
|--|------------------------|------------------------|
| Calcareous nannofossils                      |                        |                        |
| N2 B <i>Emiliana huxleyi</i>                 | 15.90–16.95            |                        |
| N3 T <i>Pseudoemiliana lacunosa</i>          | 32.45–34.90            |                        |
| N5 T <i>Reticulofenestra asanoi</i>          |                        | 60.25–65.50            |
| N6 B <i>Gephyrocapsa parallela</i>           |                        | 68.50–71.40            |
| N8 T <i>Gephyrocapsa (large)</i>             |                        | 83.70–84.75            |
| N9 T <i>Helicosphaera sellii</i>             |                        | 104.35–107.35          |
| N11 T <i>Calcidiscus macintyreii</i>         |                        | 110.00–112.70          |
| N12 B <i>Gephyrocapsa oceanica</i>           |                        | 123.50–126.44          |
| N13 B <i>Gephyrocapsa caribbeanica</i>       |                        | 132.00–133.03          |
| N14 T <i>Discoaster brouweri</i>             |                        | 165.21–168.05          |
| N18 T <i>Discoaster surculus</i>             |                        | 191.05–194.05          |
| N22 T <i>Sphenolithus abies</i>              |                        | 230.05–233.05          |
| N23 T <i>Reticulofenestra pseudoumbilica</i> |                        | 238.40–239.44          |
| Planktonic foraminifer                       |                        |                        |
| P9 T <i>Globigerinoides obliquus</i>         |                        | 146.85–151.40          |
| Polarity reversals                           |                        |                        |
| M1 Brunhes/Matuyama                          |                        | 65.45–66.95            |
| M2 T Jaramillo                               |                        | 71.34–74.44            |
| M3 B Jaramillo                               |                        | 81.14–81.93            |
| M4 T Olduvai                                 |                        | 134.18–141.89          |
| M5 B Olduvai                                 |                        | 148.86–149.70          |
| M6 Matuyama/Gauss                            |                        | 194.23–200.36          |

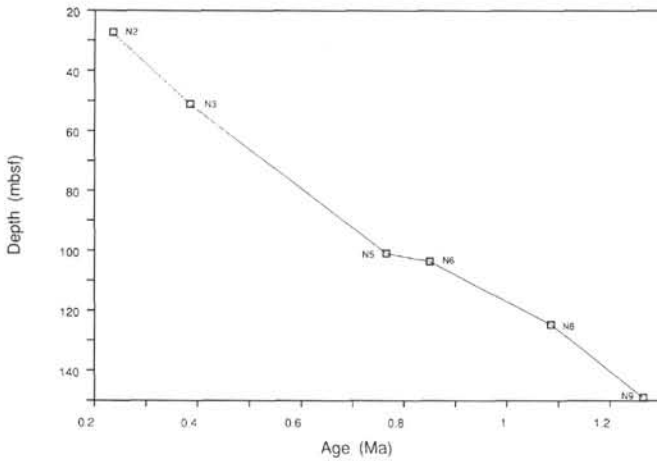


Figure 9. Sedimentation rate curve for Site 725.

Table 8. Biostratigraphic events at Site 725.

| Event                               | Hole 725C depth (mbsf) |
|-------------------------------------|------------------------|
| Calcareous nannofossils             |                        |
| N2 B <i>Emiliana huxleyi</i>        | 25.78–28.50            |
| N3 T <i>Pseudoemiliana lacunosa</i> | 50.18–51.68            |
| N5 T <i>Reticulofenestra asanoi</i> | 99.48–102.23           |
| N6 B <i>Gephyrocapsa parallela</i>  | 102.23–104.90          |
| N8 T <i>Gephyrocapsa (large)</i>    | 124.10–125.28          |
| N9 T <i>Helicosphaera sellii</i>    | 144.89–153.20          |

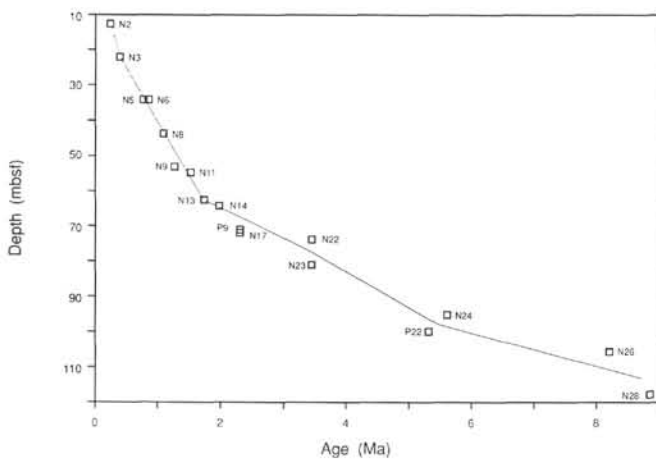


Figure 10. Sedimentation rate curve for Site 726.

Table 9. Biostratigraphic events at Site 726.

| Event                                       | Hole 726A depth (mbsf) |
|---|------------------------|
| Calcareous nannofossils                     |                        |
| N2 B <i>Emiliana huxleyi</i>                | 11.08–14.08            |
| N3 T <i>Pseudoemiliana lacunosa</i>         | 20.48–23.48            |
| N5 T <i>Reticulofenestra asanoi</i>         | 32.98–35.30            |
| N6 B <i>Gephyrocapsa parallela</i>          | 32.98–35.30            |
| N8 T <i>Gephyrocapsa (large)</i>            | 42.48–44.70            |
| N9 T <i>Helicosphaera sellii</i>            | 51.88–54.10            |
| N11 T <i>Calcidiscus macintyreii</i>        | 54.10–55.28            |
| N13 B <i>Gephyrocapsa caribbeanica</i>      | 61.28–63.60            |
| N14 T <i>Discoaster brouweri</i>            | 63.60–64.78            |
| N17 T <i>Discoaster pentaradiatus</i>       | 70.63–73.20            |
| N22 T <i>Sphenolithus abies</i>             | 73.20–74.38            |
| N23 T <i>Reticulofenestra pseudombilica</i> | 77.85–83.98            |
| N24 T <i>Discoaster quinqueramus</i>        | 93.58–96.58            |
| N25 T <i>Discoaster berggrenii</i>          | 93.58–96.58            |
| N26 B <i>Discoaster quinqueramus</i>        | 98.10–113.24           |
| N27 B <i>Discoaster berggrenii</i>          | 98.10–113.24           |
| N28 T <i>Discoaster hamatus</i>             | 116.24–119.24          |
| Planktonic foraminifers                     |                        |
| P9 T <i>Globigerinoides obliquus</i>        | 68.75–73.20            |
| P22 B <i>Globorotalia tumida tumida</i>     | 97.55–102.10           |

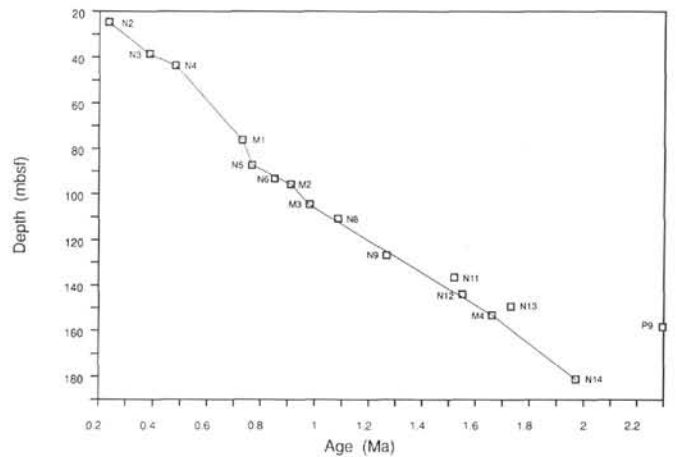


Figure 11. Sedimentation rate curve for Site 727.



**Table 10. Biostratigraphic and paleomagnetic events at Site 727.**

| Event                                  | Hole 724A depth (mbsf) |
|--|------------------------|
| <b>Calcareous nannofossils</b>         |                        |
| N2 B <i>Emiliana huxleyi</i>           | 23.08–26.08            |
| N3 T <i>Pseudoemiliana lacunosa</i>    | 37.80–38.98            |
| N4 B <i>Helicosphaera inversa</i>      | 41.98–44.98            |
| N5 T <i>Reticulofenestra asanoi</i>    | 85.80–88.70            |
| N6 B <i>Gephyrocapsa parallela</i>     | 91.70–94.70            |
| N8 T <i>Gephyrocapsa</i> (large)       | 109.18–112.15          |
| N9 T <i>Helicosphaera sellii</i>       | 125.18–128.14          |
| N11 T <i>Calcidiscus macintyreii</i>   | 134.96–138.06          |
| N12 B <i>Gephyrocapsa oceanica</i>     | 143.06–144.78          |
| N13 B <i>Gephyrocapsa caribbeanica</i> | 147.78–150.74          |
| N14 T <i>Discoaster brouweri</i>       | 179.90–182.40          |
| <b>Planktonic foraminifer</b>          |                        |
| P9 T <i>Globigerinoides obliquus</i>   | 153.30–163.00          |
| <b>Polarity reversals</b>              |                        |
| M1 Brunhes/Matuyama                    | 75.80–76.50            |
| M2 T Jaramillo                         | 95.40–95.83            |
| M3 B Jaramillo                         | 104.14–104.69          |
| M4 T Olduvai                           | 152.72–153.55          |

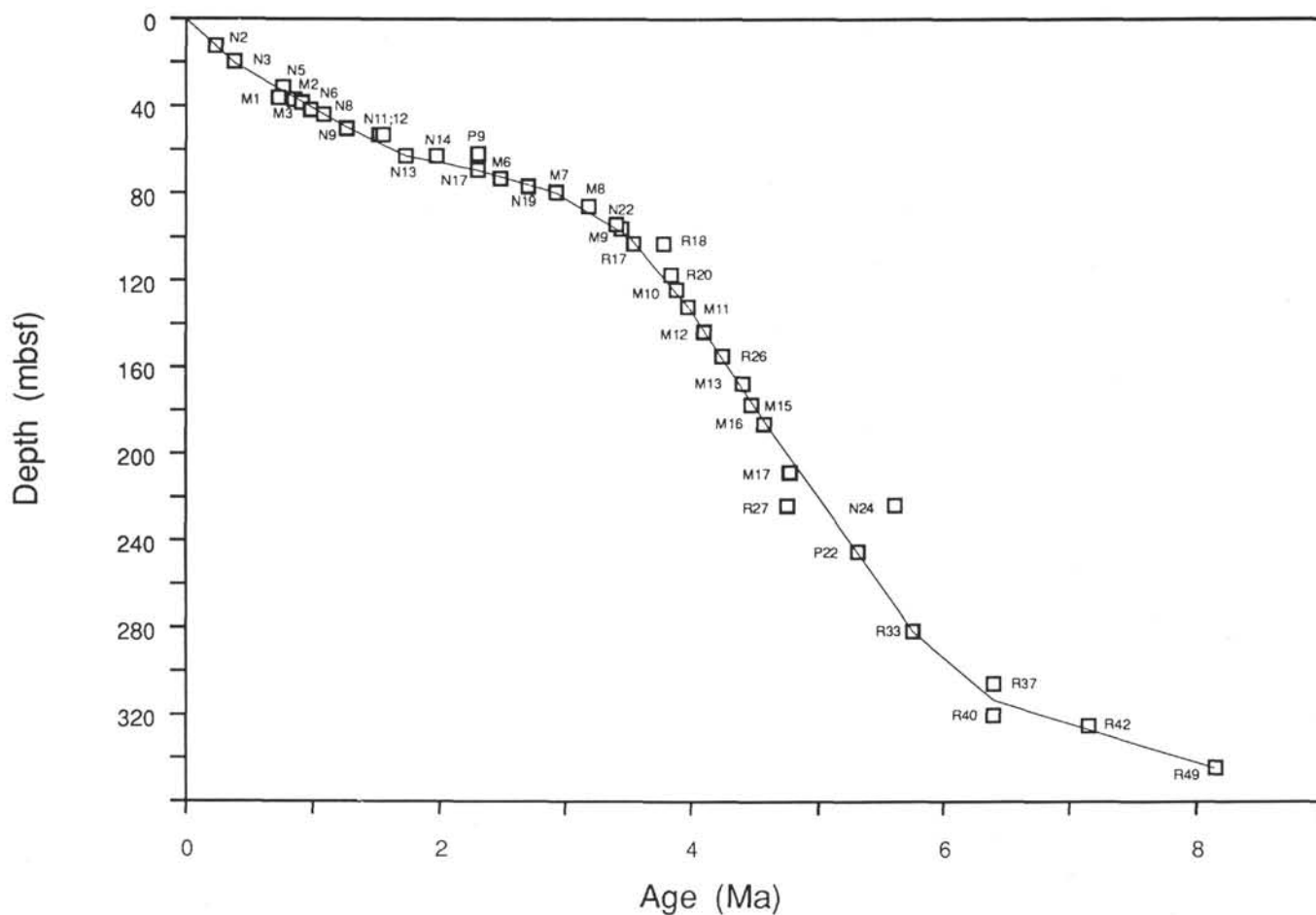


Figure 12. Sedimentation rate curve for Site 728.

**Table 11. Biostratigraphic and paleomagnetic events at Site 728.**

| Event                          |  | Hole 728A depth (mbsf) |
|--------------------------------|--|------------------------|
| <b>Calcareous nannofossils</b> |  |                        |
| N2                             | B <i>Emiliana huxleyi</i>                    | 10.78–13.78            |
| N3                             | T <i>Pseudoemiliana lacunosa</i>             | 19.10–20.28            |
| N5                             | T <i>Reticulofenestra asanoi</i>             | 29.68–32.68            |
| N6                             | B <i>Gephyrocapsa parallela</i>              | 35.68–38.00            |
| N8                             | T <i>Gephyrocapsa</i> (large)                | 42.18–45.18            |
| N9                             | T <i>Helicosphaera sellii</i>                | 48.68–51.68            |
| N11                            | T <i>Calcidiscus macintyreii</i>             | 51.68–54.68            |
| N12                            | B <i>Gephyrocapsa oceanica</i>               | 51.68–54.68            |
| N13                            | B <i>Gephyrocapsa caribbeanica</i>           | 61.25–64.25            |
| N14                            | T <i>Discoaster brouweri</i>                 | 61.25–64.25            |
| N17                            | T <i>Discoaster pentaradiatus</i>            | 67.68–70.68            |
| N19                            | T <i>Discoaster tamalis</i>                  | 76.10–77.28            |
| N22                            | T <i>Sphenolithus abies</i>                  | 95.30–96.48            |
| N23                            | T <i>Reticulofenestra pseudoumbilica</i>     | 95.30–96.48            |
| N24                            | T <i>Discoaster quinqueramus</i>             | 222.01–225.00          |
| <b>Planktonic foraminifers</b> |  |                        |
| P9                             | T <i>Globigerinoides obliquus</i>            | 57.00–66.50            |
| P22                            | B <i>Globorotalia tumida tumida</i>          | 240.10–249.80          |
| <b>Radiolarians</b>            |  |                        |
| R17                            | T <i>Phormostichoartus doliolum</i>          | 100.65–104.90          |
| R18                            | B <i>Amphirhopalum ypsilon</i>               | 100.65–104.90          |
| R20                            | B <i>Spongaster tetras tetras</i>            | 114.50–119.85          |
| R24                            | T <i>Spongodiscus ambus</i>                  | 139.25–143.50          |
| R26                            | T <i>Didymocyrtis penultima</i>              | 148.85–153.10          |
| R27                            | T <i>Solenosphaera omnitubus</i>             | 220.80–226.15          |
| R29                            | T <i>Siphostichartus corona</i>              | 245.45–249.80          |
| R30                            | T <i>Stichocorys delmontensis</i>            | 245.45–249.80          |
| R33                            | T <i>Stichocorys johnsoni</i>                | 278.80–284.15          |
| R36                            | B <i>Spongodiscus ambus</i>                  | 288.50–293.82          |
| R35                            | <i>S. delmontensis</i> → <i>S. peregrina</i> | 298.10–307.80          |
| R37                            | T <i>Calocyrella caepa</i>                   | 303.45–307.80          |
| R41                            | B <i>Stichocorys peregrina</i>               | 313.15–317.50          |
| R40                            | B <i>Solenosphaera omnitubus</i>             | 317.50–322.85          |
| R42                            | T <i>Diartus hughesi</i>                     | 322.85–327.20          |
| R46                            | T <i>Didymocyrtis laticonus</i>              | 342.15–346.40          |
| R49                            | T <i>Botryocyrtis miralestensis</i>          | 342.15–346.60          |
| <b>Polarity reversals</b>      |  |                        |
| M1                             | Brunhes/Matuyama                             | 35.70–36.45            |
| M2                             | T Jaramillo                                  | 37.95–38.45            |
| M3                             | B Jaramillo                                  | 40.72–42.20            |
| M6                             | Matuyama/Gauss                               | 72.20–73.70            |
| M7                             | T Kaena                                      | 78.80–79.55            |
| M8                             | B Mammoth                                    | 84.80–86.15            |
| M9                             | Gauss/Gilbert                                | 92.90–93.65            |
| M10                            | T Cochiti                                    | 122.45–125.35          |
| M11                            | B Cochiti                                    | 131.40–132.15          |
| M12                            | T Nunivak                                    | 142.60–144.10          |
| M13                            | B Nunivak                                    | 153.63–155.80          |
| M14                            | T Sidufjall                                  | 167.08–167.79          |
| M15                            | B Sidufjall                                  | 176.75–177.49          |
| M16                            | T Thvera                                     | 185.78–186.32          |
| M17                            | B Thvera                                     | 207.84–208.70          |

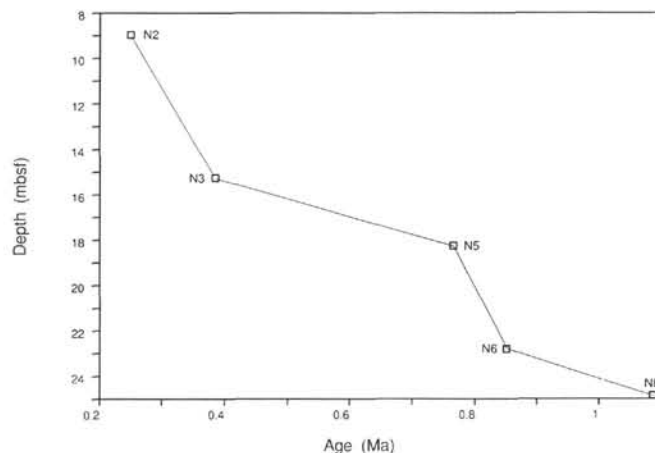


Figure 13. Sedimentation rate curve for Site 729.

**Table 12. Biostratigraphic events at Site 729.**

| Event                          |                                  | Hole 729A depth (mbsf) |
|--------------------------------|----------------------------------|------------------------|
| <b>Calcareous nannofossils</b> |                                  |                        |
| N2                             | B <i>Emiliana huxleyi</i>        | 7.25–10.25             |
| N3                             | T <i>Pseudoemiliana lacunosa</i> | 13.75–16.75            |
| N5                             | T <i>Reticulofenestra asanoi</i> | 16.75–19.75            |
| N6                             | B <i>Gephyrocapsa parallela</i>  | 22.30–23.35            |
| N8                             | T <i>Gephyrocapsa</i> (large)    | 23.35–26.35            |

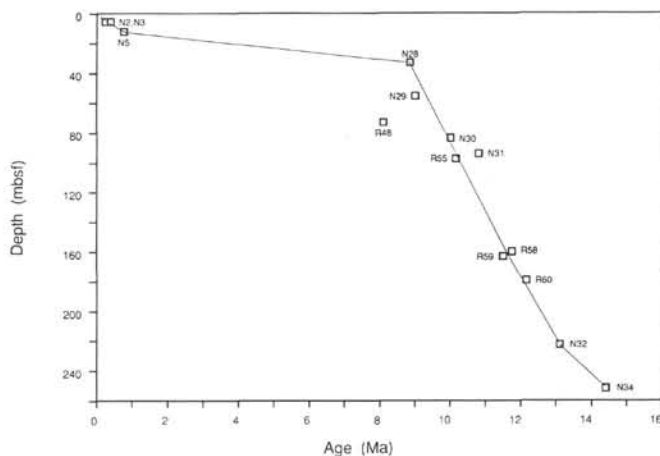


Figure 14. Sedimentation rate curve for Site 730.

Table 13. Biostratigraphic events at Site 730.

| Event                   |                                      | Hole 730A depth<br>(mbsf) |
|-------------------------|--------------------------------------|---------------------------|
| Calcareous nannofossils |                                      |                           |
| N2                      | B <i>Emiliana huxleyi</i>            | 4.05-7.05                 |
| N3                      | T <i>Pseudoemiliana lacunosa</i>     | 4.05-7.05                 |
| N5                      | T <i>Reticulofenestra asanoi</i>     | 11.91-12.31               |
| N28                     | T <i>Discoaster hamatus</i>          | 31.35-34.35               |
| N29                     | T <i>Catinaster coalitus</i>         | 53.35-56.85               |
| N30                     | B <i>Discoaster hamatus</i>          | 82.25-84.90               |
| N31                     | B <i>Catinaster coalitus</i>         | 91.95-95.65               |
| N32                     | B <i>Discoaster kugleri</i>          | 221.25-224.25             |
| N33                     | T <i>Cyclicargolithus floridanus</i> | 221.25-224.25             |
| N34                     | T <i>Sphenolithus heteromorphus</i>  | 250.25-253.25             |
| Planktonic foraminifers |                                      |                           |
| P30                     | T <i>Globorotalia fohsi</i>          | 162.20-164.35             |
| P32                     | B <i>Orbulina universa</i>           | 291.55-297.60             |
| Radiolarians            |                                      |                           |
| R48                     | T <i>Stichocorys wolffii</i>         | 70.85-75.20               |
| R55                     | T <i>Cyrtocapsella japonica</i>      | 94.60-99.95               |
| R58                     | T <i>Lithopera thornburgi</i>        | 152.60-157.95             |
| R59                     | T <i>Cyrtocapsella cornuta</i>       | 157.95-162.20             |
| R56                     | B <i>Diartus petterssoni</i>         | 167.55-171.90             |
| R60                     | T <i>Lithopera renzae</i>            | 177.25-181.60             |