

17. CALCAREOUS NANNOFOSSIL BIOSTRATIGRAPHY: SITES 840 (TONGA RIDGE) AND 841 (TONGA TRENCH)¹

Paula J. Quintero²

ABSTRACT

Ocean Drilling Program Leg 135 drilled at two sites on the Tonga Ridge. Calcareous nannofossils recovered at Site 840 on the Tonga Ridge date the sedimentary sequence as late Pleistocene or Holocene (CN15) through late Miocene (CN9) in age. A hiatus occurs in the mid Pliocene. Site 841 in the Tonga Trench yielded a sedimentary sequence with nannofossils from middle Pleistocene Subzone CN14b through the middle or late Eocene Subzones CP14a–CP15b overlying a rhyolitic volcanic basement. Part of the Eocene interval contains the shallow-water nannofossil taxa *Braarudosphaera*, *Micrantholithus*, and *Pemma*. A major unconformity separates lower Oligocene Zone CP16 from lower middle Miocene Zone CN4 strata.

INTRODUCTION

The purpose of Leg 135 was to study the geological evolution of the Lau backarc basin and the adjacent Tofua/Tonga oceanic arc system and to apply the findings to oceanic arc and backarc spreading systems in general. A total of eight sites were drilled in the Lau Basin and on the adjacent Tonga Ridge during Ocean Drilling Program (ODP) Leg 135 (Fig. 1). This study presents the calcareous nannofossil biostratigraphy of Sites 840 and 841, which are located on the Tonga Ridge and in the Tonga Trench, respectively. Table 1 shows the longitude, latitude, water depth, and meters penetrated in holes at Sites 840 and 841. For a discussion of calcareous nannofossils from the Lau backarc basin (Sites 834 through 839), see Styzen (this volume).

A primary objective at Site 840 was to determine the age of a seismic reflector, "Horizon A," thought to be a regional unconformity of latest Miocene and earliest Pliocene age; however, it could not be identified from our biostratigraphic results although the base of the section recovered is dated as being older than "Horizon A." See Scholl and Vallier (1985) and Shipboard Scientific Party (1992a) for a discussion of "Horizon A."

Site 841 was chosen to study the sedimentary section and basement for comparison with forearc sites previously drilled in the Mariana and Bonin arc systems (Shipboard Scientific Party, 1992b).

METHODS

Smear slides were prepared by mixing sediment with distilled water in a small vial; agitating with a "vortex" mixer; extracting an aliquot with a micropipette onto a glass slide; moving the micropipette back and forth over the slurry on the slide until most of the water had evaporated and ridges of sediment had formed; evaporating the remainder of the water on a hot plate; and covering the slide with a glass cover slip to which piccolyte had been added. The slides were examined with a transmitting light microscope at $\times 1000$ magnification using phase contrast and cross-polarized light.

Abundance and Preservation

Abundance and preservation data for Holes 840 and 841 are presented in Tables 2 through 6. Abundance estimates of the nannofossils

in each slide were made by counting a transect of 100 fields at $\times 1000$ and assigning the following abundance codes:

V (very abundant) = >10 specimens per field of view,
A (abundant) = 1–10 specimens per field of view,
C (common) = 1 specimen in 2–10 fields of view,
F (few) = 1 specimen in 11–100 fields of view,
R (rare) = 1 specimen in 101–1000 fields of view,
f = few reworked specimens, and
r = rare reworked specimens.

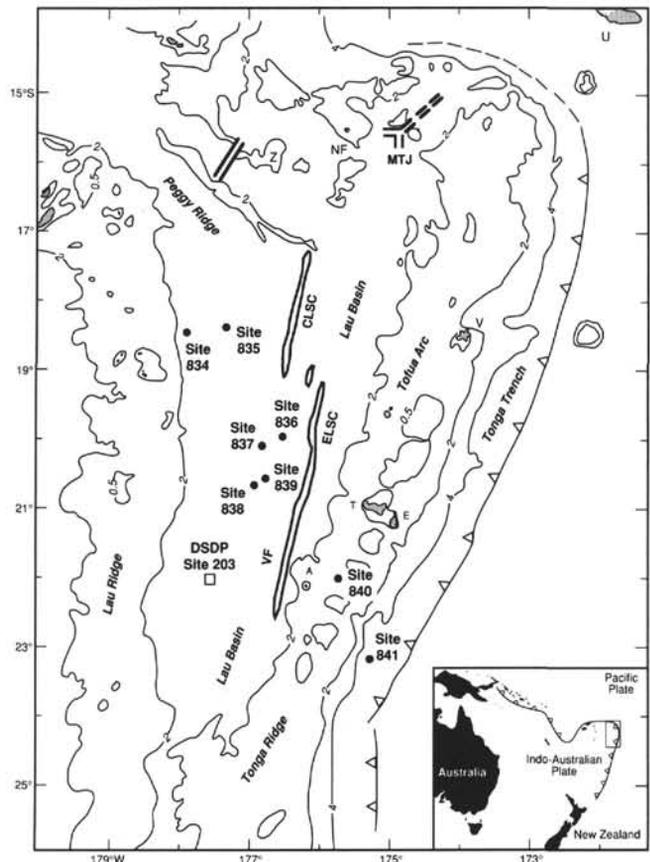


Figure 1. Location of Leg 135 sites. Islands include T = Tongatapu, E = 'Eua, V = Vava'u, A = 'Ata, NF = Niuafu'ou, and U = Upolu. Contour interval in kilometers.

¹ Hawkins, J., Parson, L., Allan, J., et al., 1994. *Proc. ODP, Sci. Results*, 135: College Station, TX (Ocean Drilling Program).

² U.S. Geological Survey, M.S. 999, 345 Middlefield Road, Menlo Park, CA 94025, U.S.A.

Table 1. Location, water depth, and penetration of Sites 840 and 841.

Hole	Latitude (°S)	Longitude (°W)	Water depth (m)	Penetration (m)
840A	22°13.249'	175°44.916'	743.3	4.5
840B	22°13.259'	175°44.918'	743.3	597.3
840C	22°13.234'	175°44.925'	743.3	259.5
841A	23°20.746'	175°17.871'	4809.8	186.6
841B	23°20.741'	175°17.872'	4809.0	834.2

The following codes were assigned to describe nannofossil preservation:

- G (good) = little or no dissolution and/or overgrowth of specimens;
 M (moderate) = moderate dissolution and/or overgrowth of specimens, identification of species somewhat impaired; and
 P (poor) = extreme dissolution and/or overgrowth of specimens.

BIOSTRATIGRAPHY

The calcareous nannofossil zonations (CP for the Paleogene and CN for the Neogene) of Bukry (1973, 1975, 1981) and Okada and Bukry (1980) are used. Perch-Nielsen (1985) summarizes the problems of subdividing the Pleistocene. She points out that small members of the family Prinsiacae are largely used, and these are best studied quantitatively and by transmission and scanning electron microscopy (TEM and SEM). Furthermore, the numerous studies (e.g., Pujos-Lamy, 1977a, 1977b) that have been done are difficult to compare and some subdivisions may be valid only locally. Recovery of Pleistocene sediment at both Sites 840 and 841 was poor; therefore, I have not attempted to do the necessary detailed quantitative and TEM or SEM work to subdivide this unit using the various species of *Gephyrocapsa*.

Site 840

Site 840 is located on the western Tonga Ridge approximately 130 km south-southwest of 'Eua and Tongatapu. Three holes were drilled at this site at a water depth of 743 m, but basement was not reached. The recovered sequence ranges in age from late Miocene (CN9) to late Pleistocene or Holocene CN15 (Table 7), and consists mainly of vitric volcanoclastic sediments with interbeds of calcareous oozes and chalks. Miocene volcanoclastic turbidites are common, and sediments coarsen downhole to volcanoclastic breccia and conglomerate.

Core recovery was good for Holes 840A (91.3%) and 840C (60.6%); however, they were only drilled to depths of 4.5 and 259.5 m, respectively. Recovery was poor (29.5%), especially in the upper 260 m of Hole 840B, which was drilled to 597.3 mbsf. No samples were recovered for Cores 135-840B-5X through -8X, and only core-catcher samples were recovered for several other cores throughout the hole. The lack of closely spaced, continuous samples, the presence of abundant volcanic glass, and the common occurrence of turbidites limit biostratigraphic resolution at this site. When present, nannofossils are abundant and of moderate preservation in most samples from Site 840 (Tables 2–4); however, small placoliths (<4 μm) often dominate the assemblages. These forms are difficult to identify with the light microscope at ×1000, especially when they are not well preserved. Therefore, for this study, they have been grouped as “small placoliths.” Barren samples and samples with only sparse, nondiagnostic species are not plotted in Tables 2 through 4 but are listed as footnotes to the tables.

Pleistocene and Pliocene

The first occurrence (FO) of *Emiliana huxleyi* defines the base of Zone CN15 and was identified by SEM in Sample 135-840A-1H-1, 45–46 cm. The base of Subzone CN14b is defined by the last occurrence (LO) of *Emiliana ovata* and the top by the FO of *E. huxleyi*. This zone was recognized in Holes 840A and 840B (Tables 2–3). For this study, I have kept the counts of *P. lacunosa* (circular morphotype) separate from *Emiliana ovata* (oval morphotype), but the ranges were the same (Tables 3–4) in the limited number of samples studied.

Only a core-catcher sample, consisting of several rock chips, was recovered in Core 135-840B-2X. One chip, Sample 135-840B-2X-CC (a), contains the overlap of *Gephyrocapsa oceanica* and *E. ovata* and is, therefore, assigned to Subzone CN14a. Another chip, Sample 135-840B-2X-CC (b), contains *Calcidiscus macintyreii*, *Ceratolithus rugosus*, *Discoaster brouweri*, *Gephyrocapsa caribbeanica*, and *Helicosphaera sellii*, and is assigned to Subzone CN13b. Although *D. brouweri* is present, it occurs with *G. caribbeanica*, and is considered reworked. Barred *gephyrocapsids* resembling *G. caribbeanica* occur in the core catcher below (135-840B-3X-CC) with discoasters including *D. brouweri*, *D. pentaradiatus*, and *D. surculus*, which are considered reworked, so the sample is also assigned to Subzone CN13b. The sample below (135-840B-4X-CC) contains abundant volcanic glass and few nannofossils. Discoasters are absent, and small barred *gephyrocapsids* are rare; it is assigned to Zone CN13.

The upper sediment was washed through in Hole 840C, and the first core was taken at 38 mbsf. Nannofossil preservation in Samples 135-840C-1H-2, 105–106 cm, and -1H-CC is moderate to poor (Table

Table 2. Abundance, preservation, and distribution of calcareous nannofossils, Hole 840A.

Zone or Subzone	Core, section, interval (cm)	Abundance		Preservation		<i>Calcidiscus leptoporus</i>		<i>Ceratolithus cristatus</i>		<i>Ceratolithus rugosus</i>		<i>Ceratolithus simplex</i>		<i>Discoaster</i> spp.		<i>Emiliana huxleyi</i>		<i>Gephyrocapsa caribbeanica</i>		<i>Gephyrocapsa oceanica</i>		<i>Hayaster perplexus</i>		<i>Helicosphaera carteri</i>		<i>Helicosphaera inversa</i>		<i>Helicosphaera wallichii</i>		<i>Oolithus fragilis</i>		<i>Pontosphaera indoceanica</i>		<i>Pontosphaera</i> spp.		<i>Rhabdosphaera</i> spp.		<i>Scyphosphaera</i> spp.		<i>Umbellosphaera irregularis</i>		Small <i>gephyrocapsids</i> (with bars)		Small placoliths	
		V	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G	F	G								
CN15	135-840A-1H-1, 45	V	G	F	F	.	F	V	F?	A	.	C	.	F	.	F	F	F	F	A	.	.	A	.	.	.	A					
CN14b	1H-2, 3	V	G	F	F	A	F	C	.	.	C	.	C	C	C	.	C	A	.	C	.	C	A					
	1H-2, 140	V	G	R	R	A	R	R	.	.	F	.	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R	R				
	1H-3, 32	V	G	R	.	r	A	A	.	C	.	.	C	.	C	C	C	C	A	F	.	A	.	A	.	A	.	A	.	A	.				
	1H-CC	V	G	A	C	.	C	.	.	C	.	C	C	C	F	.	C	.	C	.	C	.	A	.	A	.	A	.				

Notes: Abundance: V = very abundant, A = abundant, C = common, F = few, R = rare, and lowercase letters = reworked. Preservation: G = good, M = moderate, and P = poor.

4). The late Pliocene CN12 subzonal marker species, *D. pentaradiatus* and *D. brouweri*, are sparse, and *D. surculus* is absent. The sparseness and moderate to poor preservation of the discoasters makes age assignment difficult. However, Sample 135-840C-1H-2, 105–106 cm, and -1H-CC are assigned to Subzone CN12c on the basis of the presence of *D. brouweri* and *D. pentaradiatus* and lack of *D. surculus*.

Subzones CN12b–d could not be identified from the material recovered from Hole 840B. This is probably because of the lack of sediment recovery in Cores 135-840B-5X through -8X. Samples 135-840B-9X-CC and -10X-CC and 135-840C-2H-CC through -4H-CC are assigned to Subzone CN12a on the basis of the presence of *Discoaster tamalis* and the absence of *Reticulofenestra pseudoumbilica*. Bukry (1991) divided late Pliocene *D. tamalis* Subzone CN12a into three parts (A, B, and C) based on the extinctions of *Sphenolithus* spp. and *Discoaster variabilis* at tropical sites in the Pacific and Atlantic oceans. The three units contain assemblages typical of the *D. tamalis* subzone, but Subzone CN12aA must contain *S. spp.*; Subzone CN12aB must contain *D. variabilis* without *S. spp.*; and Subzone CN12aC must lack both *S. spp.* and *D. variabilis*. Using these criteria, Subzone CN12aA can be recognized in Core 840B-10X-CC. Subzone CN12aB can be recognized in Sample 135-840C-4H-CC. Subzone CN12aC can be recognized in Samples 135-840B-9X-CC and in Samples 135-840C-2H-CC through -4H-4, 69 cm (Tables 3–4).

Early Pliocene Zone CN11, characterized by the presence of *Reticulofenestra pseudoumbilica*, *Sphenolithus* spp., and a *D. asymmetricus* acme (in Subzone CN11b), but lacking *Amaurolithus* spp., could not be recognized in Holes 840B or 840C. In Hole 840C this interval was not cored, having been washed. In Hole 840B the absence of this zone is interpreted as a hiatus. However, this zone has been recognized in samples from the Tonga Platform by Bukry (1985). He reported nannofossils typical of this zone in two samples from Dredge 3 (22°46.3'S, 175°39.2'W) recovered at a water depth of 700–850 m.

Subzones CN10a through CN10d were recognized in Holes 840B and 840C in spite of low abundances of marker species and poor core recovery throughout much of this interval in Hole 840B. The absence of *Amaurolithus* species above Samples 135-840B-11X-1, 22–23 cm, and 135-840C-5H-1, 53–54 cm, marks the top of Zone CN10. The overlapping occurrence of *Amaurolithus* and *D. asymmetricus* in 135-840B-11X-1, 22–23 cm, through -11X-CC, and in Sample 135-840C-5H-1, 53 cm, indicates Subzone CN10d of Bukry (1981). *Ceratolithus rugosus*, the marker species for the base of Subzone CN10c, first appears in Samples 135-840B-13X-1, 74–75 cm, and 135-840C-5H-CC. A specimen showing some of the morphologic characteristics of both *Ceratolithus acutus* and *C. rugosus* is present in Sample 135-840B-11X-2, 19–20 cm.

Subzone CN10b is defined by the range of the marker species *C. acutus*, which is present in Hole 840B from Samples 135-840B-13X-CC through -18X-CC and in Hole 840C in Samples 135-840C-8H-CC and -10H-CC, with a questionable occurrence in Sample 135-840C-7H-CC.

Subzone CN10a is recognized in Samples 135-840B-20X-CC and -22X-CC and in Samples 135-840C-11H-CC and -12H-CC by the presence of *Triquetrorhabdulus rugosus* and the absence of *Ceratolithus acutus* and *D. quinqueramus*. The single specimen of *Amaurolithus amplifiscus* observed in Sample 135-840B-13X-CC (Subzone CN10b) is reworked. It is absent from the lower 12 samples (135-840B-15X-CC through -26X-CC), but it occurs again in 10 samples between 135-840B-28X-2, 41–42 cm, and -52X-CC. Gartner (1990) places both the first and last occurrences of this species within Subzone CN9b.

Miocene

An extensive section of late Miocene Zone CN9 is present in Hole 840B from Samples 135-840B-23X-CC to -63X-CC at the bottom of the hole. The occurrence together of *D. quinqueramus* and *Amaurolithus* species in Samples 135-840B-23X-CC through -52X-CC allows

assignment to Subzone CN9b. Samples 135-840B-55X-CC through -63X-CC are tentatively assigned to Subzone CN9a on the basis of the presence of *D. quinqueramus* and the absence of *Amaurolithus* species. Only the deepest sample from Hole 840C (135-840C-13H-CC) contained *D. quinqueramus* and *Amaurolithus* species.

Site 841

Site 841 is located in the Tonga Trench, west of the axis of the trench, at a water depth of 4810 m and approximately 48 km south southeast of Site 840. Three holes were drilled at Site 841; core recovery at Holes 841A and 841B was 37.7% and 27.9%, respectively. Hole 841A was cored from the seafloor surface to 186.6 mbsf. Hole 841B was washed to a depth of 169.8 mbsf and then cored to a total depth of 834.2 mbsf. Hole 841C was drilled specifically for logging, and no samples were taken. The sedimentary sequence cored in Hole 841A ranges from Pleistocene Subzone CN14b to late Miocene (Zone CN8). Hole 841B recovered a late Miocene (CN8) to middle or late Eocene (Subzones CP14a–CP15b) sedimentary sequence above a silicic volcanic basement. Poor core recovery and the presence of ash layers and turbidites limit biostratigraphic resolution at this site.

In general, nannofossils are less abundant and less well preserved at Site 841 than at Site 840, but exceptions do occur. For example, preservation is good and abundance is relatively high in the middle part of the interval assigned to late Miocene Subzone CN9b (Samples 135-841A-8H-4, 110–111 cm, through -10X-CC) (Table 5). Also, nannofossils are abundant and moderately well preserved in the Eocene and early Oligocene parts of Hole 841B, as represented by Samples 135-841B-46R-1, 139–140 cm, through -42R-2, 59–60 cm (Table 6). As at Site 840, small placoliths (<4 µm), which are difficult to identify to species with the light microscope, dominate the assemblages, and these have been grouped as “small placoliths” in Tables 5 and 6. Barren intervals, probably indicating deposition below the calcite compensation depth (CCD), are identified between the upper Miocene and middle Pleistocene in Hole 841A (Samples 135-841A-1H-5, 28–29 cm, through -6H-4, 95–96 cm). Barren intervals and those with poor and no age-diagnostic nannofossils in the middle and upper Miocene in Hole 841B, which are represented by Samples 135-841B-19R-3, 70–71 cm, through -32R-1, 45–46 cm, occur in volcanic conglomerates. Barren samples and most of those that lack age-diagnostic nannofossils are listed as footnotes to Tables 5 and 6.

Pleistocene

Zone CN14 was recognized in two samples from Hole 841A (Table 5). Both samples contain siliceous microfossil debris and reworked nannofossils. Both also contain specimens of *Coccolithus pelagicus* with a bar (Plate 1, Figs. 1–2). Sample 135-841A-1H-1, 56–59 cm, is assigned to Subzone CN14b on the basis of the presence of *G. oceanica* and lack of *E. ovata* and *E. huxleyi*. Rare, reworked Miocene *Discoaster bollii* and *Triquetrorhabdulus rugosus* are present. Nannofossils in Sample 135-841A-1H-3, 72–73 cm, are poorly preserved, showing signs of dissolution. This sample can only be assigned to Subzone CN14a or CN14b based on the presence of *G. oceanica* and, because of poor preservation, the inability to determine whether *E. ovata* is present. Reworked Eocene and Miocene nannofossils are common in this sample and include *Cyclicargolithus floridanus*, *Dictyococcites bisectus*, *Discoaster sublodoensis*, and *Triquetrorhabdulus farnsworthii*.

Miocene

A barren section consisting of clay with interbeds of vitric sand, vitric silt, and fine ash, extending from Samples 135-841A-1H-5, 28–29 cm, through -6H-4, 95–96 cm, separates the Pleistocene (CN14) from the upper Miocene (CN9). The absence of nannofossils may be a result of dissolution caused by deposition well below the CCD.

Table 3 (continued).

Subzones	Core, section, interval (cm)	<i>Helicosphaera sellii</i>	<i>Helicosphaera wallitchii</i>	<i>Helicosphaera</i> spp.	<i>Minyitha convallis</i>	<i>Oolithus fragilis</i>	<i>Pontosphaera</i> spp.	<i>Pseudoemiliania lacunosa</i>	<i>Reticulofenestra pseudoumbilica</i>	<i>Reticulofenestra</i> spp.	<i>Rhabdosphaera</i> spp.	<i>Scyphosphaera</i> spp.	<i>Sphenolithus</i> spp.	<i>Triquetrorhabdulus farnsworthii</i>	<i>Triquetrorhabdulus rugosus</i>	<i>Umbellosphaera irregularis</i>	Small geophycapsids	Small placoliths
CN14b	135-840B-1X-1, 33 1X-CC, 18 1X-CC	F	F	.	.	.	F	F	A	A
CN14a	2X-CC (a)	.	.	A	.	A	F	A	.	C	A	A	A
CN13b	2X-CC (b) 3X-CC	A	.	.	.	C	R	A	.	A	.	A	r	.	.	F	.	A
CN13a/b	4X-CC	.	R	.	.	R	R	F	.	A	R	A
CN12aC CN12aA	9X-CC 10X-CC	C	C	C	.	C	.	F	.	.	.	F	.	V
CN10d	11X-1, 22 11X-1, 65 11X-2, 19 11X-2, 60 11X-CC	.	.	.	f	C	.	.	F	A	.	F	C	V
CN10c	12X-1, 70 12X-CC 13X-1, 74	R	.	C	.	C	C	.	C	C	F	F	A	V
CN10b/c	13X-3, 64	R	.	C	.	R	R	.	C	A	R	F	A	r	.	.	.	V
CN10b	13X-CC 15X-CC 17X-CC 18X-CC	C	.	.	.	R	C	.	R	A	F	F	A	V
CN10a/b	19X-CC	C	.	.	.	C	C	.	A	F	.	F	A	.	r	.	.	V
CN10a	20X-CC 22X-CC	.	.	C	.	C	C	.	V	A	C	C	A	R	F	.	.	V
CN9b	23X-CC 24X-1, 50 24X-1, 140 24X-CC 26X-1, 48 26X-CC 28X-2, 41 28X-CC 29X-CC 30X-CC 31X-CC 32X-CC 33X-1, 75 33X-CC 34X-1, 90 34X-CC 36X-1, 50 36X-CC 37X-CC 38X-CC 40X-CC 42X-CC 43X-CC 45X-CC 50X-CC 51X-CC 52X-CC	C	C	.	C	V	C	C	A	.	F	.	.	A
CN9a	55X-CC 56X-CC 63X-1, 136 63X-4, 2 63X-CC	R	R	.	A	.	.	R	A	.	R	.	.	A

Discoaster berggrenii and *D. quinqueramus*, the marker species for Zone CN9, are present from Samples 135-841A-8H-4, 110–111 cm, through -11X-CC where preservation is good. The underlying Cores 135-841A-12X, -13X, and -14X are barren of nannofossils. The marker species for Zone CN9 are absent in Sample 135-841A-15X-1, 11–12 cm; however, *Amaurolithus* sp. aff. *A. amplificus* is present, and because *Amaurolithus* spp. first occur at the base of Subzone CN9b, this sample and those in Zone CN9 stratigraphically above were assigned to Subzone CN9b. Although the boundary between Zones CN8 and CN9 is not well defined in Hole 841A, Samples 135-841A-15X-CC through -17X-CC, 25–26 cm, have been assigned to Zone CN9 on the basis of the occurrence of *D. quinqueramus* in the latter sample. Samples 135-841A-18X-CC through -21X-CC are assigned to CN8 on the basis of the presence of *Minylitha convallis* and the absence of *D. berggrenii* and *D. quinqueramus*.

The LO of *Discoaster hamatus* marks the base of Zone CN8 (Bukry, 1973; Okada and Bukry, 1980). This species is not present in Holes 841A or 841B, but *M. convallis*, which first appears in Zone CN8 (Bukry, 1973) and whose FO has been used to approximate closely the base of Zone CN8 (Rio et al., 1990), is present. *M. convallis* occurs in the upper Miocene down to the bottom of Hole 841A and from Cores 135-841B-2R-CC through -16R-CC in Hole 841B. Other species present in this interval that typically occur in CN8 include *Discoaster bollii* and *D. neorectus* (FO in Subzone CN8b). The boundary between Subzones CN8a and CN8b is tentatively placed at the lowest occurrence of *D. sp. cf. D. neorectus* in Sample 135-841B-8R-CC. The boundary is questionable because nannofossil preservation is poor to moderate, and this species occurs only sporadically in this hole.

From Samples 135-841B-17R-4, 11–12 cm, through -32R-1, 45–46 cm, samples are barren or nannofossils are too sparse to determine the age of the sequence.

The LO of *H. ampliaperta* defines the boundary between Zones CN3 and CN4; and the LO of *S. heteromorphus* defines the boundary between Zones CN4 and CN5. Samples 135-841B-32R-1, 107–108 cm, through -41R-2, 92–93 cm, contain *Sphenolithus heteromorphus* and *Cyclicargolithus floridanus* but not *H. ampliaperta*, suggesting assignment to Zone CN4. However, it is not possible to determine if *H. ampliaperta* is absent because of an evolutionary LO, or if its absence is an artifact of preservation or because of ecologic factors. Rio et al. (1990) summarize possible alternative species that can be used to determine the boundary, but none are fully satisfactory for Site 841. In their study, they use the acme end of *D. deflandrei*, proposed by Bukry (1973), which is coincident with the FO of the *Discoaster signus*–*D. tuberi* group to mark the CN3/CN4 boundary. An incomplete section, because of a hiatus below Sample 135-841B-41R-2, 92–93 cm, and the few to rare, poorly preserved specimens of *D. deflandrei* above, does not permit the end of the acme to be determined in Hole 841B; however, the presence of *D. signus* in Sample 135-841B-40R-CC supports the Zone CN4 age assignment.

Triquetrorhabdulus sp. aff. *T. farnsworthii* is present in Sample 135-841B-41R-2, 92–93 cm, the lowest sample assigned to Zone CN4, and it occurs in several other samples above. The FO of *T. rugosus* at this site is higher, at Sample 135-841B-32R-1, 107–108 cm. Although *T. farnsworthii* is considered a junior synonym of *T. rugosus* (Lipps, 1969), it might prove useful to record their stratigraphic ranges separately to determine if they differ.

Oligocene and Eocene

A major hiatus occurs at Hole 841B where the lower middle Miocene sedimentary sequence rests unconformably on a lower Oligocene sequence. Nannofossils below the unconformity are common to abundant and moderately well preserved in most samples.

The three uppermost samples below the unconformity lack both *Discoaster barbadiensis* and *D. saipanensis*, the marker species for

the top of Subzone CP15b, but both contain *Ericsonia formosa*, the marker for the top of Subzone CP16b. The three samples are, therefore, assigned to the early Oligocene Subzones CP16a or CP16b. The co-occurrence in Sample 135-841B-43R-CC of *Isthmolithus recurvus*, the marker for the base of Subzone CP15b, and *D. saipanensis* allows assignment to late Eocene Subzone CP15b.

Samples 135-841B-44R-2, 27–28 cm, through -47R-2, 89–90 cm, contain nannofossils that can only be assigned to late middle or late Eocene Zones CP14 and CP15. Species present that are characteristic of these zones include *Criboecium reticulatum*, *Dictyococcites bisectus*, *Discoaster barbadiensis*, *D. saipanensis*, and *Ericsonia subdisticha*. The LOs of *C. reticulatum*, *D. barbadiensis*, and *D. saipanensis* in Subzone CP15b follow the same order as noted by Okada (1990) in samples from the Indian Ocean, with *C. reticulatum* disappearing first, followed by *D. barbadiensis*, and finally, *D. saipanensis* at the top of Subzone CP15b (Table 6).

Shallow-water nannofossils of the family Braarudosphaeraceae occur consistently in low to moderate abundance in Cores 135-841B-44R-CC through -47R-2, 20–22 cm; they are most abundant in Cores 135-841B-45R-1, 80–81 cm, through -45R-CC (Table 6). In most samples only isolated segments of pentoliths were found, but occasionally an entire pentolith is present. Species present include the following taxa: *Braarudosphaera bigelowii*, *B. discula*, *B. spp.*, *Micrantholithus* sp. aff. *M. flos*, *M. pinguis*, *M. spp.*, *Pemma basquense crassum*, and *P. papillatum* (Plate 1, Fig. 3). *Zygrhahlithus bijugatus*, a shallow-water holococcolith, is present throughout much of this Eocene and Oligocene section. Braarudosphaerids are not generally used for Cenozoic biostratigraphy. Bukry's zonal scheme is based primarily on open-ocean species. Gartner's (1971) zonation of the Eocene, which is based on samples from the Blake Plateau, is useful in a general way, but the order of FOs of his marker species differs somewhat from the order of the species at Hole 841B. This may be caused by the lack of a complete section for study because of the poor core recovery, local variations in ranges of species, poor preservation, or reworking. Nevertheless, the shallow-water assemblages at Site 841B support the Eocene age for Cores 135-841B-44R through -47R.

Chaproniere (this volume) and Nishi and Chaproniere (this volume) discuss Eocene larger shallow-water foraminifers, which are present in samples from Cores 135-841B-46R and -47R. These foraminifers are most abundant and diverse in Sample 135-841B-46R-CC, where they indicate water depths of 50–60 m. Thus, Chaproniere (this volume) concludes that Site 841 was at approximately 50 m water depth during the middle and late Eocene and has subsided to its present depth of 4809 m. Although the presence of braarudosphaerids usually indicates a shallow-water environment, the actual depth range for this group is unknown, especially for extinct such species as *Pemma papillatum*. Braarudosphaerids are present but sparse, possibly because of poor nannofossil preservation, in Sample 135-841B-46R-CC, where larger foraminifers are best represented. Poor nannofossil preservation in Samples 135-841B-47R-2, 20–21 cm, and -46R-2, 7–8 cm (Table 6) may also be the reason for the sparseness of braarudosphaerids in these samples.

Braarudosphaerids, especially *Pemma papillatum*, are most abundant in Samples 135-841B-45R-1, 80–81 cm, through -45R-CC, where larger foraminifers disappear, and planktonic foraminiferal abundances increase, indicating water depths greater than 100 m. This increase in braarudosphaerid abundance correlates with the beginning of an increase in the abundance of *Coccolithus pelagicus*, a cool water indicator, as well as with an increase of other oceanic nannofossils such as *Cyclicargolithus floridanus*, *Dictyococcites bisectus*, *Reticulofenestra*, and *Discoaster* species. However, braarudosphaerids are no longer present when these oceanic species reach their maximum abundances in Sample 135-841B-43R-CC and above. This suggests that the braarudosphaerids were able to tolerate greater water depths and cooler temperatures than the larger foraminifers. The trend to cooler waters during the late Eocene and early Oligocene is also

Table 4. Abundance, preservation, and distribution of calcareous nannofossils, Hole 840C.

Subzones	Core, section, interval (cm)	Abundance	Preservation	<i>Amaurolithus bizarrus</i>	<i>Amaurolithus delicatus</i>	<i>Amaurolithus ninae</i>	<i>Amaurolithus primus</i>	<i>Amaurolithus delicatus/primus</i> intergrade	<i>Amaurolithus tricorniculatus</i>	<i>Amaurolithus</i> spp.	<i>Calcidiscus leporinus</i>	<i>Calcidiscus macintyreii</i>	<i>Calcidiscus</i> sp. A	<i>Ceratolithus acutus</i>	<i>Ceratolithus armatus</i>	<i>Ceratolithus rugosus</i>	<i>Coccolithus pelagicus</i>	<i>Dicyclococites</i> spp.	<i>Dicyclococites</i> spp. (<4 µm)	<i>Discoaster archipelagoensis</i>	<i>Discoaster asymmetricus</i>	<i>Discoaster blackstockae</i>	<i>Discoaster brouweri</i>	<i>Discoaster challengeri</i>	<i>Discoaster dilatatus</i>	<i>Discoaster exilis</i>	<i>Discoaster intercalaris</i>	<i>Discoaster pansus</i>
CN12c	135-840C-1H-2, 105 1H-CC	C A	P M	-	-	-	-	-	-	-	R R	-	-	-	-	R	F	-	-	-	-	-	R R	-	-	-	-	-
CN12aC	2H-CC 3H-1, 28 3H-CC 4H-2, 97 4H-4, 69	A V A V V	P G M M G	-	-	-	-	-	-	-	C A C A	R C C C	-	-	-	F - C	-	-	-	-	F C C	-	R C A C	-	-	-	-	-
CN12aB	4H-CC	A	G	-	-	-	-	-	-	-	C	F	F	-	-	-	F	-	-	-	F	F	-	-	-	-	-	-
CN10d	5H-1, 53	V	M	-	R	-	-	-	R	R	A	-	C	-	-	-	C	-	-	C	R	F	-	-	-	-	F	-
CN10c	5H-2, 127 5H-CC	V A	M M	R	R	-	-	-	R	-	A	F	F	-	-	-	C	-	-	F	R?	C	C	C	-	-	-	C
CN10b/c	7H-CC	A	M	-	R	R	-	R	R	R	C	R	-	R?	-	-	C	-	-	-	-	-	F	-	R	-	-	R
CN10b	8H-CC 10H-CC	V V	M M	-	F	R	-	R	R	-	A	F	-	R	R	-	C	-	-	-	-	-	F	-	-	-	-	F
CN10a	11H-CC 12H-CC	A A	M P	-	F	-	-	R	-	R	C	-	-	-	-	-	C	-	-	-	-	-	F	-	-	-	-	-
CN9b	13H-CC	A	P	-	R	-	F	-	-	-	C	-	-	-	-	-	F	-	-	-	-	R	-	-	-	-	-	-

Notes: Abbreviations as in Table 2. The following samples are barren, or nearly barren, and contain no age-diagnostic nannofossils: 135-840C-2H-CC, 13 cm; -6H-CC; and -9H-CC.

reflected in the planktonic foraminifers, with increased abundances of cool-water indicators such as *Catapsydrax* in Sample 135-841B-45R-CC and above (Nishi and Chaproniere, this volume).

It is not known how calcareous nannoplankton species that are restricted to shallow-water deposits can have worldwide distribution. Yet, pentoliths of Eocene age have been reported from widely separated geographic areas. Bybell (1975) has discussed this problem in detail, so I will only summarize her work here. Pentoliths of Eocene age have been found in hemipelagic deposits in the United States (California and the Gulf Coast), Mexico, Europe, USSR, India, off the shore of the Blake Plateau, and in our samples from the South Pacific on the Tonga Ridge. With only a few exceptions, they are absent from open-ocean deposits of Eocene age. Many possible explanations have been offered for the restricted distribution, including the effects of turbidity and salinity, paleocurrents, and physiochemical factors affecting preservation.

Parke and Adams (1960) have noted in laboratory culture experiments that some coccolith species have different phases in their life cycles and produce one form of coccolith (calcareous plate) when in a hemipelagic environment, and another form when living in an open-ocean environment. For example, the coccolithophorid *Crystallolithus hyalinus* is the motile phase in the life history of the non-motile *Coccolithus pelagicus*. Tappan (1980) reported that *Crystallolithus hyalinus* produces plates composed of small loosely packed crystals that disintegrate easily and are not likely to be preserved in the geologic record. *Coccolithus pelagicus* produces larger, heavier plates with a more complex and stable crystal structure that favors preservation.

Laboratory studies are needed to determine if the shallow-water Braarudosphaeraeae have this type of life cycle. If they do, it would provide a mechanism for the organism to be geographically widespread, but usually restricted to shallow-water deposits. The form more susceptible to disintegration could live, reproduce, and be widely dispersed in the oceanic environment, but it would not be preserved in oceanic sediments.

SUMMARY AND CONCLUSIONS

Tables 7 and 8 summarize the nannofossil biostratigraphy for Sites 840 and 841. In spite of the poor recovery in some sections of the sequence drilled, and the abundance of volcanic material and turbidite beds, a relatively complete biostratigraphic zonation has been possible.

Nannofossil assemblages from Site 840 on the Tonga Ridge range in age from late Pleistocene or Holocene Zone CN15 through late Miocene Zone CN9. The absence of mid Pliocene Zone CN11 at Site 840B is interpreted as a hiatus.

At Site 841 in the Tonga Trench, the age of the sedimentary sequence drilled ranges from middle Pleistocene Subzone CN14b through middle or late Eocene Zones CP14 or CP15 and overlies a rhyolitic volcanic basement. A major hiatus occurs where lower middle Miocene strata (CN4) rest unconformably on lower Oligocene strata (CP16). A shallow-water Eocene nannofossil assemblage consisting of *Braarudosphaera*, *Micrantholithus*, and *Pemma* is present in Cores 44 through 47. A trend toward cooler and deeper water starts to be evident in the middle or late Eocene (Sample 135-841B-45R-

Table 4 (continued).

<i>Discoaster pentaradiatus</i>	<i>Discoaster</i> sp. aff. <i>Discoaster pentaradiatus</i>	<i>Discoaster quadramus</i>	<i>Discoaster quinqueramus</i>	<i>Discoaster surculus</i>	<i>Discoaster tamafis</i>	<i>Discoaster triradiatus</i>	<i>Discoaster variabilis</i>	<i>Discoaster</i> spp.	<i>Emilitiana ovata</i>	<i>Havaster perplexus</i>	<i>Helicosphaera carteri</i>	<i>Helicosphaera</i> sp. aff. <i>Helicosphaera intermedia</i>	<i>Helicosphaera sellii</i>	<i>Helicosphaera</i> spp.	<i>Minylitha convallis</i>	<i>Oolithonus fragilis</i>	<i>Pontosphaera</i> spp.	<i>Pseudoemiliania lacunosa</i>	<i>Reticulofenestra pseudoumbilica</i>	<i>Reticulofenestra</i> spp.	<i>Rhabdosphaera claviger</i>	<i>Scyphosphaera</i> spp.	<i>Sphenolithus</i> spp.	<i>Triquetrorhabdulus farnsworthii</i>	<i>Triquetrorhabdulus rugosus</i>	<i>Umbellosphaera irregularis</i>	Small <i>gephyrocapsids</i> (with bars)	Small placoliths
R	-	-	-	-	-	-	-	R	F	-	R	-	-	R	-	-	-	-	-	C	-	-	F	r	-	-	R?	F
F	R	-	-	-	-	-	-	-	C	-	F	-	R	-	-	R	F	C	-	-	-	-	-	-	-	R	-	A
F	F	-	-	-	R	F	-	-	C	R	F	-	F	-	-	C	R	C	-	-	-	-	-	-	r	-	-	A
A	C	R	-	C	C	-	-	C	A	C	C	-	C?	-	-	C	C	A	-	-	C	F	-	-	-	-	-	V
C	R	-	-	C	C	F	-	-	C	C	-	-	R	A	-	C	C	C	-	-	A	F	-	-	-	-	-	A
A	C	-	-	A	A	-	-	A	A	C	C	-	C	A	-	A	C	A	-	-	A	C	-	-	-	-	-	V
A	C	-	-	A	C	F	-	C	A	C	C	-	A	C	-	A	C	A	-	-	A	C	-	-	-	C	-	V
C	-	-	-	F	F	-	R	C	C	C	C	-	C	C	-	F	F	C	-	-	F	F	-	-	-	-	-	A
C	C	-	-	F	-	-	R	C	-	C	C	-	C	C	r?	C	C	-	F	A	C	F	A	-	-	-	-	V
C	C	-	-	C	R?	-	C	C	-	-	C	F	-	-	f?	F	F	-	-	C	C	F	A	-	-	-	-	V
F	C	-	-	C	-	-	-	F	-	-	F	-	-	-	f?	R	R	-	-	C	-	F	C	-	-	-	-	A
C	R	-	-	F	-	-	F	-	-	R	C	-	-	-	-	-	-	-	-	C	R	R	C	-	r	-	-	A
A	-	-	-	C	-	-	C	-	-	C	C	C	R	C	-	-	C	-	-	A	C	C	A	-	-	-	-	V
A	R	-	-	C	-	-	A	-	-	A	-	-	-	-	-	-	C	-	F	A	F	A	A	-	-	-	-	V
F	-	-	-	R	-	-	F	C	-	R	C	-	-	C	-	F	F	-	F	A	F	-	A	-	R	-	-	A
F	-	-	-	R	-	-	F	C	-	R	F	-	-	F	-	R	-	-	R	C	C	R	C	-	-	-	-	C
F	-	-	R	-	-	-	F	F	-	-	C	-	-	-	-	F	-	F	-	C	-	R	C	-	-	-	-	A

CC) and becomes more pronounced during the latest Eocene and early Oligocene (Samples 135-841B-43R-CC and above).

ACKNOWLEDGMENTS

I appreciate very much the review of an earlier version of this manuscript by David Bukry (U.S.A.), Samir Shafik (Australia), To-shiaki Takayama (Japan), and Michael Styzen (U.S.A.). I especially thank David Bukry for his discussions on the taxonomy of *Calcidiscus*.

TAXONOMIC NOTES

Calcidiscus spp.

Three morphotypes of *Calcidiscus* were recognized in samples from Holes 840B and 840C. Specimens 10 µm or larger are listed as *C. macintyreii* on Tables 3 and 4. Those <10 µm and having a nearly closed, circular central area are listed as *C. leptoporus*. Specimens assigned to *C. sp. A* are similar in size to *C. leptoporus* but have a structure within the central area. In phase contrast illumination (Plate 1, Fig. 4), *C. sp. A* appears to have a honeycomb or gridlike structure in the central area and, thus, somewhat resembles *Cycloperfoliithus carlae* Lehotayova and Priewalder (1978). However, the gridlike structure in *C. carlae* has many pores arranged in fivefold symmetry, whereas *C. sp. A* appears to have a total of seven or eight pores. Also, *C. carlae* is generally larger (7.5–12 µm) than *C. leptoporus*.

Three samples from Hole 840B in which *C. sp. A* had been identified by light microscope were examined by SEM. No specimens with a honeycomb structure were observed, but one specimen with projections extending into the center was present (Plate 1, Fig. 5). These projections may be the remnants of partially dissolved rim elements, which only appear to have a honeycomb-like structure when observed in phase contrast illumination. The short bars extend-

ing partly across the central perforation in specimens of *C. leptoporus*, which were figured by Gartner (1967), may be similar to the structures in *C. sp. A*.

Coccolithus pelagicus

Coccolithus pelagicus with a bar (Plate 1, Figs. 1–2) is present in Samples 135-841A-1H-1, 56–59 cm, and 135-841A-1H-3, 72–73 cm. Parke and Adams (1960) recorded *C. pelagicus* with a bar and assigned it to *Crystallolithus hyalinus* (the motile phase in the life history of the non-motile *C. pelagicus*). Perch-Nielsen (1985) figured the barred morphotype but did not discuss the biostratigraphic or paleoecologic significance. Bukry (1991) noted the presence of both barred and nonbarred *C. pelagicus* but did not separate them for his counts. It may be useful to study the distribution of the barred and nonbarred morphotypes to see if an ecological significance exists.

Discoaster sp. aff. *D. pentaradiatus*

Birefringent discoasters that are more robust than typical *D. pentaradiatus* and have five nonbifurcating bluntly pointed rays are listed in Tables 3 and 4 as *D. sp. aff. D. pentaradiatus*. *Discoaster ono*, a new species described by Styzen (this volume) was not counted separately, but was included in the tables as *D. sp. aff. D. pentaradiatus* along with larger forms having longer rays.

Helicosphaera spp.

Specimens with an optically discontinuous bridge and resembling *H. intermedia* or *H. euphratis* are listed in Tables 3 through 6 as *H. sp. aff. H. intermedia*. They occur in samples as young as Pliocene Subzone CN10c. However, species with an optically discontinuous bridge are reported to have their LO during the early or early middle Miocene (Perch-Nielsen, 1985; Aubry, 1990).

Table 5 (continued).

Zone or Subzone	Core, section, interval (cm)	<i>Discoaster subdoensis</i>	<i>Discoaster surculus</i>	<i>Discoaster toralus</i>	<i>Discoaster variabilis</i>	<i>Discoaster</i> spp.	<i>Emiliana ovata</i>	<i>Ericsonia formosa</i>	<i>Gephyrocapsa caribbeanica</i>	<i>Gephyrocapsa oceanica</i>	<i>Helicosphaera carteri</i>	<i>Helicosphaera</i> sp. aff. <i>Helicosphaera intermedia</i>	<i>Helicosphaera</i> spp.	<i>Micrantholithus</i> spp.	<i>Minylitha convallis</i>	<i>Pemna papillatum</i>	<i>Pontosphaera</i> spp.	<i>Pseudoemiliana lacunosa</i>	<i>Reticulofenestra pseudumbilica</i>	<i>Reticulofenestra</i> spp.	<i>Rhabdosphaera</i> spp.	<i>Sphenolithus</i> spp.	<i>Triquetrorhabdulus farnsworthii</i>	<i>Triquetrorhabdulus rugosus</i>	<i>Umbellosphaera irregularis</i>	Small placoliths	Pentolith fragments
CN14b	135-841A 1H-1, 56	C	F	r	R	F	.	f	r	R	A	.
CN14a/b	1H-3, 72	r	.	.	.	c	R?	.	F	C	F?	f	F	.	f	r	.	.	A	.
CN9b	8H-4, 110	.	.	R	F	F	.	r	.	.	F	F	R	r	F	.	R	.	.	C	.	C	F	.	.	C	.
	8H-CC	.	R	.	.	F	R	.	F	.	.	.	C	.
	9X-1, 3	F	F	R	R	.	.	F	.
	9X-CC	.	.	.	F	F	F	.	.	F	.	.	C	.
	10X-CC	.	R	.	C	C	R	C	.	.	C	.	.	C	r
11X-CC	R	R	.	R?	R	.	
15X-1, 11	.	.	.	F	F	R	.	.	C	.	R	.	.	C	.	.	R	R	.	.	A	.
CN9a/b	15X-CC	.	.	R	F	F	R	F	.	R	F	F	.	R	R	.	.	F	.
	16X-CC	.	.	.	F	F	C	F	C	C	.	.	R	.	F	F	.	C	R	.	.	C	.
	17X-CC, 25	.	.	.	F	R	.	r	.	.	F	F	.	C	r	.	R	.	R	C	.	C	.	.	.	C	r
CN8	18X-CC	R	F
	19X-CC	R
	20X-CC	.	.	R	F	C	F	.	C	F	F	.	C	.	.	C	.	
	21X-CC	.	.	.	F	C	C	F	C	.	C

Discoaster extensus Hay (1967)
Discoaster giganteus (Theodoridis, 1984)
Discoaster intercalaris Bukry (1971)
Discoaster kuepperi Stradner (1959)
Discoaster loeblichii Bukry (1971)
Discoaster mendumobensis Wise (1973)
Discoaster moorei Bukry (1971)
Discoaster neorectus Bukry (1971)
Discoaster pansus (Bukry and Percival, 1971) Bukry (1973)
Discoaster pentaradiatus Tan (1927) emend. Bramlette and Riedel (1954)
Discoaster perclarus Hay in Hay et al. (1967)
Discoaster phylloides Hay (1967)
Discoaster quadramus Bukry (1973)
Discoaster quinqueramus Gartner (1969)
Discoaster saipanensis Bramlette and Riedel (1954)
Discoaster signus Bukry (1971)
Discoaster sublodoensis Bramlette and Sullivan (1961)
Discoaster surculus Martini and Bramlette (1963)
Discoaster tamalis Kamptner (1967)
Discoaster tani tani Bramlette and Riedel (1954)
Discoaster tani nodifer Bramlette and Riedel (1954)
Discoaster toralus Ellis, Lohmann and Wray (1972)
Discoaster triradiatus Tan (1927)
Discoaster variabilis Martini and Bramlette (1963)
Emiliana huxleyi (Lohmann, 1902) Hay and Mohler in Hay et al. (1967)
Emiliana ovata (Cohen, 1964) Bukry (1973)
Ericsonia formosa (Kamptner, 1963) Haq (1971)
Ericsonia subdisticha (Roth and Hay in Hay et al., 1967) Roth in Baumann and Roth (1969)
Gephyrocapsa caribbeanica Boudreaux and Hay (1969)
Gephyrocapsa oceanica Kamptner (1943)
Hayaster perplexus (Bramlette and Riedel, 1954) Bukry (1973)
Helicosphaera bramlettei Müller (1970)
Helicosphaera burkei Black (1971)

Helicosphaera carteri (Wallich, 1877) Kamptner (1954)
Helicosphaera compacta Bramlette and Wilcoxon (1967)
Helicosphaera intermedia Martini (1965)
Helicosphaera inversa Gartner (1980)
Helicosphaera reticulata Bramlette and Wilcoxon (1967)
Helicosphaera salebrosa Perch-Nielsen (1971)
Helicosphaera sellii Bukry and Bramlette (1969)
Helicosphaera wallichii (Lohmann, 1902) Boudreaux and Hay (1969)
Isthmolithus recurvus Deflandre (1954)
Isthmolithus unipons Bramlette and Sullivan (1961)
Micrantholithus flos Deflandre in Deflandre and Fert (1954)
Micrantholithus pinguis Bramlette and Sullivan (1961)
Minylitha convallis Bukry (1973)
Oolithus fragilis (Lohmann, 1912) Martini and Muller (1972)
Pemna basquense crassum (Bouche) Bybell and Gartner (1972)
Pemna papillatum Martini (1959)
Pontosphaera indoceanica Čepik (1973)
Pseudoemiliana lacunosa (Kamptner, 1963) Gartner (1969)
Reticulofenestra dictyoda (Deflandre in Deflandre and Fert, 1954) Stradner in Stradner and Edwards (1968)
Reticulofenestra hillae Bukry and Percival (1971)
Reticulofenestra pseudumbilica (Gartner, 1967) Gartner (1969)
Reticulofenestra umbilica (Levin, 1965) Martini and Ritzkowski (1968)
Rhabdosphaera claviger Murray and Blackman (1898)
Scyphosphaera sp. Lohmann (1902)
Sphenolithus abies Deflandre in Deflandre and Fert (1954)
Sphenolithus anarrhopus Bukry and Bramlette (1969)
Sphenolithus celsus Haq (1971)
Sphenolithus compactus Backman (1980)
Sphenolithus distentus (Martini, 1965) Bramlette and Wilcoxon (1967)
Sphenolithus heteromorphus Deflandre (1953)
Sphenolithus intercalaris Martini (1976)
Sphenolithus moriformis (Brönnimann and Stradner, 1960) Bramlette and Wilcoxon (1967)

Table 6. Abundance, preservation, and distribution of calcareous nannofossils, Hole 841B.

Zone or Subzone	Core, section, interval (cm)	Abundance	Preservation	<i>Biantholithus sparsus</i>	<i>Braarudosphaera bigelowii</i>	<i>Braarudosphaera discuta</i>	<i>Braarudosphaera</i> spp.	<i>Bramletteius serraculoides</i>	<i>Calcidiscus leptoporus</i>	<i>Calcidiscus mucintyreii</i>	<i>Calcidiscus protoannulatus</i>	<i>Chiasmolithus</i> sp. cf. <i>Chiasmolithus modestus</i>	<i>Coccolithus eopelagicus</i>	<i>Coccolithus pelagicus</i>	<i>Cribocentrum reticulatum</i>	<i>Cyclargolithus abisectus</i>	<i>Cyclargolithus floridanus</i>	<i>Dietycococites bisectus</i>
CN8b	135-841B-2R-3, 32	C	M	R
	2R-CC	A	M	F	R	.	.	.	C
	3R-1, 12	F	P	r	.	.
	4R-CC	A	M	F	F	.	.	.	r
	5R-2, 80	R	M
	5R-2, 132	R	M
	5R-CC	C	M	F	F	.	.	.	F	.	.	.	r
	6R-CC	A	M	F	R	.	.	.	F
	7R-CC	C	M	R	.	.	.	F
	8R-1, 52	C	P	f	f	.
8R-CC	C	P	R	R	
CN8a	9R-1, 91	R	P	r	.
	9R-CC	C	P	R	R
	10R-CC	F	P
	11R-CC	F	P	R
	12R-CC	F	P	R
	14R-CC	C	P	F	R	.	.	.	F
	15R-CC	F	P	F	R	.	.	.	F
	16R-CC	F	P	R
CN4	32R-1, 107	F	M	R	.	F	F	.
	32R-CC	C	M	R	C	.
	33R-3, 44	F	P	f	R	.	F	F	.
	33R-CC	A	M	R	.	F	A	.
	34R-CC	A	M	R	R	.	F	C	.
	35R-1, 31	C	P	F	C	.
	35R-1, 140	F	M
	35R-CC	C	M	F	.	R	C	r
	36R-CC	F	M	R	.	R	F	.
	37R-1, 122	F	P	F	F	.
	39R-CC	C	M	F	F	.
40R-CC	C	P	.	r	.	.	.	R	F	.	F	C	.	
41R-2, 92	F	M	R	F	.	
CP16	42R-2, 59	A	M	r	.	C	.	.	A	C
	42R-4, 61	A	M	A	.	F	A	C
	42R-CC	A	M	f	C	A	.	.	A	A
CP15b	43R-CC	A	M	.	.	.	R	C	.	.	A	A	
CP14/CP15	44R-2, 27	A	M	F	C	.	.	F	C
	44R-CC	A	M	R	F	.	F	C	C	.	C	C
	45R-1, 80	A	M	C	.	F	F	R	C	C	.	C	C
	45R-1, 140	A	M	.	.	R	R	C	C	.	C	C
	45R-CC	A	M	.	F	.	F	R?	C	C	.	.	C	C
	46R-1, 139	A	M	.	.	.	R	C	R	R	C	F
	46R-2, 7	C	P	R	.	.	R	F	F	.	.	C	F
	46R-CC	C	P	R	.	.	R	.	.	.	R	.	.	R	.	.	F?	F
	47R-2, 20	C	P	.	.	.	R	F	.	F	.
	47R-2, 89	C	M	F	R	R	F	R

Notes: Abbreviations as in Table 2. The following samples are barren, or nearly barren, and contain no age-diagnostic nannofossils: 135-841B-4R-2, 61 cm; -10R-1, 60 cm; -11R-2, 10 cm; -13R-4, 13 cm; -16R-1, 45 cm; -16R-3, 33 cm; -17R-4, 11 cm; -17R-4, 64 cm; -18R-1, 107 cm; -18R-2, 95 cm; -19R-3, 70 cm; -20R-1, 88 cm; -21R-1, 79 cm; -21R-1, 89 cm; -22R-1, 86 cm; -23R-2, 92 cm; -24R-1, 83 cm; -26R-1, 124 cm; -27R-1, 6 cm; -28R-2, 18 cm; -29R-2, 25 cm; -29R-2, 79 cm; -30R-2, 18 cm; -31R-CC; -32R-1, 45 cm; -32R-1, 127 cm; -33R-3, 63 cm; -34R-1, 136 cm; -34R-2, 2 cm; -35R-1, 140 cm; -35R-3, 59 cm; -35R-3, 126 cm; -36R-1, 47 cm; -37R-2, 80 cm; -38R-2, 38 cm; -38R-4, 33 cm; -39R-2, 5 cm; -39R-2, 106 cm; -40R-2, 79 cm; -40R-2, 149 cm; -41R-3, 107 cm; -41R-3, 119 cm; -41R-3, 144 cm; and -54R-2, 51 cm.

Table 6 (continued).

Zone or Subzone	Core, section, interval (cm)	<i>Helicosphaera carteri</i>	<i>Helicosphaera compacta</i>	<i>Helicosphaera</i> sp. aff. <i>Helicosphaera intermediata</i>	<i>Helicosphaera reticulata</i>	<i>Helicosphaera salebrosa</i>	<i>Helicosphaera sellii</i>	<i>Helicosphaera</i> spp.	<i>Ishmolithus recurvus</i>	<i>Ishmolithus unipons</i>	<i>Micrantholithus flos</i>	<i>Micrantholithus pinguis</i>	<i>Micrantholithus</i> spp.	<i>Minyolitha convallis</i>	<i>Oolithus</i> spp.	<i>Pemma basquense crassum</i>
CN8b	135-841B-2R-3, 32	R	R
	2R-CC	C	.	F	.	.	.	C	C	.	.
	3R-1, 12
	4R-CC	F	.	R	C	.	.
	5R-2, 80	R	.	.
	5R-2, 132	F	.	.
	5R-CC	R	C	.	.
	6R-CC	C	.	.
	7R-CC	R	C	F	.
	8R-1, 52	C	.	.
8R-CC	C	.	.	
CN8a	9R-1, 91	R	.	.
	9R-CC	C	.	.
	10R-CC	F	.	.
	11R-CC	F	.	.
	12R-CC	F	.	.
	14R-CC	F	F	.	.
	15R-CC	F	.	.
16R-CC	F	.	.	
CN4	32R-1, 107
	32R-CC
	33R-3, 44	R
	33R-CC	R
	34R-CC	F
	35R-1, 31	F
	35R-1, 140	R	R	?	R	.
	35R-CC	R	.	R
	36R-CC
	37R-1, 122	.	.	R	R	.
39R-CC	F	R	
40R-CC	F	F	
41R-2, 92	R	
CP16	42R-2, 59	.	R	F	.	r?
	42R-4, 61	.	F	R
	42R-CC	.	C	F
CP15b	43R-CC	.	R	R	
CP14/CP15	44R-2, 27	.	F
	44R-CC	.	.	.	R
	45R-1, 80	.	.	R	R	.	.	R
	45R-1, 140	R	F	.	.	.	C
	45R-CC	F	.	.	F	.	F	.	.	.
	46R-1, 139
	46R-2, 7	R	.	.	.
	46R-CC	F	.	.	.
	47R-2, 20	R	R	.	.	.
47R-2, 89	

Table 7. Summary chart of nannofossil biostratigraphy, Site 840.

Epoch	Zone or subzone	Hole 840A	Hole 840B	Hole 840C
Holocene or Pleistocene	CN15	1H-1, 45		
Pleistocene	CN14b	1H-2, 3 1H-2, 140 1H-3, 32 1H-CC	1X-1, 33 1X-CC, 18 1X-CC	
	CN14a		2X-CC (a)	
late Pliocene	CN13b		2X-CC (b) 3X-CC	
	CN13a/b		4X-CC	
	CN12c			1H-2, 105 1H-CC
	CN12aC		9X-CC	2H-CC 3H-1, 28 3H-CC 4H-2, 97 4H-4, 69
	CN12aB			4H-CC
	CN12aA		10X-CC	
early Pliocene	CN10d		11X-1, 22 11X-1, 65 11X-2, 19 11X-2, 60 11X-CC	5H-1, 53
	CN10c		12X-1, 70 12X-CC 13X-1, 74	5H-2, 127 5H-CC
	CN10b/c		13X-3, 64	7H-CC
	CN10b		13X-CC 15X-CC 17X-CC 18X-CC	8H-CC 10H-CC
	CN10a/b		19X-CC	
	CN10a		20X-CC 22X-CC	11H-CC 12H-CC
late Miocene	CN9b		23X-CC 34X-1, 90 24X-1, 50 34X-CC 24X-1, 140 36X-1, 50 24X-CC 36X-CC 26X-1, 48 37X-CC 26X-CC 38X-CC 28X-2, 41 40X-CC 28X-CC 42X-CC 29X-CC 43X-CC 30X-CC 45X-CC 31X-CC 50X-CC 32X-CC 51X-CC 33X-1, 75 52X-CC 33X-CC	13H-CC
	CN9a		55X-CC 56X-CC 63X-1, 136 63X-4, 2 63X-CC	

Sphenolithus neobabies Bukry and Bramlette (1969)
Sphenolithus obtusus Bukry (1971)
Sphenolithus predistentus Bramlette and Wilcoxon (1967)
Sphenolithus pseudoradians Bramlette and Wilcoxon (1967)
Sphenolithus quadrispinatus Perch-Nielsen (1980)
Sphenolithus radians Deflandre in Grassé (1952)
Sphenolithus tribulosus Roth (1970)
Sphenolithus verensis Backman (1978)
Triquetrorhabdulus farnsworthii (Gartner, 1967) Perch-Nielsen (1984)
Triquetrorhabdulus rugosus Bramlette and Wilcoxon (1967)
Umbellosphaera irregularis Paasche in Markali and Paasche (1955)
Umbellosphaera sp. Lohmann (1902)
Zygrhablithus bijugatus (Deflandre in Deflandre and Fert, 1954) Deflandre (1959)

Table 8. Summary chart of nannofossil biostratigraphy, Site 841.

Epoch	Zone or subzone	Hole 841A	Hole 841B
Pleistocene	CN14b	1H-1, 56	
	CN14a/b	1H-3, 72	
late Miocene	CN9b	8H-4, 110 10X-CC 8H-CC 11X-CC 9X-1, 3 15X-1, 11 9X-CC	
	CN9a/b	15X-CC 16X-CC 17X-CC, 25	
	CN8b	18X-CC 19X-CC 20X-CC 21X-CC	2R-3, 32 5R-CC 2R-CC 6R-CC 3R-1, 12 7R-CC 4R-CC 8R-1, 52 5R-2, 80 8R-CC 5R-2, 132
	CN8a		9R-1, 91 12R-CC 9R-CC 14R-CC 10R-CC 15R-CC 11R-CC 16R-CC
middle Miocene	CN4		32R-1, 107 35R-CC 32R-CC 36R-CC 33R-3, 44 37R-1, 122 33R-CC 39R-CC 34R-CC 40R-CC 35R-1, 31 41R-2, 92 35R-1, 140
early Oligocene	CP16		42R-2, 59 42R-4, 61 42R-CC
late Eocene	CP15b		43R-CC
middle and late Eocene	CP14/CP15		44R-2, 27 46R-1, 139 44R-CC 46R-2, 7 45R-1, 80 46R-CC 45R-1, 140 47R-2, 20 45R-CC 47R-2, 89

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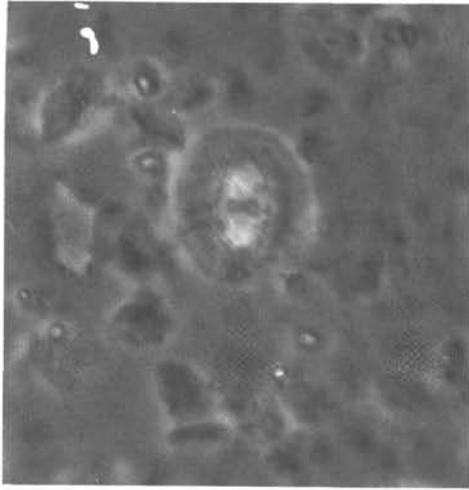
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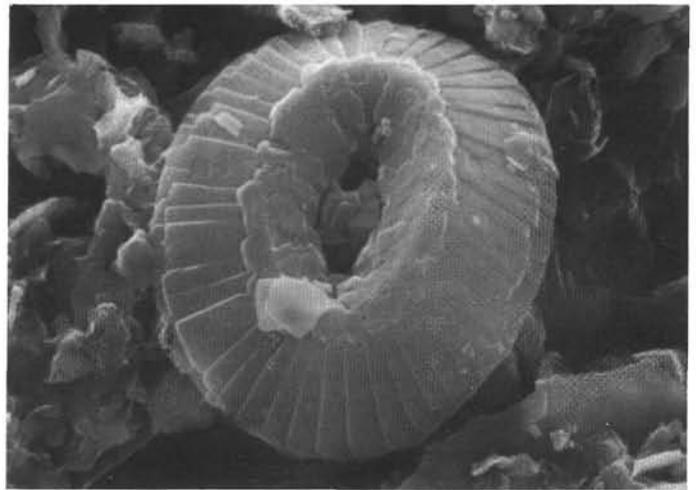
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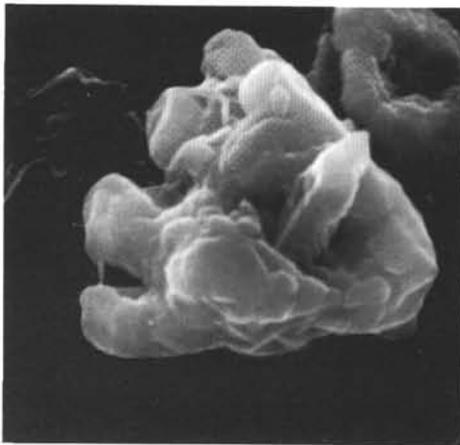


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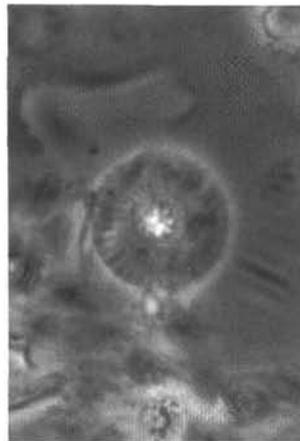
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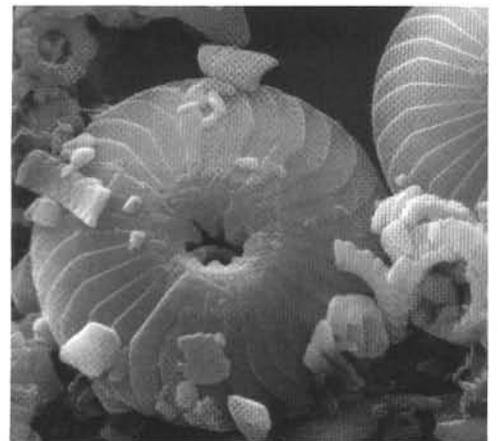


3

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4



5

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Plate 1. Cenozoic calcareous nannofossils. PC = phase contrast (magnifications approximate). 1. *Coccolithus pelagicus* with bar, PC, $\times 1800$. Sample 135-841A-1H-1, 56–59 cm. 2. *C. pelagicus* with bar, SEM. Sample 135-841A-1H-1, 56–59 cm. 3. Isolated segment of *Pemma papillatum*, SEM. Sample 135-841B-45R-1, 80–81 cm. 4. *Calcidiscus* sp. A, PC, $\times 2100$. Sample 135-840C-4H-2, 97–98 cm. 5. *C.* sp. A (?), SEM. Sample 135-840B-11X-CC.