

34. DATA REPORT: CENOZOIC RADIOLARIANS FROM LEG 143, HOLE 869A, EQUATORIAL PACIFIC OCEAN¹

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ABSTRACT

Cenozoic (lower Eocene to lower Miocene) radiolarians are abundant and exceptionally well-preserved in cores from Leg 143 Hole 869A. The Eocene/Oligocene boundary sequence appears conformable, whereas erosional hiatuses are present between the early Oligocene and early Miocene and between the middle and late Eocene. These hiatuses may represent periods of emergence.

INTRODUCTION

Hole 869A (Leg 143), located at 11°0.009'N, 164°44.696'W, is situated 45 nmi (83 km) southwest of the atoll-guyot pair of Bikini (formerly Bikini) Atoll and Wodejebatao (formerly Sylvania) Guyot in the equatorial Pacific Ocean. Hole 869A was an APC/XCB-cored hole that had 100% recovery in the APC-cored section. The hole was drilled in a water depth of 4826.7 m and penetrated 166.5 m of radiolarian ooze.

The area examined has long been known as one in which well-preserved radiolarian faunas are present. Riedel and Sanfilippo (1971) presented an extensive report on Cenozoic tropical radiolarians from the western Pacific Ocean in the *Initial Reports* volume of Deep Sea Drilling Project (DSDP) Leg 7. Abundant and exceptionally well-preserved radiolarians were recovered from cores throughout Hole 869A (Table 1). The primary purpose of this paper is to provide radiolarian age constraints on sediments recovered from this section.

METHODS

Samples were taken at least one per core, and most core-catcher samples were examined. More detailed examinations were conducted in the vicinity of particular sedimentological or radiolarian events. To obtain clean radiolarian concentrates for microscopic examination, sediments were disaggregated and sieved to remove the clay-silt fraction. A 5-cm³ sample was placed in a 400-cm³ beaker that contained 150 mL of a 10% solution of hydrogen peroxide and a small amount of Calgon (to aid in disaggregating the sediment). If calcareous components were evident, they were dissolved by adding hydrochloric acid. The residue was sieved through a 63-μm sieve, and the remaining siliceous microfossils were pipetted evenly onto labeled glass slides. The accompanying water then was evaporated under a heat lamp, after which the remaining residue was mounted using Norland Optical Adhesive and covered with a 22 × 50 mm cover slip. Two slides were prepared and examined for each sample. Qualitative assessments of the radiolarians in each slide were not recorded for abundance and preservation as, in all cases, radiolarians were abundant and well preserved. Radiolarians were profusely abundant in some samples, and complete faunal lists are not presented herein but are the topic of ongoing study to be presented later (Aitchison and Flood, unpubl. data).

BIOSTRATIGRAPHIC FRAMEWORK

The Cenozoic radiolarian zonation of Sanfilippo et al. (1985), derived for the tropical equatorial Pacific, was used at all sites. Sanfilippo et al. (1985) summarized the taxonomy and evolutionary lineages of all stratigraphically important radiolarian taxa commonly found in low-latitude regions of this zonation. When suggesting tentative "absolute" ages for radiolarian datum levels and zonal boundaries, the schemes of Nigrini (1985) and Barron et al. (1985), established on the basis of DSDP Leg 85 sites in the equatorial Pacific, were followed. Although much of the material obtained during Leg 85 could not be directly dated paleomagnetically, sufficient duplicate sites were available in which all major microfossil events could be identified, some of which had been correlated to the polarity time scale in nearby piston cores. Thus, the ages of Pacific radiolarian events, estimated by Foreman (1981), Nigrini (1985), and Barron et al. (1985), are considered to provide a satisfactory working model. Tentative correlation with calcareous nannofossil stratigraphy has been based on correlation charts of Haq et al. (1987).

LITHOSTRATIGRAPHY

Two lithostratigraphic units were identified in Hole 869A: (Unit I) cyclically bedded very pale brown to yellow-brown to dark reddish brown nannofossil ooze, radiolarian nannofossil ooze, and clayey nannofossil ooze; and (Unit II) cyclically bedded white to very pale brown to dark yellowish-brown clayey nannofossil ooze to nannofossil radiolarian ooze containing porcellanite and chert.

Unit I (0–88.20 mbsf, lower Miocene to upper Eocene)

The dominant sedimentary theme of Unit I is one of cycles, on various scales, which are characterized by regular changes in color and composition. The principal sedimentary components are nannofossils, radiolarians, sponge spicules, and clay; the colors vary from very pale brown to yellow-brown to dark reddish-brown, and the compositions range from nannofossil ooze through radiolarian nannofossil ooze to clayey nannofossil ooze. Thicknesses of different lithologies are on the scale of tens of centimeters, although some thinner bands do exist. Sub-unit IA is separated from Subunit IB by a stratigraphic hiatus. There is some reworking of Subunit IB into the basal bed of Subunit IA.

Unit II (88.2–166.5 mbsf, upper Eocene to middle Eocene)

As in Unit I, the dominant sedimentary theme of Unit II is one of cycles on various scales, characterized by regular changes in color and composition. In Core 143-869A-10X, the first porcellanite appears that is associated with clayey nannofossil ooze. Porcellanites and cherts appear intermittently at first, but increase in percentage

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Table 1. Age-diagnostic radiolarians present in samples from Hole 869A.

Epoch	Radiolarian zone	Core, section, interval (cm)	<i>Calocyctella virginis</i>	<i>Liriospyris mutuaria</i>	<i>Stichocorys delmontensis</i>	<i>Didymocyrta prismatica</i>	<i>Dorcadospyrus ateuchus</i>	<i>Centrobotrys thermophila</i>	<i>Lynchnocanoma elongata</i>	<i>Lithocyclia angusta</i>	<i>Tristylospyris triceros</i>	<i>Lynchodictyon audax</i>	<i>Artophormis gracilis</i>	<i>Centrobotrys petrushevskaya</i>	<i>Theocyrtis tuberosa</i>	<i>Thyscyrtis tetricantha</i>	<i>Thyscyrtis triacantha</i>	<i>Podocyrtis helene</i>	<i>Podocyrtis pasciolaria</i>	<i>Podocyrtis chalara</i>	<i>Calocyclas bandycra</i>	<i>Eusyringium fistuligerum</i>	<i>Calocyclus turris</i>	<i>Lithochytris vespertilio</i>	<i>Selachocyrtis triconicus</i>	<i>Podocyrtis mitra</i>	<i>Podocyrtis trachodes</i>	<i>Spongatractus pachystylus</i>	<i>Lithocelia ocellata</i>	<i>Thyscyrtis rhizodon</i>	<i>Dictyopora mongolfieri</i>	<i>Padocyrtis sinuosa</i>	<i>Eusyringium lagera</i>	<i>Phaeocystis striata</i>	<i>Podocyrtis ampla</i>	<i>Thyscyrtis tenuis</i>	<i>Theocytyle tenerulensis</i>	<i>Thyscyrtis hirsuta</i>	<i>Padocyrtis dimessa</i>				
early Miocene	<i>Stichocorys delmontensis</i>	1H-CC	X X X																																								
		2H-CC	X X X X X X																																								
		3H-CC	X X X X X X																																								
		4H-1, 90-92	X X X X X X	X X																																							
Oligocene	<i>Theocyrtis tuberosa</i>	5H-1, 90-92					X X	X X																																			
		5H-CC		X			X X X	X X																																			
		6H-CC			X		X X X X X X																																				
		7H-CC				X	X X																																				
		8H-CC					X X	X X																																			
		9H-1, 100-102					X X																																				
		9H-CC						X X	X X	X X X X X X	X X	X X X X X X	X X																														
late Eocene	<i>T. bromia</i>	10X-1, 55-56																																									
		10X-1, 100-102																																									
middle Eocene	<i>Podocyrtis mitra</i>	11X-1, 100-102																																									
		11X-CC																																									
		12X-CC																																									
	<i>P. ampla</i>	13X-4, 73-75																																									
		13X-CC																																									
		14X-1, 112-114																																									
		14X-2, 95-97																																									
	<i>Thyscyrtis triacantha</i>	14X-3, 148-150																																									
		14X-4, 129-131																																									
		14X-5, 69-71																																									
		14X-CC																																									
	<i>Dictyopora mongolfieri</i>	15X-2, 114-116																																									
		15X-3, 82-84																																									
		15X-CC																																									

downhole until they constitute the dominant recovered lithology. Also included in this section are Cores 143-869B-2R to -9R whose recovered lithology is dominated by porcellanite and chert, colored various shades of brown, locally containing radiolarians, and accompanied in some places by nannofossil limestone.

Some sections also contain several millimeter- to centimeter-thick levels of graded radiolarian sands that exhibit sharp contacts with the enveloping clayey radiolarian ooze. Radiolarian ooze, colored in various shades of yellowish-brown, becomes a progressively more important sediment type in Cores 143-869A-12X through -15X, where it contains subsidiary and variable amounts of clay and nannofossils. Those levels that contain abundant nannofossils are chalky in texture.

RESULTS

Abundant, well-preserved radiolarians were found in all samples from Hole 869A. A diverse assemblage that includes the age-diagnostic species *Calocyctella virginis*, *Liriospyris mutuaria*, *Stichocorys delmontensis*, *Didymocyrta prismatica*, *Dorcadospyrus ateuchus*, *Centrobotrys thermophila*, and *Lynchnocanoma elongata* was recovered in Samples 143-869A-1H-CC through -4H-1, 90-92 cm. The presence of these radiolarians indicates an early Miocene age, and the co-occurrence of *Stichocorys delmontensis* and *Dorcadospyrus ateuchus* permits assignment to the *Stichocorys delmontensis* Zone of Sanfilippo et al. (1985). Samples 143-869A-5H-1, 90-92 cm, through -9H-1, 100-102 cm, can be assigned an early Oligocene age, based on the occurrence of *Lithocyclia angusta*, *Tristylospyris triceros*, *Lynchodictyon audax*, *Artophormis gracilis*, *Centrobotrys petrushevskaya*, and *Theocyrtis tuberosa*. The presence of species such as *Centrobotrys petrushevskaya* and *Theocyrtis tuberosa* permits an age assignment to the *Theocyrtis tuberosa* Zone of Sanfilippo et al. (1985). The boundary between the *Stichocorys delmontensis* and *Theocyrtis tuberosa* zones was not precisely defined because Sample 143-869A-4H-3, 100-102 cm, was one of the few samples that did not contain well-preserved, abundant radiolarians.

No hiatus was detected at the Oligocene/Eocene boundary, which lies between Samples 143-869A-9H-1, 100-102 cm, and -9H-CC, with the latter containing a latest Eocene assemblage that includes *Thyscyrtis tetricantha*, *Thyscyrtis triacantha*, *Podocyrtis chalara*, *Lynchnocanoma bandycra*, *Eusyringium fistuligerum*, *Calocyclus turris*, and *Lithochytris vespertilio*. This assemblage was assigned to the *Thyscyrtis bromia* Zone of Sanfilippo et al. (1985). Reworking of middle Eocene radiolarians is evident in Samples 143-869A-10X-1, 55-56 cm, and -10X-1, 100-102 cm. These samples are latest Eocene in age, but contain elements of a middle Eocene fauna that includes *Podocyrtis mitra* and *Podocyrtis trachodes*, which are characteristic of the *Podocyrtis mitra* Zone of Sanfilippo et al. (1985), together with a latest Eocene fauna identical to that found in Sample 143-869A-9H-CC.

The remainder of the section is an apparently conformable middle Eocene succession. Sample 143-869A-13X-CC contains *Podocyrtis ampla* and *Podocyrtis sinuosa* and was assigned to the *Podocyrtis ampla* Zone of Sanfilippo et al. (1985). Samples 143-869A-14X-1, 112-114 cm, through -14X-CC contain *Thyscyrtis triacantha*, but not *Podocyrtis ampla*, suggesting that they belong to the *Thyscyrtis triacantha* Zone of Sanfilippo et al. (1985). *Thyscyrtis triacantha* is absent from Samples 143-869A-15X-2, 114-116 cm through 143-869A-15X-CC, but *Dictyopora mongolfieri* is present in great abundance together with *Thyscyrtis hirsuta*, indicating that these samples are from the middle Eocene *Dictyopora mongolfieri* Zone of Sanfilippo et al. (1985).

CONCLUSIONS

Exceptionally well-preserved, abundant middle Eocene to lower Miocene radiolarians in cores from Leg 143 Hole 869A permit refinement of shipboard age assignments on the basis of calcareous nannofossils. Radiolarian data indicate that the Eocene/Oligocene boundary sequence appears conformable, whereas significant erosional hiatuses are present between the early Oligocene and early Miocene and between the middle and late Eocene.

SPECIES LIST

Detailed original descriptions of the biostratigraphically significant radiolarian species, identified in samples from Leg 143, have already been presented. Therefore, the following list simply provides a bibliographic reference for the species mentioned here. In most cases, only the reference containing the original description is presented, except where this description differs from present consensus or has been revised. These species are listed in alphabetical order.

- Artophormis gracilis* Riedel
Artophormis gracilis Riedel, 1959, p. 300, pl. 2, figs. 12, 13.
Calocyclas bandycia (Mato and Theyer)
Lynchocanoma bandycia Mato and Theyer, 1980, p. 225, pl. 1, figs. 1–6.
Calocyclas bandycia (Mato and Theyer), Sanfilippo and Riedel in Saunders et al., 1985, p. 411, pl. 5, figs. 1, 5–6.
Calocyclas turris Ehrenberg
Calocyclas turris Ehrenberg, 1873, p. 218; 1875, pl. 18, fig. 7.
Calocyclas turris (Ehrenberg) Foreman, 1973, p. 434.
Calocyctella virginis Haeckel
Calocyctella virginis (Calocyctella) virginis Haeckel, 1887, p. 1381, pl. 74, fig. 4.
Calocyctella virginis (Haeckel) Riedel and Sanfilippo, 1970, p. 535, pl. 14, fig. 10.
Centrobotrys petrushevskayae Sanfilippo and Riedel
Centrobotrys petrushevskayae Sanfilippo and Riedel, 1973, p. 532, pl. 36, figs. 12, 13.
Centrobotrys thermophila Petrushevskaya
Centrobotrys thermophila Petrushevskaya, 1965, p. 115, text-fig. 20.
Centrobotrys thermophila (Petrushevskaya) Nigrini, 1967, p. 49, text-fig. 26, pl. 5, fig. 7.
Dictyopora mongolfieri (Ehrenberg)
Eucyrtidium mongolfieri Ehrenberg, 1854, pl. 36, fig. 18, Blower, 1873, p. 230.
Dictyopora mongolfieri (Ehrenberg) Nigrini, 1977, p. 250, pl. 4, fig. 10.
Didymocystis prismatica (Haeckel)
Pipettella prismatica Haeckel, 1887, p. 305, pl. 39, fig. 6.
Didymocystis prismatica (Haeckel), Sanfilippo and Riedel, 1980, p. 1010.
Dorcadospyris ateuchus (Ehrenberg)
Ceratospyris ateuchus Ehrenberg, 1873, p. 218; 1875, pl. 21, fig. 4D.
Dorcadospyris ateuchus (Ehrenberg), Riedel and Sanfilippo, 1970, p. 523, pl. 15, fig. 4.
Eusyringium fistuligerum (Ehrenberg)
Eucyrtidium fistuligerum Ehrenberg, 1873, p. 229; 1875, pl. 9, fig. 3.
Eusyringium fistuligerum (Ehrenberg) Riedel and Sanfilippo, 1970, p. 527, pl. 8, figs. 8, 9.
Eusyringium lagena (Ehrenberg)
Lithopera lagena Ehrenberg, 1873, p. 241; 1875, pl. 3, fig. 4.
Eusyringium lagena (Ehrenberg), Riedel and Sanfilippo, 1970, p. 527, pl. 8, figs. 5–7.
Liriospyris mutuaria Goll
Liriospyris mutuaria Goll, 1968, p. 1428, pl. 175, figs. 6, 10, 11 and 14, text-fig. 9.
Lithochytris vespertilio Ehrenberg
Lithochytris vespertilio Ehrenberg, 1873, p. 239.
Lithocyclia angusta (Riedel)
Trigonactura angusta Riedel, 1959, p. 292, pl. 1, fig. 6.
Lithocyclia angusta (Riedel), Riedel and Sanfilippo, 1970, p. 522, pl. 13, figs. 1, 2.
Lithocyclia ocellus Ehrenberg
Lithocyclia ocellus Ehrenberg, 1854, pl. 36, fig. 30; 1873, p. 240.
Lithocyclia ocellus (Ehrenberg), Riedel and Sanfilippo, 1970, p. 522, pl. 5, figs. 1, 2.
Lychnodictyon audax Riedel
Lychnodictyon audax Riedel, 1953, p. 810, pl. 85, fig. 9.
Lychnocanoma elongata (Vinassa de Regny)
Tetrahedrina elongata Vinassa de Regny, 1900, p. 243, pl. 2, fig. 31.
Lychnocanoma bipes Riedel, 1959, p. 294, pl. 2, figs. 5–6.
Lychnocanoma elongata (Vinassa de Regny), Sanfilippo et al., 1973, p. 221, 222, pl. 5, figs. 19–20.
Phormocystis striata striata Brandt
Phormocystis striata Brandt in Wetzel, 1935, p. 55, pl. 9, fig. 12.
Phormocystis striata striata (Brandt) Foreman, 1973, p. 438, pl. 7, figs. 5, 6, 9.
Podocystis (Podocystotges) ampla Ehrenberg
Podocystis (?) ampla Ehrenberg, 1873, p. 248, 1875, pl. 16, fig. 7.
Podocystis ampla (Ehrenberg) Riedel and Sanfilippo, 1970, p. 533, pl. 12, figs. 5, 7, 8.
Podocystis (Podocystotges) ampla (Ehrenberg) Sanfilippo and Riedel, 1992, p. 14, pl. 5, fig. 4.
Podocystis (Lampterium) chalara Riedel and Sanfilippo
Podocystis (Lampterium) chalara Riedel and Sanfilippo, 1970, p. 535, pl. 12, figs. 2, 3.
Podocystis (Lampterium) chalara Riedel and Sanfilippo, 1978, p. 71, pl. 8, fig. 3, text fig. 3.
Podocystis (Podocystotges) diamesa Riedel and Sanfilippo
Podocystis (Podocystotges) diamesa Riedel and Sanfilippo, 1970, p. 533 (partim), pl. 12, fig. 4, non figs. 5, 6.
Podocystis (Podocystotges) diamesa Sanfilippo and Riedel 1973, p. 531, pl. 20, figs. 9, 10, pl. 35, figs. 10, 11.
Podocystis (Podocystotges) diamesa (Riedel and Sanfilippo) Sanfilippo and Riedel, 1992, p. 14.
Podocystis (Lampterium) fasciolata (Nigrini)
Podocystis (Podocystotges) ampla fasciolata Nigrini, 1974, p. 1069, pl. 1K, figs. 1, 2, pl. 4, figs. 2, 3.
Podocystis (Lampterium) fasciolata (Nigrini) Sanfilippo et al., 1985, p. 697, fig. 30.7.
Podocystis (Lampterium) helenae Nigrini
Podocystis (Lampterium) helenae Nigrini, 1974, p. 1070, pl. 1L, figs. 9–11, pl. 4, figs. 4, 5.
Podocystis (Lampterium) mitra Ehrenberg
Podocystis (Lampterium) mitra Ehrenberg, 1854, pl. 36, fig. B20: 1873, p. 251; non Ehrenberg, 1875, pl. 15, fig. 4.
Podocystis (Lampterium) mitra (Ehrenberg) Riedel and Sanfilippo, 1970, p. 534; 1978, text-fig. 3.
Podocystis (Lampterium) sinuosa Ehrenberg
Podocystis (Lampterium) sinuosa Ehrenberg, 1873, p. 253; 1875, pl. 15, fig. 5; Riedel and Sanfilippo, 1970, p. 534, pl. 11, fig. 3; 1978, text-fig. 3.
Podocystis (Lampterium) trachodes Riedel and Sanfilippo
Podocystis (Lampterium) trachodes Riedel and Sanfilippo 1970, p. 535, pl. 11, fig. 7, pl. 12, fig. 1.
Sethochytris triconiscus Haeckel
Sethochytris triconiscus Haeckel, 1887, p. 1239, pl. 57, fig. 13.
Spongatractus pachystylus (Ehrenberg)
Spongosphera pachystyla Ehrenberg, 1873, p. 256; 1875, pl. 26, fig. 3.
Spongatractus pachystylus (Ehrenberg) Sanfilippo and Riedel, 1973, p. 519, pl. 1, 2, figs. 4–6, pl. 25, fig. 3.
Stichocorys delmontensis (Campbell and Clark)
Eucyrtidium delmontensis, Campbell and Clark, 1944, p. 56, pl. 7, figs. 19, 20.
Stichocorys delmontensis (Campbell and Clark) Sanfilippo and Riedel, 1970, p. 451, pl. 1, fig. 9.
Theocystole venezuelensis Riedel and Sanfilippo
Theocystole venezuelensis Riedel and Sanfilippo, 1970, 525, pl. 6, figs. 9, 10, pl. 7, figs. 1, 2.
Theocystis tuberosa Riedel emend. Sanfilippo et al.
Theocystis tuberosa Riedel, 1959, p. 298, pl. 2, figs. 10, 11; Sanfilippo et al., 1985, p. 701, figs. 32, 1a–1d.
Thrysocystis (Thrysocystis) hirsuta (Krasheninnikov)
Podocystis hirsutus Krasheninnikov, 1960, p. 300, pl. 3, fig. 16.
Thrysocystis (Thrysocystis) hirsuta (Krasheninnikov) Sanfilippo and Riedel, 1982, p. 173, pl. 1, figs. 3, 4.
Thrysocystis (Thrysocystis) rhizodon Ehrenberg
Thrysocystis rhizodon Ehrenberg, 1873, p. 262; 1875, p. 94, pl. 12, fig. 1; Sanfilippo and Riedel, 1982, p. 173, pl. 1, figs. 14–16, pl. 3, figs. 12–17.
Thrysocystis (Thrysocystis) robusta Riedel and Sanfilippo
Thrysocystis hirsuta robusta Riedel and Sanfilippo, 1970, p. 526, pl. 8, fig. 1.
Thrysocystis (Thrysocystis) robusta (Riedel and Sanfilippo) Sanfilippo and Riedel, 1982, p. 174, pl. 1, fig. 5.
Thrysocystis (Pentalacorys) tensa Foreman
Thrysocystis hirsuta tensa Foreman, 1973, p. 442, pl. 3, figs. 13–16, pl. 12, fig. 8.
Thrysocystis (Pentalacorys) tetricantha (Ehrenberg)
Podocystis tetricantha Ehrenberg, 1873, p. 254; 1875, pl. 13, fig. 2.
Thrysocystis (Pentalacorys) tetricantha (Ehrenberg) Sanfilippo and Riedel, 1982, p. 176, pl. 1, figs. 11, 12, pl. 3, fig. 10.
Thrysocystis (Pentalacorys) triacantha (Ehrenberg)
Podocystis triacantha Ehrenberg, 1873, p. 254; 1875, pl. 13, fig. 4.
Thrysocystis (Pentalacorys) triacantha (Ehrenberg) Sanfilippo and Riedel, 1982, p. 176, pl. 1, figs. 8–10, pl. 3, figs. 3, 4.

Tristylospyris triceros (Ehrenberg)

Ceratospyris triceros Ehrenberg, 1873, p. 220; 1875, pl. 21, fig. 5.
Tristylospyris triceros (Ehrenberg) Haeckel, 1887, p. 1033.

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