

## 5. EOCENE TO MIOCENE OCEANOGRAPHIC AND PROVENANCE CHANGES IN A SEQUENCE STRATIGRAPHIC FRAMEWORK: BENTHIC FORAMINIFERS OF THE NEW JERSEY MARGIN<sup>1</sup>

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### ABSTRACT

Drilling on the New Jersey continental slope and rise provides an Eocene bathyal to abyssal depth transect and an Oligocene to Miocene bathyal transect that allow the calibration of benthic foraminiferal abundance changes to independent depth estimates through time. New Jersey Transect benthic foraminiferal faunal changes reflect global, regional, and local influences and provide constraints on paleobathymetry and sediment provenance.

The New Jersey Transect yields cosmopolitan Eocene deep-water benthic foraminiferal faunas. Early to middle Eocene bathyal biofacies are characterized by *Lenticulina* spp., *Alabamina wilcoxensis*, and *Osangularia* spp., whereas coeval abyssal locations contain a deeper *Nuttallides truempyi*-dominated biofacies. A late middle Eocene shift to a bathyal biofacies dominated by *Osangularia* spp., *Bulimina alazanensis*, and *Pullenia bulloides* reflects a global event that affected bathyal and abyssal sites. However, we lack an abyssal record to evaluate the full regional impact of this global event. Late Eocene bathyal faunas on the New Jersey Transect contain high abundances of *Bulimina alazanensis* (up to 50%), similar to peak *B. alazanensis* abundances reported from other Atlantic locations. We speculate that this uniform biofacies indicates that circum-Atlantic and Gulf of Mexico sites were ventilated by similar intermediate to upper deep-water masses in the late Eocene.

A cosmopolitan *Lenticulina* spp.-dominated biofacies prevailed in the Oligocene to early Miocene bathyal zone; polymorphinids and *A. wilcoxensis* were common secondary components of this biofacies. A *Uvigerina hispida*-*B. alazanensis*-dominated biofacies characterized the middle Miocene. A coeval increase in *Uvigerina* spp.-dominated faunas occurred in the Gulf of Mexico, indicating that this increase may be regional in extent. The shift from the *Lenticulina* spp.-dominated biofacies to the *Uvigerina hispida*-*B. alazanensis*-dominated biofacies may be the regional expression of a global late early to early middle Miocene benthic foraminiferal turnover event and associated abundance changes.

Oligocene to Miocene benthic foraminiferal biofacies changes at the slope sites exhibit sequence stratigraphic variations: (1) downslope-transported shelf taxa above several sequence boundaries (this may represent the slope expression of the shelf lowstand systems tracts) and in the upper parts of several sequences (within the slope equivalent of the shelf highstand systems tracts); (2) a preponderance of *U. proboscidea* occurs in the lower parts of several sequences (this may represent in situ biofacies within the slope expression of the lowstand systems tracts); (3) higher abundances of *Stilostomella* spp. occur in the upper parts of several sequences (this may represent the slope equivalent of the highstand systems tracts); and (4) and higher abundances of *M. pompilioides* span several sequence boundaries. We tentatively interpret the sections immediately above sequence boundaries as correlative to the lowstand systems tracts on the shelf. These sections contain predominantly in situ faunas with minor abundances of transported benthic foraminifera (<10% of the total fauna). Minor abundances of transported benthic foraminifera reappear in the upper parts of several sequences, and may reflect minor shedding of shelf sediments during highstand systems tracts progradation on the shelf.

Sedimentation in the early middle Miocene and older intervals was dominated by pelagic settling at our slope sites (as it is today), and there is little benthic foraminiferal evidence for widespread downslope transport prior to the middle middle Miocene at our sites. In the Leg 150 region of the New Jersey slope, a dramatic change in the dominant mode of sedimentation occurred between Reflectors m3 (13.5 Ma) and m2 (12.5 Ma). This change is indicated by (1) a major increase in downslope transport of neritic (shelf) species; (2) dramatically increased sedimentation rates (>60 cm/yr); (3) increased terrigenous input; (4) submarine channel cutting; (5) high terrestrial carbon input; and (6) mobilization of biogenic carbonate to form diagenetic nodules. Transported benthic foraminifera dominate the upper middle Miocene to Pleistocene section of the slope cores, reflecting increased sediment supply due to the progradation of clinoforms that progressed from 90 km landward of Site 903 in the early Miocene to ~60 km landward in the middle Miocene to less than 20 km landward in the Pleistocene.

### INTRODUCTION

One of the primary goals of Ocean Drilling Program (ODP) Leg 150 was to evaluate the nature of slope sedimentation and its response to global sea-level changes (Miller and Mountain, 1994). This evaluation involves reconstructing sediment provenance changes and changes in dominant depositional processes (e.g., gravity flows vs. hemipelagic settling) on both long-term (10 m.y.) and shorter term

(~1 m.y.) scales. Although changes in provenance are often evaluated using sedimentological criteria (e.g., Mazzullo, 1986), benthic foraminifera also provide reliable tracers of neritic sediment sources for continental slope deposits (Brunner and Culver, 1992). For example, benthic foraminifera can be especially useful in sorting out the origins of multiple-source materials on the lower slope and upper rise (e.g., McHugh et al., this volume).

Previous foraminiferal studies demonstrate that the dominant pattern of sedimentation on the New Jersey continental slope today is pelagic settling (Miller and Lohmann, 1982; Streeter and Lavery, 1982; Christensen and Miller, 1991) with minimal evidence for downslope transport processes. Observations made from submersibles also indicate that most of the middle slope is draped with (hemi-)pelagic muddy oozes in the Carteret Canyon area (K.G. Miller, pers. comm., 1989). In contrast, widespread erosion characterized

<sup>1</sup>Mountain, G.S., Miller, K.G., Blum, P., Poag, C.W., and Twichell, D.C. (Eds.), 1996. *Proc. ODP, Sci. Results*, 150: College Station, TX (Ocean Drilling Program).

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the New Jersey slope in the past (Poag, Watts, et al., 1987); benthic foraminiferal studies show the downcore presence of transported material on the floors and southwestern walls of Lindenkohl and Carteret Canyons and on the crests of their southwestern walls (interfluvial) (Christensen and Miller, 1991). Leg 150 drilling established that intervals of pervasive downslope transport were concentrated in Pleistocene glacial periods, although transported benthic foraminifera also dominated interglacial periods (Christensen et al., this volume). One of the topics addressed by Leg 150 is whether pre-Pleistocene intervals on the New Jersey slope were characterized by tranquil pelagic settling or dominated by downslope transport.

Benthic foraminifera also can be used to evaluate changes in source and depositional environments within sequences (i.e., stratigraphic sections bracketed by unconformities). Changes in neritic benthic foraminiferal biofacies have been used successfully to decipher sequences deposited on continental shelves (e.g., Miller et al., 1994; Olsson et al., 1987; Christensen et al., in press). However, no benthic foraminiferal biofacies model for slope sequences has been published, and it is unclear whether benthic foraminifera exhibit a predictable succession correlative to the classic systems tracts model (Posamentier et al., 1988). For example, are there (1) "lowstand systems tract" biofacies on the slope that are characterized by transported benthic foraminiferal biofacies; (2) "transgressive systems tract" biofacies characterized by benthic foraminifera more typical of pelagic settings; and/or (3) "highstand systems tract" biofacies characterized by benthic foraminifera typical of regions with high supply of terrigenous sediments? A sequence stratigraphic framework for the New Jersey slope was developed by tracing seismic reflectors from the shelf to four Leg 150 boreholes (e.g., Miller and Mountain, 1994), providing the opportunity to evaluate intrasequence and longer term changes on the slope using the benthic foraminiferal record.

Drilling on the New Jersey continental slope and rise by ODP Leg 150 (Sites 902, 903, 904, and 906; 811, 444, 1123, and 913 m water depth, respectively), Deep Sea Drilling Project (DSDP) Leg 95 (Sites 612 and 613, 1404 and 2323 m water depth, respectively; Poag, Watts, et al., 1987), and DSDP Leg 11 (Site 108, 1815 m water depth; Hollister, Ewing, et al., 1972), provides an Eocene bathyal to abyssal transect and an Oligocene to Miocene bathyal transect (Fig. 1; Table 1; see Miller and Katz 1987a, for locations of Sites 108, 390, and 613). Site 390 (DSDP Leg 44, 2665 m water depth; Benson, Sheridan, et al., 1978), drilled on the Blake Nose, is included in this study because it is an abyssal location in the same region as the New Jersey Transect. Similar depth transects have proven valuable for reconstructing age-depth changes through time (e.g., Katz and Miller, 1993). This transect approach provides an opportunity to evaluate Eocene to Miocene benthic foraminiferal abundance changes through depth and time on the New Jersey slope by calibrating them to independent depth estimates obtained from backtracking (Berger and Winterer, 1974) and backstripping (Steckler and Watts, 1982). In addition, the New Jersey Transect shows benthic foraminiferal faunal changes that reflect global, regional, and local influences and provide constraints on sediment provenance.

## METHODS

### Benthic Foraminifera

Samples were washed with sodium metaphosphate (5.5 g/L) in tap water through a 63- $\mu$ m sieve and air-dried. Benthic foraminifera were picked from aliquots of the greater than 150- $\mu$ m size fraction and mounted on reference slides. We chose this fraction to be consistent with previous studies on this margin (e.g., Streeter and Lavery, 1981; Miller and Katz, 1987a) and the deep sea (e.g., Thomas, 1990). We avoid the 63–150- $\mu$ m size fraction preferred by some authors (e.g., Schröder et al., 1987) because of problems in identifying small taxa. While we lose information obtainable from the 63–150- $\mu$ m size

fraction, we gain information on larger taxa (i.e., in an aliquot picked from the >63- $\mu$ m size fraction, many larger taxa are underemphasized while small, difficult to identify taxa are emphasized).

In general, 300–400 specimens were picked per sample. However, samples with as few as 85 specimens were used in the faunal analyses; samples that yielded too few specimens were omitted from analyses. Three hundred and nineteen samples were examined for this study (Table 1). Closely spaced samples (2-m sampling interval) from the m5 sequence at Sites 903 and 904 that were examined for intrasequence biofacies changes were not included in the multivariate analyses.

Stratigraphic control was obtained from planktonic foraminifera (Snyder et al., this volume), nannofossils (Aubry in Mountain, Miller, Blum, et al., 1994), and strontium isotope stratigraphy (Miller et al., this volume). Sample ages were assigned by interpolating between planktonic biostratigraphic datums and Sr isotope ages and by extrapolating where necessary (Table 2). The time scale of Berggren et al. (1985) was used to be consistent with shipboard studies and Mountain, Miller, Blum, et al. (1994). Cande and Kent (1992) made substantial revisions (>2 m.y. in some cases) to much of the Paleogene geomagnetic polarity time scale (GPTS). The boundary calibrations of chronostratigraphic units, zones, and isotopic correlations have only recently been calibrated to the GPTS (Berggren et al., 1995); in order to maintain consistency in view of a rapidly evolving time scale, Leg 150 participants decided to report ages based on the Berggren et al. (1985) geological time scale. Although the numerical ages presented here will change with the Berggren et al. (1995) time scale, the relative faunal patterns will remain the same.

Our identifications of most of the benthic foraminifera follow the taxonomy of van Morkhoven et al. (1986) and Tjalsma and Lohmann (1983). Our taxonomic base for most of the uvigerinids follows Boersma (1984). Not all species present were identified; for example, species of some common (*Lenticulina*, *Oridorsalis*, and *Gyroidinoides*) and some rare (*Lagena*, *Fissurina*, and *Oolina*) genera were not differentiated. We revised the taxonomy of Miller and Katz (1987a) for Sites 108, 390, 612, and 613 to be consistent with the taxonomy of the Leg 150 samples (see Appendix).

We performed Q-mode principal components and varimax factor analyses on the relative abundance (percentage) data using modifications of programs provided by Lohmann (1980). The Q-mode principal components and varimax factor programs utilize a cosine-theta matrix, standardizing each sample to unit length. Each factor represents a unique benthic foraminiferal biofacies. Factor analysis of the New Jersey Margin samples reveals stratigraphically meaningful biofacies successions. We used factor analysis to identify trends in the benthic foraminiferal biofacies at Sites 902, 903, 904, and 906. We analyzed each site individually and then combined the data from Sites 902, 903, 904, and 906 with Sites 108, 390, 612, and 613 to form two data sets, the Eocene and the Oligocene to Miocene, to emphasize depth rather than age differences. However, there is virtually no Oligocene to middle Miocene section at the deeper Sites 108, 390, 612, and 613; therefore, the lowermost bathyal to abyssal depths are represented only for the Eocene. The Eocene data set consists of 96 samples and 114 taxa, and the Oligocene to Miocene data set consists of 102 samples and 104 taxa. We limited our data sets used in the factor analyses to samples that yielded primarily in situ biofacies.

We use "bubble plots" to display three-dimensional data (age-paleodepth-species data/factor analysis results) in two dimensions (e.g., Tjalsma and Lohmann, 1983; Katz and Miller, 1993). They are well suited for displaying (1) long-term (m.y.) trends in species data that often contain high-frequency signals (e.g., Milankovitch scale); and (2) data that have uncertainties in age (typically  $\pm 0.5$  m.y.) or paleodepth (typically  $\pm$  several hundred meters). Bubble plots clearly show how benthic foraminiferal species abundances and biofacies have changed through time across a paleobathymetric range. Bubble size is proportional to each factor loading or species percentage.

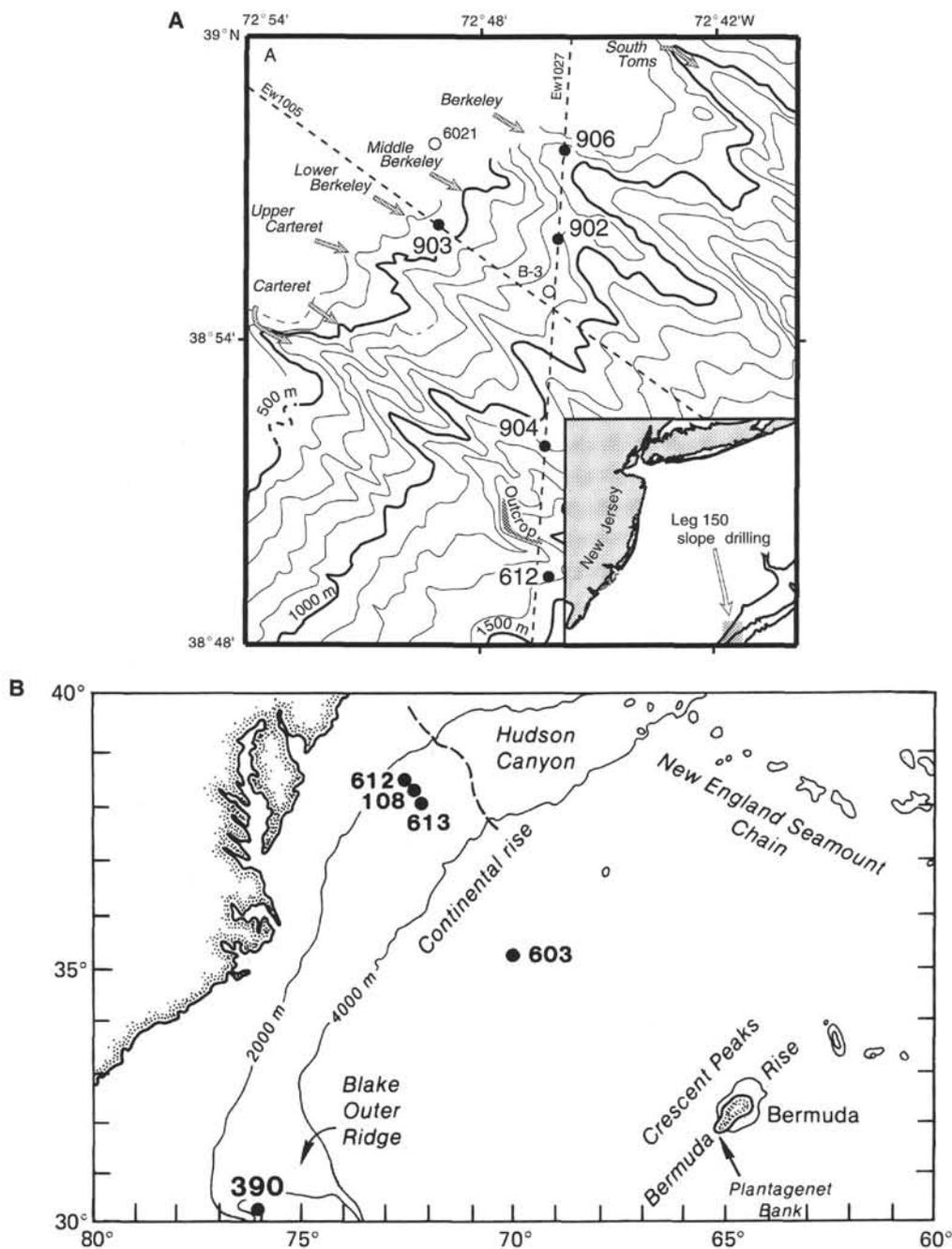


Figure 1. **A.** SeaBeam bathymetric location map of the middle continental slope (500–1500 m water depth) drilled by Leg 150 showing ODP Sites 902, 903, 904, and 906, and DSDP Site 612. SeaBeam data were collected by *Atlantis II* 120 and 124 and processed by W.B.F. Ryan and D. Twichell, unpubl. data, 1989 (after Mountain, Miller, Blum, et al., 1994). Ew1005 and Ew1027 are multichannel seismic profiles illustrated in Mountain, Miller, Blum, et al. (1994). **B.** Location map showing DSDP Sites 108, 390, 612, and 613 (after Miller and Katz, 1987a).

### Paleobathymetry

Benthic foraminifers have been recognized for their usefulness in assessing paleobathymetry (e.g., Natland, 1933; Bandy, 1960), although it is clear that factors other than water depth control the distributions. For example, deep-sea benthic foraminiferal distributions are often associated with water mass (e.g., Streeter, 1973; Bremer and Lohmann, 1982), productivity (e.g., Thomas et al., 1995), or substrate (e.g., Miller and Lohmann, 1982), characteristics that may vary independently of water depth. Independent estimates of paleobathymetry (backtracking of Berger and Winterer, 1974; backstrip-

ping of Steckler and Watts, 1982) provide calibrations for benthic foraminiferal abundance changes and species ranges through time across paleodepths (e.g., Tjalsma and Lohmann, 1983). Once faunal trends are calibrated using independent paleodepth estimates, benthic foraminiferal distributions can be used to evaluate paleodepths for sections lacking independent depth estimates.

The Eocene to Holocene sections at Sites 108, 612, 613, 902, 903, 904, and 906 overlie very thick (>10 km) Jurassic–Holocene sediments and thinned (transitional) continental to oceanic crust (Grow and Sheridan, 1988). Thus, they cannot be rigorously backtracked using conventional age-subsidence curves (e.g., Berger and Winterer,

**Table 1. DSDP and ODP sites included in this study.**

Site	Location	Present water depth (m)	Number of samples	No. of samples in analyses	No. of taxa in individual analyses	Backstripped paleodepth (m)	Backtracked paleodepth (m)
108	38°48.27'N, 72°39.21'W	1815	2	2	n.a.	1680	1650
390	30°08.54'N, 76°06.74'W	2665	5	5	n.a.		2365–2425
612	38°49.21'N, 72°46.43'W	1404	31	31	n.a.	1280–1305	1320–1430
613	38°46.26'N, 72°30.43'W	2323	11	7	n.a.	2200–2215	2275–2310
902	38°56.08'N, 72°46.38'W	811	41	41	108	790–830	1050–1070
903	38°56.30'N, 72°49.03'W	444	118	34	132	470–500	790–935
904	38°51.81'N, 72°46.08'W	1123	68	45	141	1025–1120	1155–1215
906	38°57.90'N, 72°46.00'W	913	43	31	109	870–930	1075–1135

Note: n.a. = not applicable.

1974), although we previously backtracked Sites 108, 612, and 613 (Miller and Katz, 1987a). We followed Miller and Katz (1987a) in computing a first-order paleodepth estimate by accounting for Airy loading and simple thermal subsidence in the “backtracking” form:

$$\text{Paleodepth} = \text{Initial depth} - A + A \cdot e^{(-t/\tau)} - S,$$

where  $t$  = basement age – age level considered;  $\tau$  = empirical decay constant, 62.5;  $A$  = empirical scaling constant, –3650; and  $S$  = the Airy correction,  $0.66 \cdot (\text{basement depth} - \text{depth of level considered})$  (see Miller and Katz, 1987a).

We backtracked paleodepths assuming crustal ages of (1) 180 Ma and 13 km of sediments for Sites 902, 903, 904, 906 (Mountain, Miller, Blum, et al., 1994) and 612 (Poag, 1985); (2) 180 Ma and 10 km of sediments for Site 613 (Poag, 1985); (3) 180 Ma and 11 km of sediments for Site 108 (Poag, 1985); and (4) 160 Ma and 380 m of sediments for Site 390 (Benson, Sheridan, et al., 1978).

M. Steckler (pers. comm., 1995) rigorously “backstripped” Sites 108, 612, 613, 902, 903, 904, and 906, which lie on transitional continental crust. He used the method of Steckler et al. (1988), correcting for the effects of compaction, loading, and thermal subsidence, and the long-term sea-level change of Kominz (1984). M. Steckler (pers. comm., 1995) assumed an age of 185 Ma for the crust with 12 km of sediment, and he used Airy isostasy. The differences between the backtracked and backstripped paleodepths primarily reflect the effects of decompaction taken into account in the backstripping method (Table 1). The backstripped paleodepth estimates were generally less than 150 m shallower than the backtracked paleodepth estimates for the deeper water sites (108, 613, 612, and 904). The shallower sites have thicker Pleistocene sections that need to be removed via backstripping in order to calculate reliable paleodepths. The effects of the thick Pleistocene sections are apparent in the greater differences between the backtracked and backstripped paleodepths at the shallower sites (Table 1). In this study, we use the backstripped paleodepths for all sites (except for Site 390, for which we used the backtracking method, which is applicable to normal oceanic crust with thin sediments) to evaluate benthic foraminiferal faunal trends through depth and time (Table 3).

We use the following bathymetric zones (e.g., van Morkhoven et al., 1986; Berggren and Miller, 1989): bathyal = 200–2000 m (upper: 200–600 m; middle: 600–1000 m; lower: 1000–2000 m); and abyssal = >2000 m (upper: 2000–3000 m; lower: >3000 m).

### Sequence Stratigraphic Framework

In order to evaluate benthic foraminiferal changes within sequences, the sequence boundaries must be located precisely. Sequence boundaries were located initially using seismic profiles (Miller and Mountain, 1994) that were correlated to the slope boreholes

drilled by Leg 150 (summarized in Mountain, Miller, Blum, et al., 1994; see their table 16, Site 902; table 13, Site 903; table 12, Site 904; and table 11, Site 906). Most of the Oligocene and Miocene reflectors can be traced from the shelf, where reflector terminations indicate that they are seismic sequence boundaries (Greenlee et al., 1992). In addition, most of these reflectors correspond to lithologic changes, downhole log changes (Mountain, Miller, Blum, et al., 1994), impedance contrasts measured in cores (Mountain, Miller, Blum, et al., 1994; Lorenzo, this volume), and/or stratigraphic gaps detected in the slope boreholes (Miller et al., this volume), and are thus recognized as unconformities (sequence boundaries) on the slope. A few reflectors that do not correlate with obvious unconformities appear to correspond to “correlative conformities” (i.e., indiscernible hiatuses; e.g., Reflector m6 at Site 904) or the tops of sands that overlie unconformities (e.g., Reflector m1 at Site 903).

We used the depths for the reflectors in the boreholes given by Mountain, Miller, Blum, et al. (1994) except for minor changes<sup>4</sup> made by G.S. Mountain (pers. comm., 1995) and our own adjustments to Reflectors m3 to m5 at Sites 903 and 904. Our detailed sampling for benthic foraminiferal changes across the m3–m5 sequence boundaries required the reevaluation of the precise reflector levels. We evaluated their correlations based on additional core impedance contrast data (Lorenzo and Hesselbo, this volume), reexamination of the lithologic descriptions, Sr isotopic data (Miller et al., this volume), and our benthic foraminiferal changes. The following changes are within the uncertainties of seismic-borehole correlations:

1. Reflector m3/4 (combined) at Site 904 could be placed at ~244 mbsf at the base of a sandy silt that is bracketed by Sr isotope ages of 13.0 (238.4 mbsf) and 16.0 Ma (245 mbsf); this differs from the shipboard placement of 235 mbsf.

2. Reflector m5 at Site 904 is bracketed by Sr isotopic age estimates of 16.9 (257.39 mbsf) and 18.2 Ma (264.20 mbsf); we tentatively place it near the lower analysis from near the base of a sandy silt; this reflector was tentatively identified by shipboard studies at 272 mbsf or above.

3. Reflector m4 at Site 903 could be placed near 850 mbsf (as it was on the ship), associated with a possible short hiatus indicated by Sr isotope ages of 13.5 (836.41 mbsf depth at Hole 903D; 828.41 mbsf adjusted to Hole 903C depth) and 15.7 Ma (852.49 mbsf at Hole 903D; 846.49 mbsf adjusted to Hole 903C). However, note that Sr isotopes lack the resolution to define a hiatus this brief in this interval and that there is evidence that the reflector could be deeper. We prefer to place it at the base of a minor lithologic color change at 862 mbsf (Hole 903D depth; 854 mbsf at Hole 903C), near the level of an

<sup>4</sup>These changes include: Site 904: m1, 180.3 mbsf; m2, 220 mbsf; m6, 310 mbsf; Site 903: m3, 796 mbsf; Site 902: m6, 612 mbsf.

**Table 2. Age model parameters (LO = lowest occurrence, HO = highest occurrence, Sr = strontium isotope age).**

Depth (mbsf)	Age (m.y.)	Datum
<b>Site 902:</b>		
595.52	22.04	Sr
609.81	23.16	Sr
616.47	26.60	Sr
677.04	30.20	Sr
680.94	37.00	Arbitrary level within late NP19–20
728.54	38.00	Within NP19/20
<b>Site 903:</b>		
558.82	10.40	HO <i>P. mayeri</i> ?
756.50	12.83	Top C5AA
788.00	13.01	Base C5AA
903.72	16.30	Sr
910.50	18.20	Sr
1000.00	22.17	Within NN2
1003.00	24.00	Sr
1064.10	32.00	Extrapolated sedimentation rate from below
1064.13	37.00	Upper NP19–20
1112.40	38.50	LO <i>I. recurvus</i>
1113.00	40.00	Base NP18
1121.00	40.60	HO <i>Acarinina</i> spp.
1136.00	42.00	Top NP16
<b>Site 904:</b>		
176.50	8.00	Arbitrary level in C4n
182.00	11.47	Extrapolated sedimentation rate from below
184.00	11.55	Top C5An
198.50	12.12	Top C5Ar
226.50	12.83	Top C5AA
243.50	13.30	Level of unconformity
245.00	16.00	Sr
257.42	16.90	Sr
264.22	18.10	Sr
305.90	23.70	LO <i>G. kugleri</i>
334.30	28.20	Sr
341.21	37.00	Arbitrary level within late NP19–20
417.92	38.50	Microtektite layer
417.93	44.30	Interpolated from below
428.00	44.70	LO <i>T. pomeroli</i>
463.00	46.00	LO <i>M. lehneri</i>
480.73	47.00	HO <i>C. gigas</i>
548.47	48.80	LO <i>C. gigas</i>
565.28	52.00	NP14a; HO <i>S. sublodensis</i>
<b>Site 906:</b>		
279.20	12.50	Correlated with m2
478.199	13.50	Base of Miocene
478.20	27.00	Top of Oligocene
555.50	30.40	Base of Oligocene
563.80	37.00	Top of Eocene
599.20	38.00	Arbitrary level within early NP19–20
<b>Site 612:</b>		
136.20	36.60	HO <i>Hantkenina</i> spp.
181.39	38.50	Microtektite layer
183.00	43.00	Within range of <i>A. bullbrooki</i>
331.70	51.00	Arbitrary level within Zone P10
337.50	52.00	HO <i>M. caucasica</i>
530.60	55.20	LO <i>M. aragonensis</i>
<b>Site 613:</b>		
269.00	43.00	Within range of <i>A. bullbrooki</i>
336.00	46.00	HO <i>M. aragonensis</i>
430.00	50.20	LO <i>T. rohri</i>
440.00	52.00	HO <i>M. caucasica</i>
578.00	55.20	LO <i>M. caucasica</i>
<b>Site 390:</b>		
8.70	44.00	Arbitrary level in P12
31.05	45.90	HO <i>M. aragonensis</i>
45.00	52.00	LO <i>Hantkenina</i> spp.
70.84	55.20	LO <i>M. aragonensis</i>

increase in core impedance contrast at 860 mbsf (Hole 903D depth; 852 mbsf at Hole 903C depth). However, we indicate an interval from 852 to 862 mbsf on the Site 903 figures to reflect the uncertainty in the placement of Reflector m3.

4. Reflector m5 at Site 903 is bracketed by Sr isotopic age estimates of 16.2 Ma (903.71 mbsf at Hole 903D; 895.71 m at Hole 903C) and 18.2 Ma (910.20 mbsf at Hole 903D; 902.20 mbsf at Hole 903C), indicating correlation similar to the shipboard placement at 900 mbsf if depths are corrected to an 8-m deeper offset at Hole

903D, or 908 mbsf using Hole 903D depths (samples are plotted using Hole 903D depths in this study). Lithologic studies show the base of a glauconite unit at 908.4 m at Hole 903D (900.4 mbsf at Hole 903C), and this is where we place the reflector.

We used factor analysis, species percentages, and number of benthic foraminifers per gram to identify patterns associated with the m5 sequence at Sites 903 and 904. Factor analysis did not provide results that were stratigraphically meaningful, but helped to delineate the dominant taxa within the sections. We plotted species percentages based on the factor analysis results, and identified several species abundance changes that appear to be related to sequence stratigraphy.

## RESULTS AND DISCUSSION

### Leg 150 Biofacies

#### Site 904

At Site 904 (1123 m present water depth; ~1020–1120 m paleodepth), five factors explain 58% of the faunal variation (45 samples and 141 taxa). The lower to middle Eocene biofacies (Factor 2) is characterized by *Lenticulina* spp. and *Alabamina wilcoxensis*, with secondary faunal components (Figs. 2, 3). No biofacies change is associated with the disconformable lower/middle Eocene boundary or with the transition from porcellanitic chalks to chalks at ~525 mbsf (= Reflector e2, Fig. 3; equivalent to Horizon A<sup>c</sup> of Tucholke and Mountain, 1979). The *Lenticulina* spp.–*Alabamina wilcoxensis* biofacies was replaced by an upper Eocene biofacies characterized by *Pullenia bulloides*, *Osangularia* spp., polymorphinids, and *Gyroidinoides* spp. (Factor 3) and *Bulimina alazanensis* (Factor 1) (Figs. 2, 3). Factor 1 is dominated by *B. alazanensis* and has peaks in the upper Eocene and middle Miocene. A major faunal abundance change occurs across the disconformable Eocene/upper Oligocene contact from Factors 3 and 1 below to Factor 5 above; this is most likely the result of the nearly 10-m.y. hiatus at this contact (Mountain, Miller, Blum, et al., 1994).

The upper Oligocene to lower middle Miocene section at Site 904 contains an in situ bathyal benthic foraminiferal biofacies represented by Factor 4 (characterized by *Stilostomella* spp., *Uvigerina hispida*, *Cibicidoides pachyderma*, *Bulimina gracilis*, *Bulimina macilenta*, *Martinottiella communis*, and *Melonis barleeianum*) and Factor 5 (characterized by *Uvigerina proboscidea*, *Stilostomella modesta*, *Plectofrondicularia* spp., *M. barleeianum*, *Lenticulina* spp., and *Stilostomella* spp.) (Figs. 2, 3). Within this section, the Factor 5 biofacies characterizes the intervals immediately above Reflectors o1, m6, and m5, whereas the Factor 4 biofacies characterizes the upper parts of the sequences. This is consistent with our findings in the high-resolution intrasequence study (see below), where *U. proboscidea* percentages are high above reflectors (i.e., in the lower part of the overlying sequence) and *Stilostomella* spp. percentages are high below reflectors (i.e., in the upper part of the underlying sequence). A similar pattern is associated with Reflectors m3 and m2, with Factor 1 biofacies (*B. alazanensis*) occurring immediately above the reflectors and Factor 4 biofacies (dominated by *Stilostomella* spp.) in the upper part of the sequence (Figs. 2, 3). Part of the middle Miocene section contains transported components (dominated by *Bulimina gracilis*) mixed with predominantly in situ taxa (see below). Transported taxa dominate the upper middle to upper Miocene section (Figs. 2, 3).

#### Site 906

At Site 906 (913 m present water depth; ~870–930 m paleodepth), five factors explain 75% of the faunal variation (31 samples and 109 taxa). Middle Eocene strata were not penetrated at this site. The upper Eocene section is characterized by an *Osangularia* spp., *P. bulloides*,

Table 3. Age/paleobathymetric distributions of taxa in this study (RS = restricted to; C = most common in).

Taxon	Age	Paleodepth
<i>Alabamina wilcoxensis</i>	C early Eocene to earliest Miocene	C bathyal
<i>Anom. aragonensis/capitatus</i>	RS Eocene; C early to middle Eocene	C lower bathyal
<i>A. pseudogrosserugosus</i>	C late Oligocene to early Miocene	
<i>A. spissiformis</i>	C early to middle Eocene	
<i>Aragonia aragonensis</i>	C middle Eocene	C lower bathyal
<i>Arenobulimina truncata</i>	RS Eocene	
<i>Bolivina advena</i>	C late Oligocene to early Miocene	RS middle to upper bathyal
<i>B. furcata</i>	RS middle early Miocene	
<i>B. tectiformis</i>	RS late Eocene	
<i>Bulimina alazanensis</i>	C late Eocene and middle Miocene	
<i>B. impendens</i>	RS Eocene	
<i>B. macilenta</i>		C bathyal
<i>B. semicostata</i>	RS early to middle middle Eocene	C abyssal
<i>B. subtruncata</i>	RS late Eocene	RS bathyal
<i>B. trinitatensis</i>	RS early to middle Eocene	
<i>B. bradburytuxpamensis</i>	RS Eocene; C middle to late Eocene	C bathyal
<i>Buliminella gracilis</i>	C late middle Miocene	
<i>B. grata</i>	RS Eocene; C early Eocene	C abyssal
<i>Cassidulina</i> spp.	C late Eocene and middle Miocene	
<i>Cassidulinoides</i> spp.	RS late Oligocene to middle Miocene	
<i>Cibicidoides bradyi</i>	C late Eocene	RS bathyal
<i>C. cooki</i>	RS Eocene	RS bathyal
<i>C. crebbi</i>	RS middle early to middle Miocene	
<i>C. dickersoni</i>	C middle to late Eocene	
<i>C. eocaenus</i>	RS Eocene	
<i>C. havanensis</i>	RS Eocene	
<i>C. laurisae</i>	C late Eocene	RS bathyal
<i>C. micrus</i>	RS Eocene	
<i>C. pachyderma</i>	RS latest Oligocene to Miocene	
<i>C. praemundulus</i>		C lower bathyal
<i>Cibicidoides</i> sp. A	C late Eocene to Oligocene	
<i>Dorothia</i> sp.	RS Eocene	
<i>Eggerella</i> spp.		C bathyal
<i>Fursenkoina makiyama</i>	RS late Oligocene and middle Miocene	
<i>Globobulimina notovata</i>	C late Eocene to late Oligocene	RS lower bathyal
<i>Globobulimina</i> spp.	C late Oligocene to middle Miocene	
<i>Globocassidulina subglobosa</i>	C late Eocene to middle Miocene	
<i>Gyroidinoides</i> sp. 1	RS Eocene to Oligocene	
<i>Gyroidinoides</i> spp.	C late middle Eocene to Miocene	C bathyal
<i>Hanzawaia ammophila</i>	C early to middle Eocene	C lower bathyal
<i>Hoeglundina elegans</i>	RS Oligocene to Miocene	
<i>Karrerella chapapotensis</i>	C Eocene	
<i>K. subglabra</i>	C Eocene	
<i>Karrerella</i> spp.	C Eocene	
<i>Lenticulina peregrina</i>	RS late Oligocene to early Miocene	RS middle bathyal
<i>Lenticulina</i> spp.		C bathyal
<i>Loxostomum</i> cf. <i>applinae</i>	RS late Oligocene to middle Miocene	
<i>Marginulina hirsuta</i>	RS latest middle Eocene to middle Miocene	
<i>Martinottiella communis</i>	RS Miocene; C middle Miocene	
<i>Melonis barleeaanum</i>	C late Oligocene to middle Miocene	
<i>M. pompilioides</i>	C late Oligocene to middle Miocene	
<i>Nonion havanense</i>	RS Eocene	
<i>Nonionella pizarrensis</i>	C late early to middle Miocene	
<i>Nonionellina</i> spp.	C middle to late Eocene	C bathyal
<i>Nuttallides truempyi</i>	RS early to middle middle Eocene	C abyssal
<i>Osangularia</i> spp.	C Eocene	C bathyal
<i>Pararotalia</i> spp.	RS middle Miocene	
<i>Planulina</i> aff. <i>ambigua</i>	C middle Miocene	RS upper to middle bathyal
<i>Planulina costata</i>	C late Eocene	RS bathyal
<i>Plectina elongata</i>	RS Eocene; C middle Eocene	C lower bathyal
<i>Plectofrondicularia</i> spp.	C late Oligocene	C upper to middle bathyal
<i>Plectofrondicularia</i> spp.	C Miocene	C lower bathyal
<i>Pleurostomella</i> spp.	C Eocene	C bathyal
<i>Pleurostomellids</i>	RS Eocene	
<i>Polymorphinids</i>	C late Eocene to Miocene	
<i>Praeglobobulimina spinescens</i>	C late Oligocene	C middle to lower bathyal
<i>Pullenia bulloides</i>	C late Eocene	
<i>P. eocenica</i>	C middle Eocene	
<i>P. quinqueloba</i>	C Eocene to Oligocene	C bathyal
<i>Quadratobuliminella pyramidalis</i>	RS early to middle Eocene	RS abyssal
<i>Spirosigmoilina tenuis</i>	RS late Eocene to Miocene	
<i>Siphonina tenuicarinata</i>	C late Oligocene to early middle Miocene	
<i>Siphonina</i> sp.	RS early to earliest middle Eocene	RS lower bathyal
<i>Sphaeroidina bulloides</i>	C late Oligocene	
<i>Spiroculina</i> sp.	RS Oligocene to Miocene; C early Miocene	RS middle to lower bathyal
<i>Spiroplectammina spectabilis</i>	RS Eocene	
<i>Spirosigmoilina tenuis</i>	RS late Eocene to middle Miocene	C middle to lower bathyal
<i>Stilostomella aculeata</i>	C Eocene	
<i>Stilostomella</i> sp. C	RS late Oligocene to middle Miocene	
<i>S. modesta</i>	RS late Oligocene to middle Miocene	RS lower bathyal
<i>S. paucistriata</i>	RS late Eocene	
<i>Total Stilostomella</i> spp.	C late Eocene to middle Miocene	
<i>Trifarina danvillensis</i>	C late Eocene	
<i>Turrilina robertsi</i>	RS Eocene	C lower bathyal
<i>Uvigerina adelinensis</i>	RS middle Miocene	RS upper bathyal
<i>U. auberiana</i>	C late Eocene	
<i>U. aubertae</i>	RS early to middle Miocene	lower bathyal

Table 3 (continued).

Taxon	Age	Paleodepth
<i>U. chirana</i>	C late Oligocene	
<i>U. havanensis</i>	RS middle to late Eocene	RS bathyal
<i>U. hispida</i>	C middle Miocene	
<i>U. mexicana</i>	C latest Eocene to the late Oligocene	C middle bathyal
<i>U. pigmaea</i>	RS middle Miocene	
<i>U. proboscidea</i>	C late Oligocene to the early Miocene	
<i>U. semivestita</i>	RS late Eocene	RS lower bathyal
<i>U. spinulosa</i>	RS late Eocene to earliest Miocene	RS middle bathyal
<i>Vulvulina</i> spp.	C early to middle Eocene	

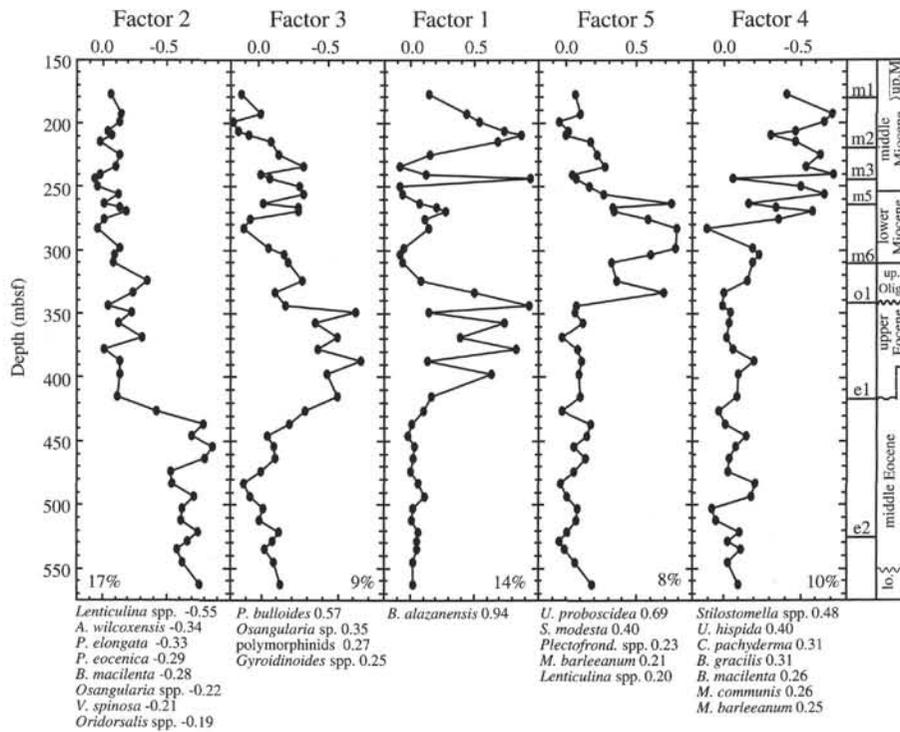


Figure 2. Site 904 Q-mode varimax factors. Percentage of the faunal variation explained by each factor and the species with the highest scores on each factor are shown. See Appendix for full taxonomic designations. Seismic stratigraphic correlations are after Mountain, Miller, Blum, et al. (1994) except where noted in the text.

*B. alazanensis*, and *Oridorsalis* spp. dominated fauna (Factor 3; Figs. 4, 5), similar to the coeval biofacies at nearby Site 902 and deeper Site 904. A shift in biofacies occurs across the disconformable Eocene/upper Oligocene contact from Factor 3 to Factors 4 and 1. Most of the upper Oligocene at Site 906 is dominated by the Factor 1 biofacies (*Lenticulina* spp., *Cibicidoides* aff. *praemundulus*, and *A. wilcoxensis*), with some samples showing high loadings on Factors 4 and 5 (*Stilostomella* sp. C, polymorphinids, *Plectofrondicularia* spp., *M. barleeaanum*, *Melonis pompilioides*, *Gyroidinoides* spp., and *Globocassidulina subglobosa*) (Figs. 4, 5). The upper Oligocene biofacies at Site 906 is comparable to that at nearby Site 902, which has a similar modern and backstripped water depth. The middle Miocene section examined at Site 906 consists of canyon fill deposited on Reflector m3; the biofacies of this canyon fill (Factor 2) are characterized by in situ taxa (*Uvigerina hispida*, *B. alazanensis*, *Cibicidoides pachyderma*, *Cibicidoides crebbsi*) mixed in with transported neritic foraminifers (*Buliminella gracilis*) (Figs. 4, 5). Of the three canyon-fill lithostratigraphic units, we only examined samples from the lower mud-clast conglomerate Unit VC (i.e., below reflector canyon 2, Fig. 5; Mountain, Miller, Blum, et al., 1994).

### Site 902

At Site 902 (811 m present water depth; ~790–830 m paleodepth), five factors explain 77% of the faunal variation (41 samples and 108 taxa). Middle Eocene strata were not penetrated at this site. The upper Eocene section is characterized by a *P. bulloides*, *B. alazanensis*, *Osangularia* spp., *Gyroidinoides* spp., and *Oridorsalis* spp. biofacies (Factors 2 and 4; Figs. 6, 7), similar to coeval biofacies at Sites 904 and 906. *Bulimina alazanensis* is characteristic of the upper Eocene section at Site 902 (Factor 2), as it is at Sites 904 and 906. Factors 1, 3, and 5 (*Lenticulina* spp. and polymorphinids, Factor 1; *Uvigerina mexicana*, *M. pompilioides*, *A. ignota*, and *Stilostomella* spp., Factor 3; and *A. wilcoxensis* and *M. barleeaanum*, Factor 5) characterize the lower Miocene section (Figs. 6, 7). There is an unusually high degree of fluctuation among Factors 1, 3, and 5 at Site 902, although these biofacies are generally similar to those found at the other Leg 150 sites at coeval levels. These faunal variations may reflect local influences, high frequency climatic variations, or be an artifact of factor analysis.

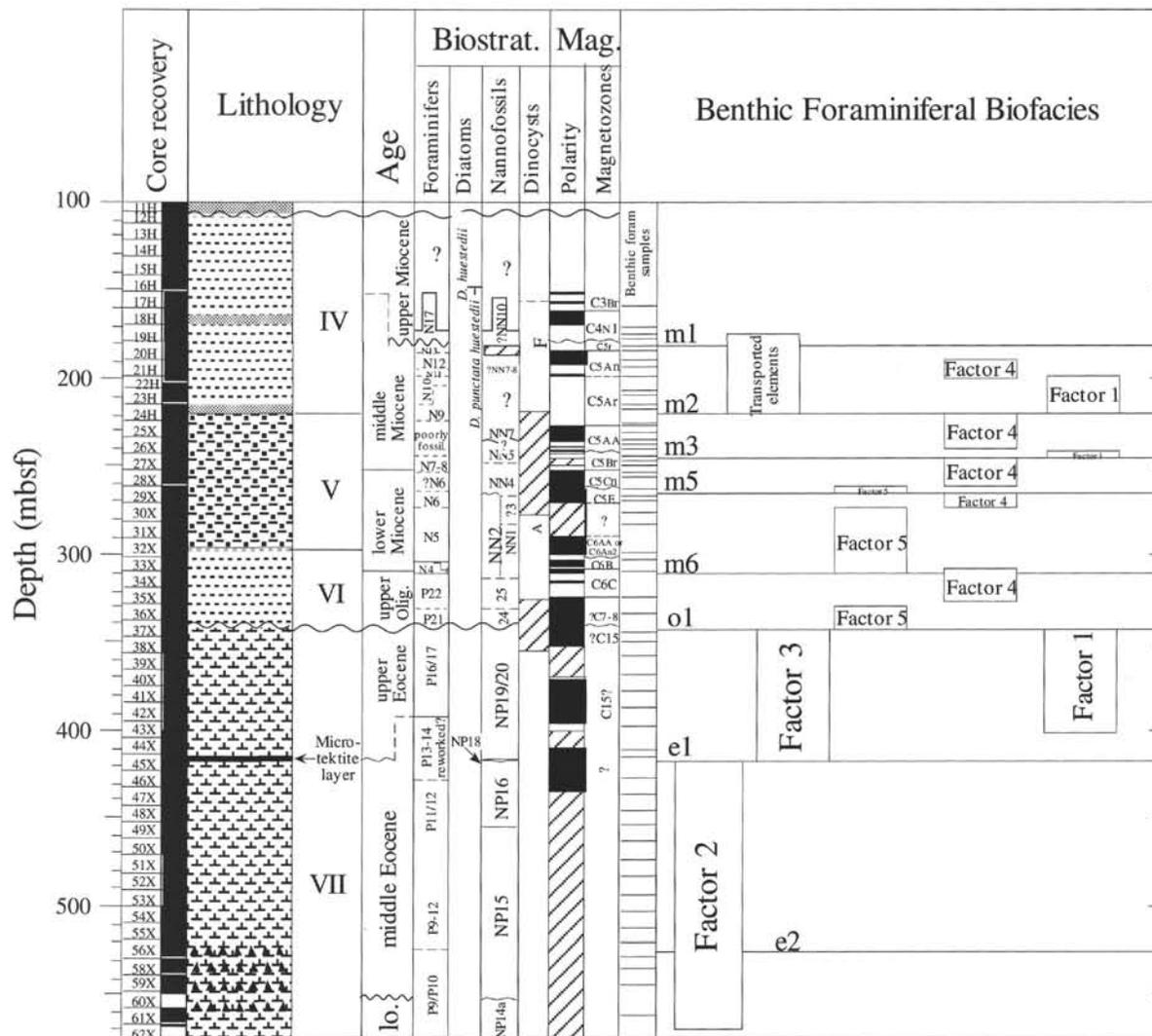


Figure 3. Site 904 biofacies represented by Q-mode varimax factors. Seismic stratigraphic correlations are after Mountain, Miller, Blum, et al. (1994) except where noted in the text. Biostratigraphy includes planktonic foraminifers (Snyder et al., this volume), nannofossils, diatoms, and dinocysts (Mountain, Miller, Blum, et al., 1994). Lithostratigraphy is after Mountain, Miller, Blum, et al. (1994). Magnetostratigraphy is after Van Fossen and Urrutia (this volume).

**Site 903**

At Site 903 (444 m present water depth; ~470–500 m paleodepth), five factors explain 65% of the faunal variation (34 samples and 132 taxa). The middle to upper Eocene section is characterized by faunas (*P. bulloides*, *B. alazanensis*, *Osangularia* spp., *Cibicidoides* aff. *praemundulus*, and *Lenticulina* spp.; Factors 1 and 3; Figs. 8, 9) similar to those found at the deeper Sites 902, 904, and 906. As at all other Leg 150 sites, a faunal change is associated with the disconformable Eocene/Oligocene boundary as the Factor 1 biofacies (*Lenticulina* spp., polymorphinids, *U. proboscidea*, *A. wilcoxensis*, *M. barleeianum*, and *Gyroidinoides* spp.) becomes dominant in the upper Oligocene section.

In the lower to lower middle Miocene section at Site 903, the dominant biofacies fluctuates among Factor 1, Factor 4 (*Cibicidoides crebbisi*, *Globocassidulina subglobosa*, and *Stilostomella* spp.), and Factor 5 (*M. pompilioides*, *Uvigerina aubertae*, *Martinottiella communis*, *U. hispida*, *Cibicidoides* aff. *mundulus*, *Stilostomella* sp. C, *U. proboscidea*, and *U. adelinensis*). The Factor 5 biofacies (dominated by *M. pompilioides*) shows peaks associated with Reflectors m6 and m4; below, we show that the taxon *M. pompilioides* has abundance

peaks that span Reflectors m4 and m5 at this site. An interval in the lower Miocene yielded rare benthic foraminifers. The upper Oligocene to lower middle Miocene biofacies at Site 903 are similar to coeval Site 902 biofacies. Whereas there are similarities to coeval Site 904 faunas, there are minor distinctions in the benthic foraminiferal composition that reflect shallower water depths at Site 903. For example, *Cibicidoides crebbisi* (dominant Factor 4 species) and *Meloniis pompilioides* (dominant Factor 5 species) are more common at Site 903 than at Site 904; these taxa tend to be most abundant shallower than ~1000 m (van Morkhoven et al., 1986; Katz and Miller, 1993). As at the other Leg 150 sites, *B. alazanensis* shows peak abundances in the upper Eocene and in the middle Miocene at Site 903. Factor 2 (*Buliminella gracilis*, *Uvigerina hispida*, *Bulimina alazanensis*, *Meloniis barleeianum*, and *Uvigerina pigmaea*) characterizes part of the middle Miocene section at Site 903.

Downslope transport becomes significant in the middle Miocene section at Site 903 near Reflector m3. *Buliminella gracilis* and *Uvigerina elongata* are most common at inner to middle neritic depths (<100 m) on the New Jersey Oligocene to Miocene coastal plain (Olsson et al., 1980), deltaic environments on the New Jersey shelf (Poag, 1978), and on the Maryland Miocene coastal plain (Gibson,

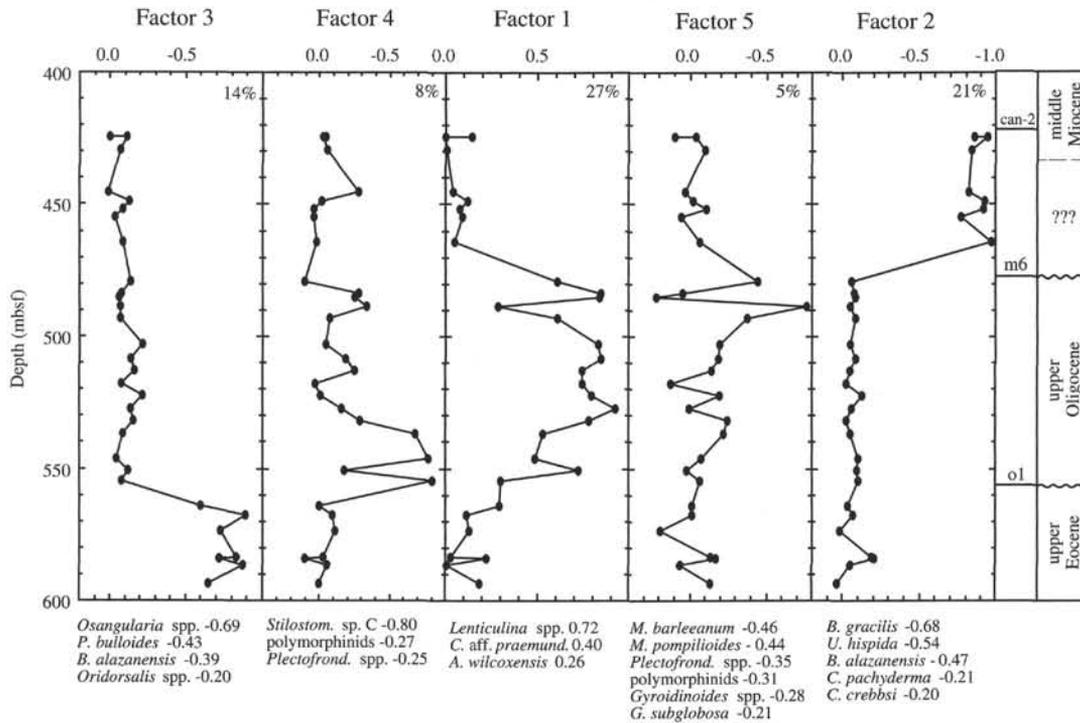


Figure 4. Site 906 Q-mode varimax factors. Percentage of the faunal variation explained by each factor and the species with the highest scores on each factor are shown. See Appendix for full taxonomic designations. Seismic stratigraphic correlations are after Mountain, Miller, Blum, et al. (1994) except where noted in the text.

1972); we use these taxa as indicators of downslope transport. Factor 2 represents mixed in situ and transported material (*B. gracilis* is <27% in Factor 2). *Buliminella gracilis* comprises 80%–100% of the biofacies above Reflector m2 (700 mbsf; Mountain, this volume), and *Uvigerina elongata* comprises nearly 100% of the faunas above Reflector m1 (604.50 mbsf; Mountain, this volume) (Fig. 9), indicating that allochthonous facies dominate much of the middle Miocene section at Site 903.

### Biofacies Age vs. Paleodepth Distribution on the New Jersey Transect

The New Jersey Transect provides a record from the bathyal to abyssal zones for most of the Eocene. As a result of poor core recovery at abyssal sites drilled prior to Leg 150 (Poag, Watts, et al., 1987; van Hinte, Wise, et al., 1987), Oligocene to Miocene records are limited to the bathyal zone in this region. The age vs. paleodepth coverage is comprehensive for the Eocene through middle Miocene bathyal samples and for the lower to middle Eocene abyssal samples (Figs. 10, 11). We present bubble plots both for factors (Figs. 10, 11) and for species percentages (Fig. 12A–L).

The age vs. paleobathymetric distributional data presented here (Table 3; Fig. 12A–L) are consistent with previous studies for the Eocene Atlantic (e.g., Tjalsma and Lohmann, 1983), the Oligocene Atlantic (M.E. Katz, K.G. Miller, and R.C. Tjalsma, unpubl. data, 1995), and the Neogene of the Gulf of Mexico (Katz and Miller, 1993). For instance, *Nuttallides truempyi* abundances decreased dramatically in the middle Eocene; on the New Jersey Margin, this species disappeared by the end of the middle Eocene (Fig. 12G), whereas it was present elsewhere in very low numbers into the late Eocene (e.g., Katz and Miller, 1991; Thomas, 1990; Tjalsma and Lohmann, 1983). Other common cosmopolitan Eocene species that are found on the New Jersey Margin include *Aragonia aragonensis*, *Bulimina impendens*, *B. semicostata*, *B. trinitatensis*, *B. tuxpamensis*, *Cibici-*

*doides micrus*, *Hanzawaia ammophila*, *Nonion havanense*, and *Quadratobulimina pyramidalis* (additional common Eocene taxa are shown in Fig. 12A–L).

Other taxa also show depth and/or age distributions on the New Jersey Margin that are similar to other Atlantic locations. For instance, *Osangularia* spp. and *Pleurostomella* spp. were most abundant in the Eocene bathyal zone at the Leg 150 sites (Fig. 12A–L), whereas polymorphinids were most abundant in the late Eocene to the Miocene (Fig. 12I). Similarly, Tjalsma and Lohmann (1983) showed that *Osangularia mexicana* was most abundant in the Eocene bathyal zone in the circum-Atlantic region. *Globocassidulina subglobosa* (Fig. 12E), *Gyroidinoides* spp. (Fig. 12E), and *Stilostomella* spp. (Fig. 12K) abundances increased in the late Eocene bathyal zone and continued to be major faunal constituents through the middle Miocene. Tjalsma and Lohmann (1983) found that abundances of *Globocassidulina subglobosa*, *Gyroidinoides* spp., and *Stilostomella* spp. increased in the middle Eocene at many of their abyssal circum-Atlantic sites, which is consistent with the New Jersey Margin distributions.

Some taxa appear to have restricted age or paleodepth distributions on the New Jersey Margin compared to other locations. *Bolivina tectiformis* (Fig. 12B), *Bulimina subtruncana* (Fig. 12B), *Stilostomella paucistriata* (Fig. 12J), and *Uvigerina semivestita* (Fig. 12L) were restricted to the late Eocene at our slope sites, but have wider distributions at other Atlantic locations. For example, *Bolivina tectiformis* has been reported at abyssal paleodepths in Oligocene to Miocene sediments in the North Atlantic (Miller and Katz, 1987b) and at a bathyal paleodepth in Oligocene South Atlantic sediments (Tjalsma, 1983). *Uvigerina semivestita* has been reported at middle to upper lower bathyal depths in upper Eocene to Oligocene Atlantic sediments (Boersma, 1984).

Eocene benthic foraminifers on the New Jersey Transect represent cosmopolitan deep-water faunas. Early to middle Eocene bathyal biofacies (Sites 108, 612, 902, 903, 904, and 906) were character-

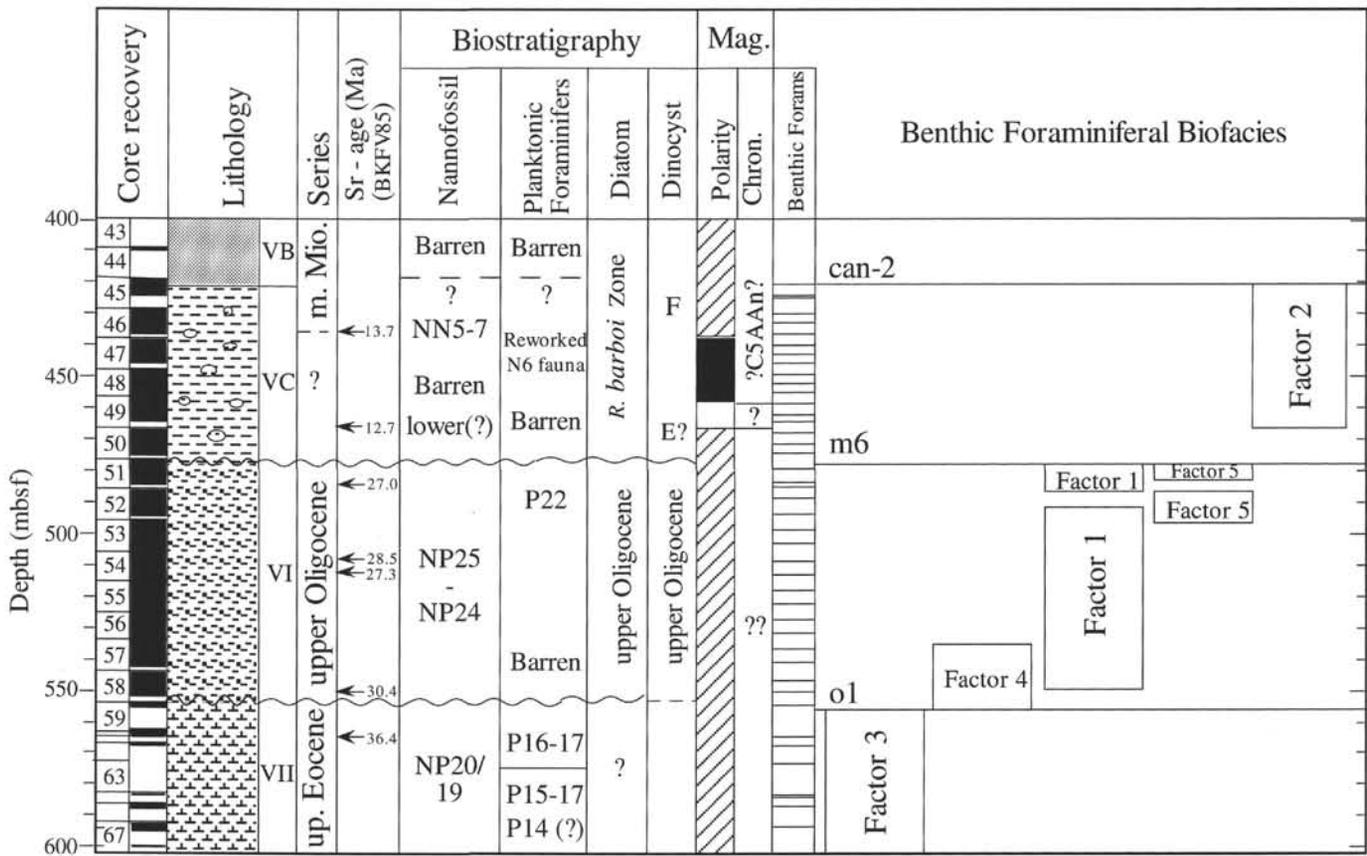


Figure 5. Site 906 biofacies represented by Q-mode varimax factors. Seismic stratigraphic correlations are after Mountain, Miller, Blum, et al. (1994) except where noted in the text. Biostratigraphy includes planktonic foraminifers (Snyder et al., this volume), nannofossils, diatoms, and dinocysts (Mountain, Miller, Blum, et al., 1994). Lithostratigraphy is after Mountain, Miller, Blum, et al. (1994). Magnetostratigraphy is after Van Fossen and Urbat (this volume).

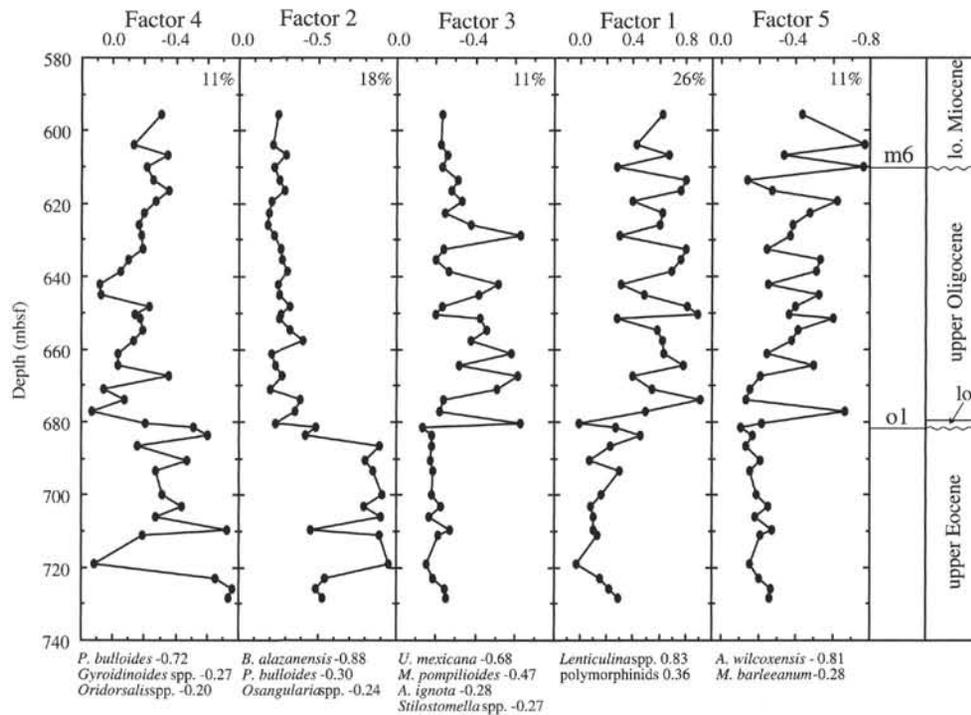


Figure 6. Site 902 Q-mode varimax factors. Percentage of the faunal variation explained by each factor and the species with the highest scores on each factor are shown. See Appendix for full taxonomic designations. Seismic stratigraphic correlations are after Mountain, Miller, Blum, et al. (1994) except where noted in the text.

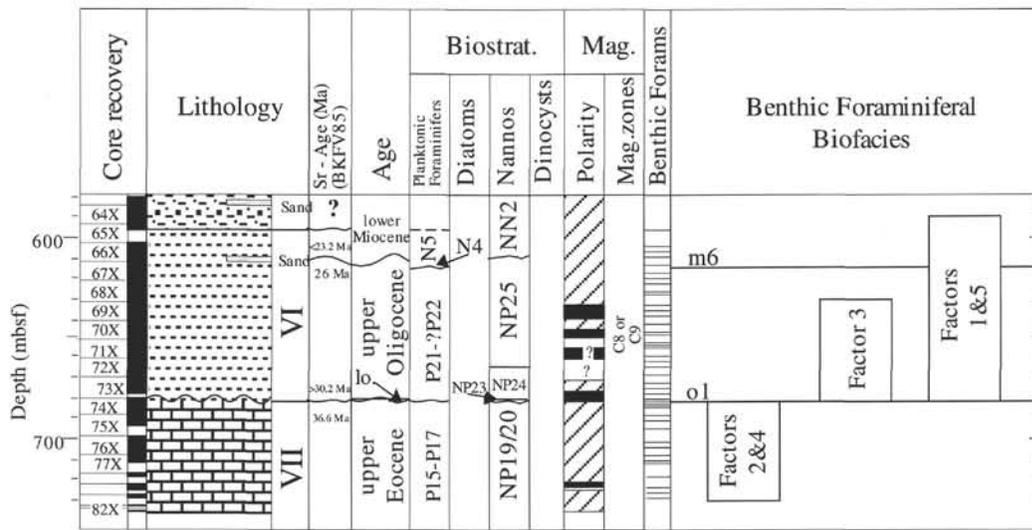


Figure 7. Site 902 biofacies represented by Q-mode varimax factors. Seismic stratigraphic correlations are after Mountain, Miller, Blum, et al. (1994) except where noted in the text. Biostratigraphy includes planktonic foraminifers (Snyder et al., this volume), nannofossils, diatoms, and dinocysts (Mountain, Miller, Blum, et al., 1994). Lithostratigraphy is after Mountain, Miller, Blum, et al. (1994). Magnetostratigraphy is after Van Fossen and Urrat (this volume).

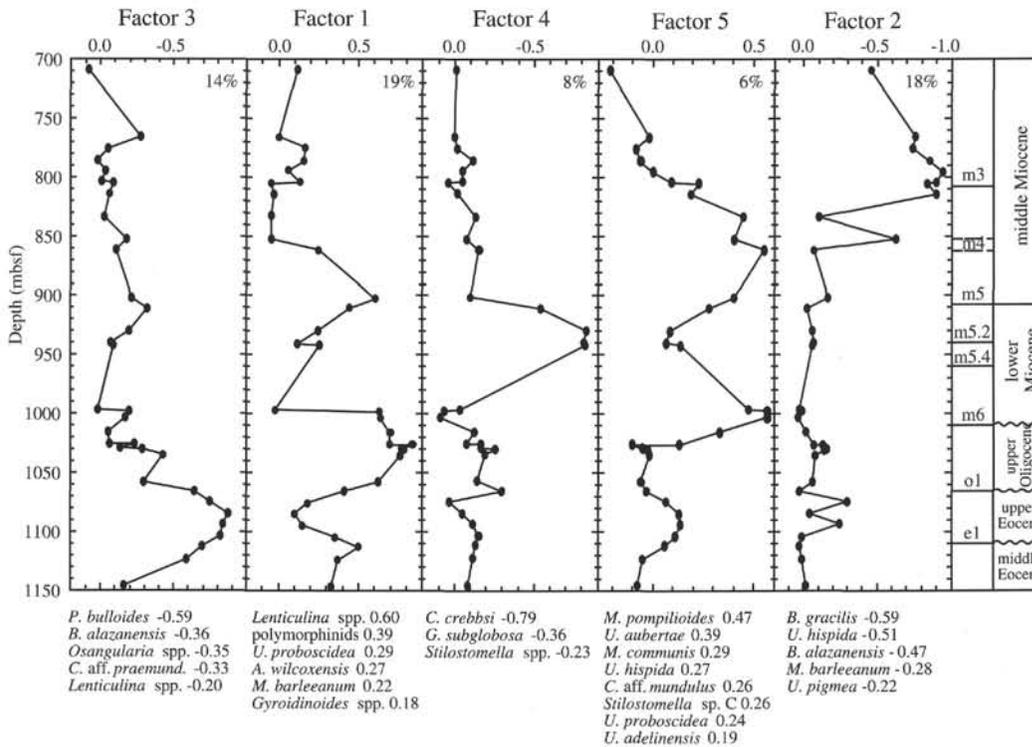


Figure 8. Site 903 Q-mode varimax factors. Percentage of the faunal variation explained by each factor and the species with the highest scores on each factor are shown. See Appendix for full taxonomic designations. Seismic stratigraphic correlations are after Mountain, Miller, Blum, et al. (1994) except where noted in the text.

ized by *Lenticulina* spp. and *A. wilcoxensis* (Factor 2) and *Osangularia* spp. (Factor 4) (Fig. 10). Coeval abyssal Sites 390 and 613 contain a deeper *N. truempyi*-dominated biofacies (Factor 5) in the early to middle Eocene (see also Miller and Katz, 1987a). Bathyal biofacies on the New Jersey Transect shifted to a biofacies dominated by *B. alazanensis* (Factor 1), *Osangularia* spp. (Factor 4), and *P. bulloides* (Factor 3) by the late Eocene. Although we lack an abyssal late Eocene record, we believe that the major bathyal faunal change that occurred near the end of the middle Eocene on the New Jersey

Transect reflects a global event that is also recorded at other bathyal and abyssal sites (e.g., Katz and Miller, 1991; Thomas, 1990; Tjalsma and Lohmann, 1983). Miller et al. (1992) and Thomas (1992a) speculated that changing deep-water conditions associated with a late middle Eocene cooling (inferred from the  $\delta^{18}O$  record) triggered these faunal changes worldwide.

Late Eocene bathyal faunas on the New Jersey Transect contain high abundances of *B. alazanensis* (up to 50%; Fig. 12B); similar coeval peak *B. alazanensis* abundances have been reported from the

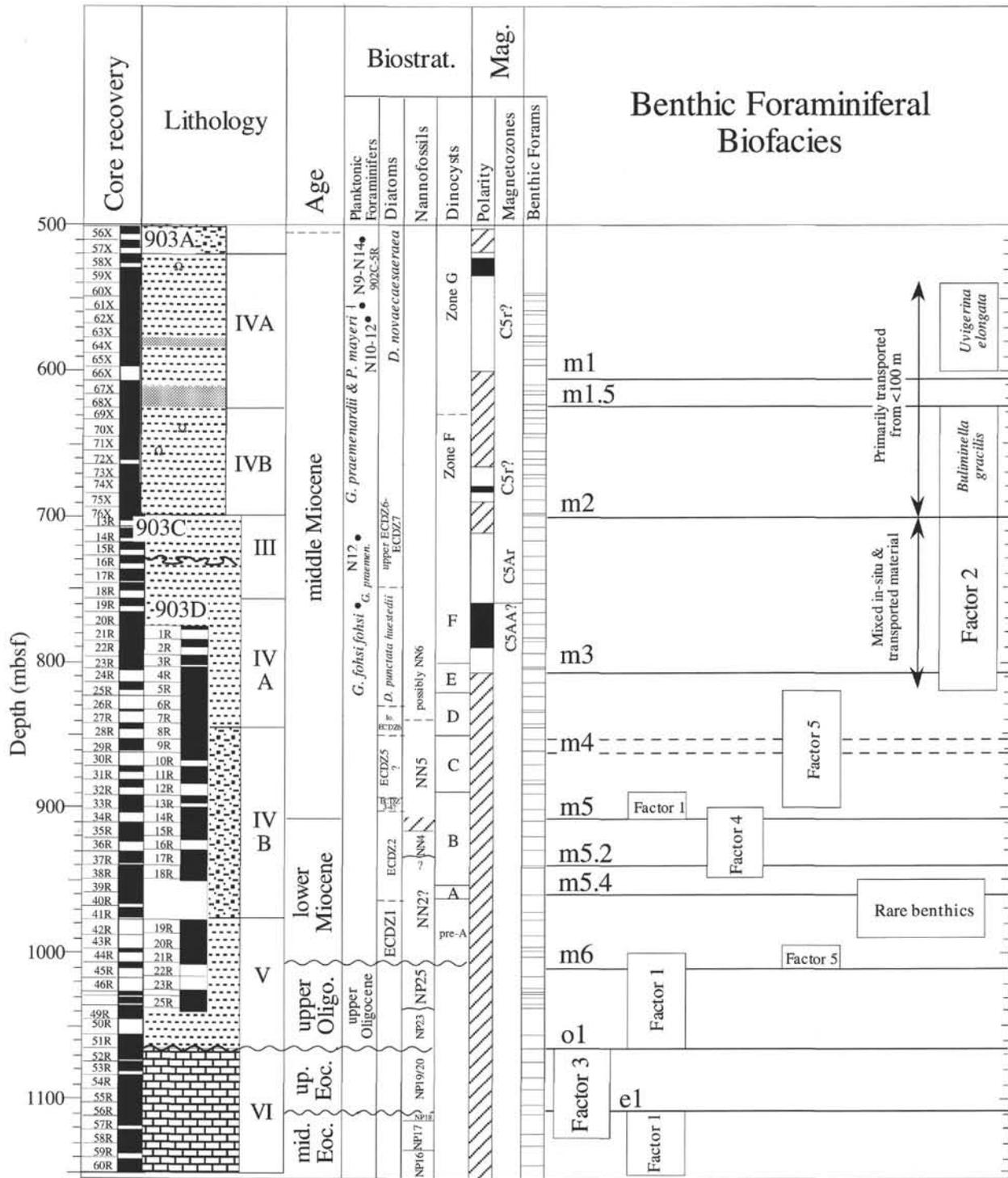


Figure 9. Site 903 biofacies represented by Q-mode varimax factors. Seismic stratigraphic correlations are after Mountain, Miller, Blum, et al. (1994) except where noted in the text. Biostratigraphy includes planktonic foraminifers (Snyder et al., this volume), nannofossils, diatoms, and dinocysts (Mountain, Miller, Blum, et al., 1994). Lithostratigraphy is after Mountain, Miller, Blum, et al. (1994). Magnetostratigraphy is after Van Fossen and Urbat (this volume).

Gulf of Mexico (E67-128, ~1600 m paleodepth; B.H. Corliss, pers. comm., 1994). Increases in *B. alazanensis* abundances (<15%) have been noted in other upper Eocene Atlantic sections: Site 516 (South Atlantic; ~1000 m paleodepth; Tjalsma, 1983), Site 548 (Irish Margin; ~1200 m paleodepth; Miller et al., 1985), Site 144 (east of Venezuela; ~2500 m paleodepth; M.E. Katz, K.G. Miller, and R.C. Tjals-

ma, unpubl. data, 1995), and Site 366 (Sierra Leone Rise; ~2800 m paleodepth; M.E. Katz, K.G. Miller, and R.C. Tjalsma, unpubl. data, 1995). We speculate that this uniform late Eocene biofacies characterized by high abundances of *B. alazanensis* indicates that circum-Atlantic sites were ventilated by similar intermediate to upper deep-water masses at this time.

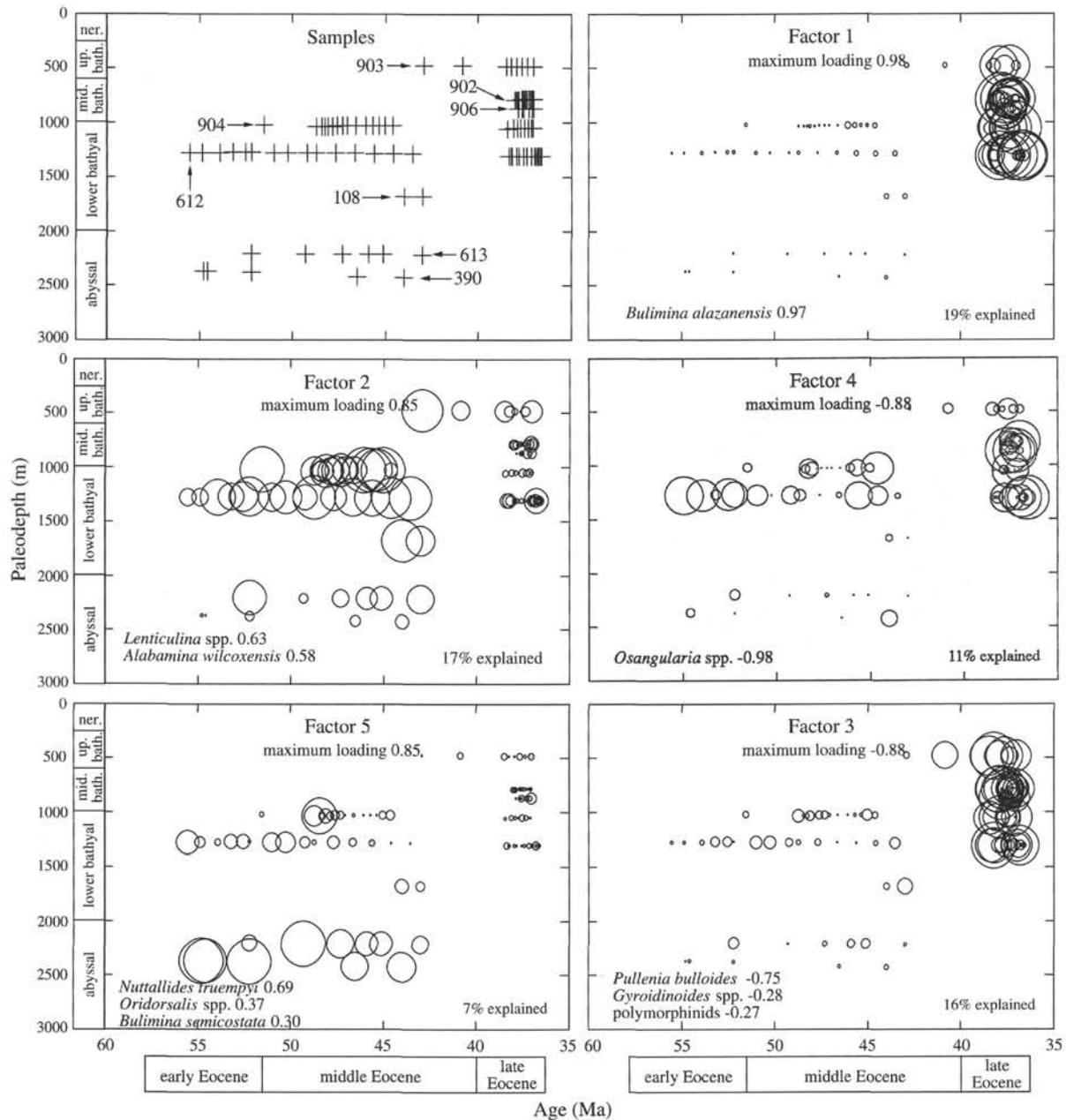


Figure 10. Eocene age-paleodepth bubble diagrams of Q-mode varimax factor combined data set loadings for the New Jersey Transect (ODP Sites 902, 903, 904, and 906 and DSDP Sites 108, 390, 612, and 613). Bubble size is proportional to each loading, with the maximum loading (largest bubble size) shown on each panel. Percentage of the faunal variation explained by each factor and the species with the highest scores on each factor are shown.

A cosmopolitan *Lenticulina* spp.-dominated biofacies prevailed in the Oligocene to early Miocene bathyal zone (note that we have no abyssal record for this interval); polymorphinids and *A. wilcoxensis* were common secondary components (Factor 1; Fig. 11). In addition, biofacies represented by Factors 4 (dominated by *U. proboscidea*, *Plectofrondicularia* spp., and *S. modesta*) and 5 (characterized by *M. pompilioides*, *Stilostomella* spp., *M. barleeianum*, *P. bulloides*, and *Plectofrondicularia* spp.) show high loadings during this interval. A *U. hispida*-*B. alazanensis*-dominated biofacies characterized the middle Miocene (Factor 3). A similar coeval shift from a *Lenticulina* spp.-dominated biofacies to a *Uvigerina* spp.-dominated biofacies occurred in the Gulf of Mexico (Katz and Miller, 1993), indicating that this *Uvigerina* increase may be regional in extent. The faunal

shift from the *Lenticulina* spp.-dominated biofacies to the *U. hispida*-*B. alazanensis*-dominated biofacies may be the regional expression of a global late early to early middle Miocene benthic foraminiferal turnover event and associated abundance changes. Previous studies have speculated that this global middle Miocene benthic foraminiferal faunal event may have been related to changes in deep-water sources and/or surface ocean productivity (e.g., Miller and Katz, 1987b; Thomas and Vincent, 1987; Thomas, 1992b; Woodruff and Savin, 1989). Katz and Miller (1993) found that the increased uvigerinid flux in the Gulf of Mexico corresponds with increased organic carbon flux in middle Miocene sediments, indicating that surface ocean productivity may have been responsible at least in part for the global middle Miocene faunal event. Further supporting this,

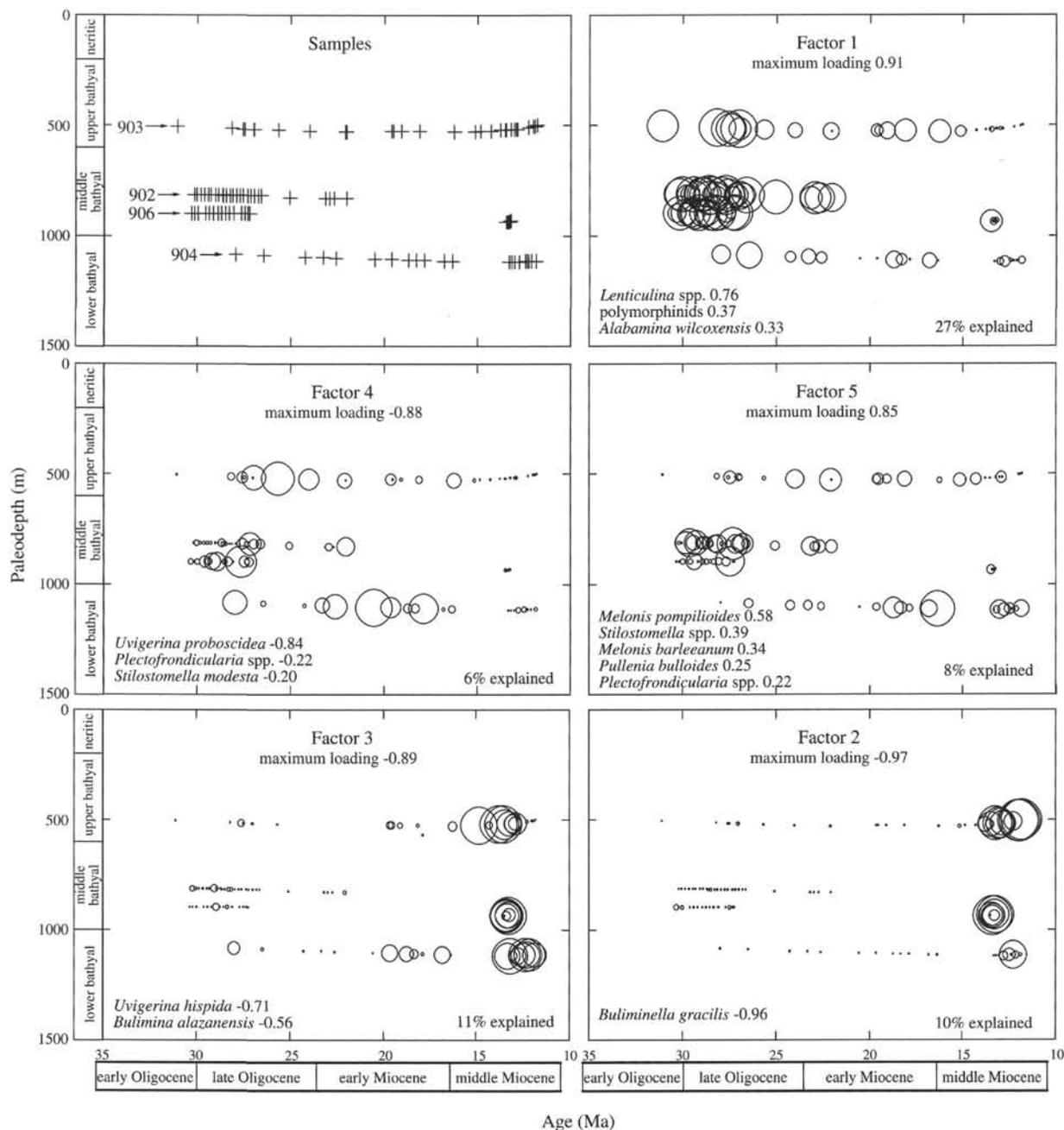


Figure 11. Oligocene to Miocene age-paleodepth bubble diagrams of Q-mode varimax factor combined data set loadings for the New Jersey Transect (ODP Sites 902, 903, 904, and 906 and DSDP Sites 108, 390, 612, and 613). Bubble size is proportional to each loading, with the maximum loading (largest bubble size) shown on each panel. Percentage of the faunal variation explained by each factor and the species with the highest scores on each factor are shown.

high abundances of Holocene *Uvigerina* spp. have been linked to a high supply of organic material to the sediments (see Thomas et al., 1995, for summary).

The Factor 2 allochthonous biofacies (characterized by *Bulminella gracilis*; Fig. 11) increased in dominance through the middle to late middle Miocene. A dramatic change occurred on the New Jersey slope between Reflectors m3 (13.5 Ma) and m2 (12.5 Ma), with a major increase in downslope transport of neritic (shelf) taxa (*B. gracilis*, *U. elongata*) (see below). This faunal change is associated with a dramatic increase in sedimentation rates (>60 cm/yr), illustrated by the increase in thickness between Reflectors m3 and m2. This change in slope sedimentation style is associated with submarine channel cut-

ting, high terrestrial carbon input, and mobilization of biogenic carbonate to form diagenetic nodules (Mountain, Miller, Blum, et al., 1994).

### Biofacies Changes Within a Sequence Stratigraphic Framework

Benthic foraminiferal biofacies changes have proven to be useful in deciphering depositional sequences in shallow-water deposits (e.g., Olsson et al., 1987; Christensen et al., in press). To evaluate whether slope benthic foraminifers exhibit similar biofacies changes, we examined samples across the m5 sequence (ca. 16.5 to 15.8 Ma;

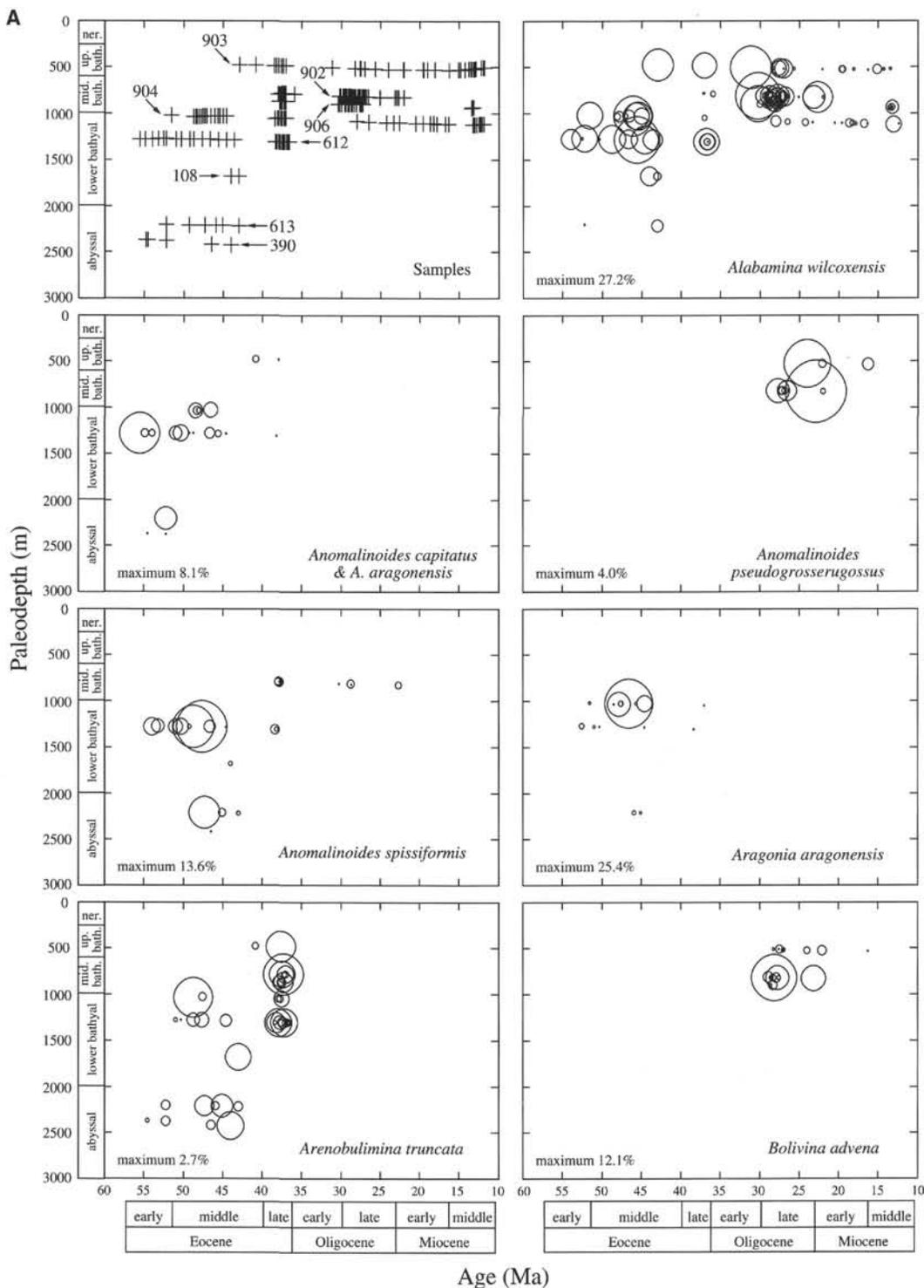


Figure 12. A–L. Eocene to Miocene age-paleodepth bubble diagrams of benthic foraminiferal species for the New Jersey Transect (ODP Sites 902, 903, 904, and 906 and DSDP Sites 108, 390, 612, and 613). Bubble size is proportional to each species percentage; maximum species percentage (largest bubble size) is shown on each panel.

Miller et al., this volume) at Site 903 (40 samples between 839.28 and 921.09 mbsf) and at Site 904 (24 samples between 223.06 and 268.70 mbsf). The m5 sequence at Site 903 is bracketed by Reflector m5 (908.4 mbsf) below and Reflector m4 (between 852 and 862 mbsf) above. The m4 sequence is missing at Site 904; here, the m5 sequence is bracketed by Reflector m5 (264.2 mbsf) below and Reflector m3 (244 mbsf) above (Figs. 13, 14).

We evaluated benthic foraminiferal absolute (number per gram) and relative abundance changes within the sequence stratigraphic framework outlined above. No good correlation was found between the number of benthic foraminifers per gram and sequences (Figs. 13, 14). However, several species relative abundance changes appear to be related to sequence boundaries. *Melinis pompilioides* shows higher percentages across the boundaries, whereas total *Stilostomella* spp.

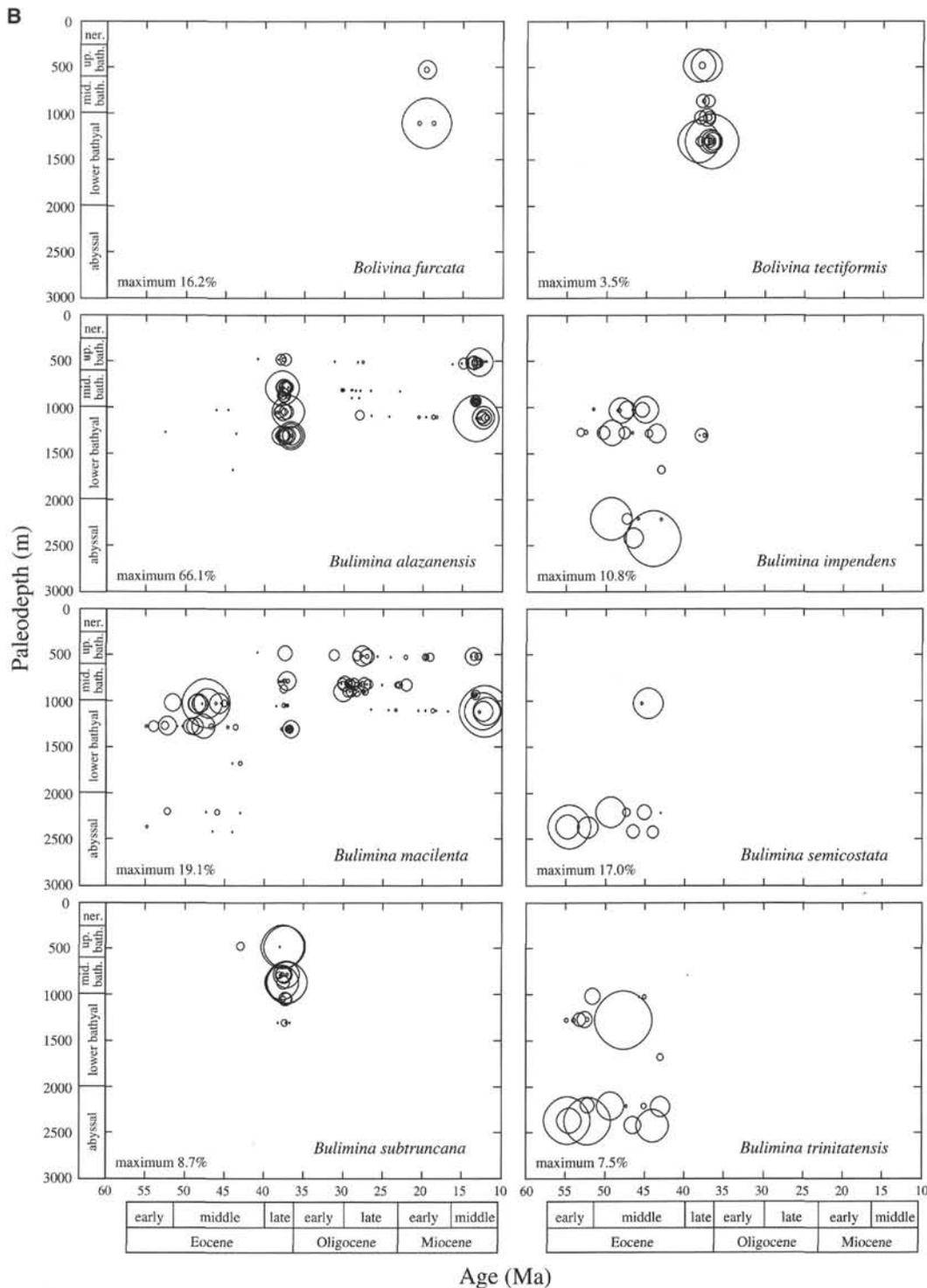


Figure 12 (continued).

increases immediately below the boundaries at Site 903 (Fig. 13); these patterns are not observed at the deeper Site 904 (Fig. 14). However, *U. proboscidea* increases above Reflector m5 at both sites, and above Reflector m4 at Site 903 (note that the corresponding m4 sequence is absent at Site 904) (Figs. 13, 14). We interpret *U. proboscidea* as in situ, indicating that the dominant biofacies immediately above the reflectors and correlative sequence boundaries are not transported (1.7 m above m5 and 0.5 m above m4 at Site 903; 1.5 m

above m5 at Site 904). Based on the higher abundances of *U. proboscidea* immediately above sequence boundaries, we suggest that Reflector m4 corresponds to 862 mbsf at Site 903. This level corresponds to the base of a minor lithologic color change (Mountain, Miller, Blum, et al., 1994).

Although the dominant benthic foraminifers are in situ, there is limited evidence of shelf transport in the m5 to m3 interval. We consider *Buliminella gracilis* and *Nonionella pizarrensis* to be down-

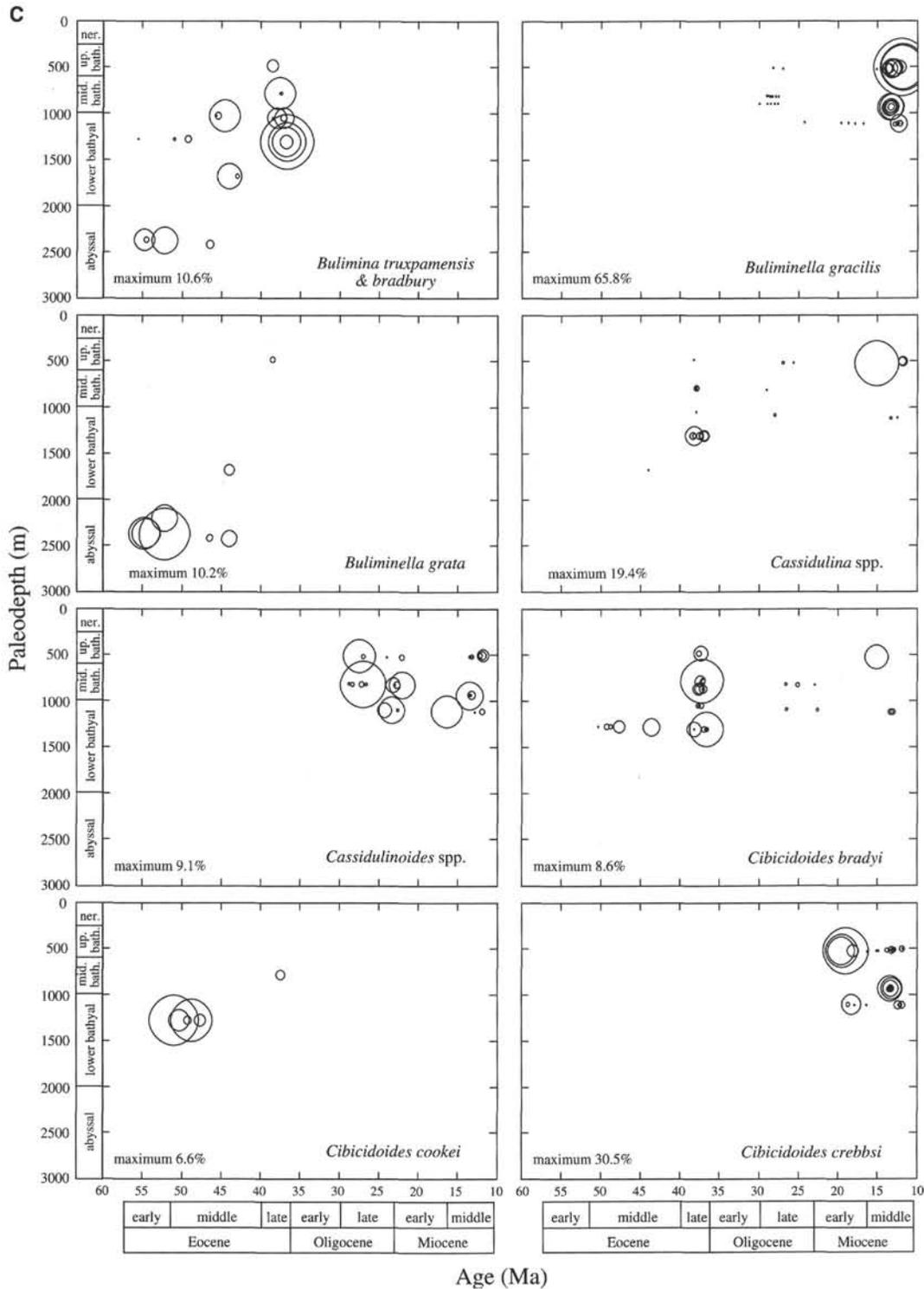


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slope-transport indicators because they typify Oligocene to Pliocene inner neritic environments on the mid-Atlantic United States margin (Gibson, 1972; Olsson et al., 1980; Poag, 1978). Neither taxon is as abundant in the m5–m3 interval as in the overlying upper middle Miocene and younger sections, where the dominant mode of sedimentation is downslope transport (Mountain, Miller, Blum, et al., 1994; Christensen et al., this volume; this study, see below). At the shallower Site 903, *N. pizarrensis* is most abundant above and below

Reflector m5, whereas *B. gracilis* is most abundant across Reflector m4 (Fig. 13). However, both taxa decrease immediately below the unconformities and show abundance peaks immediately above them. Similarly, *N. pizarrensis* decreases immediately below Reflector m5 at Site 904 and peaks immediately above it (Fig. 14). Thus, *B. gracilis* and *N. pizarrensis* occur in greatest abundances both above several sequence boundaries and in the upper parts of several sequences (Figs. 13, 14). This evidence for downslope transport, albeit a minor



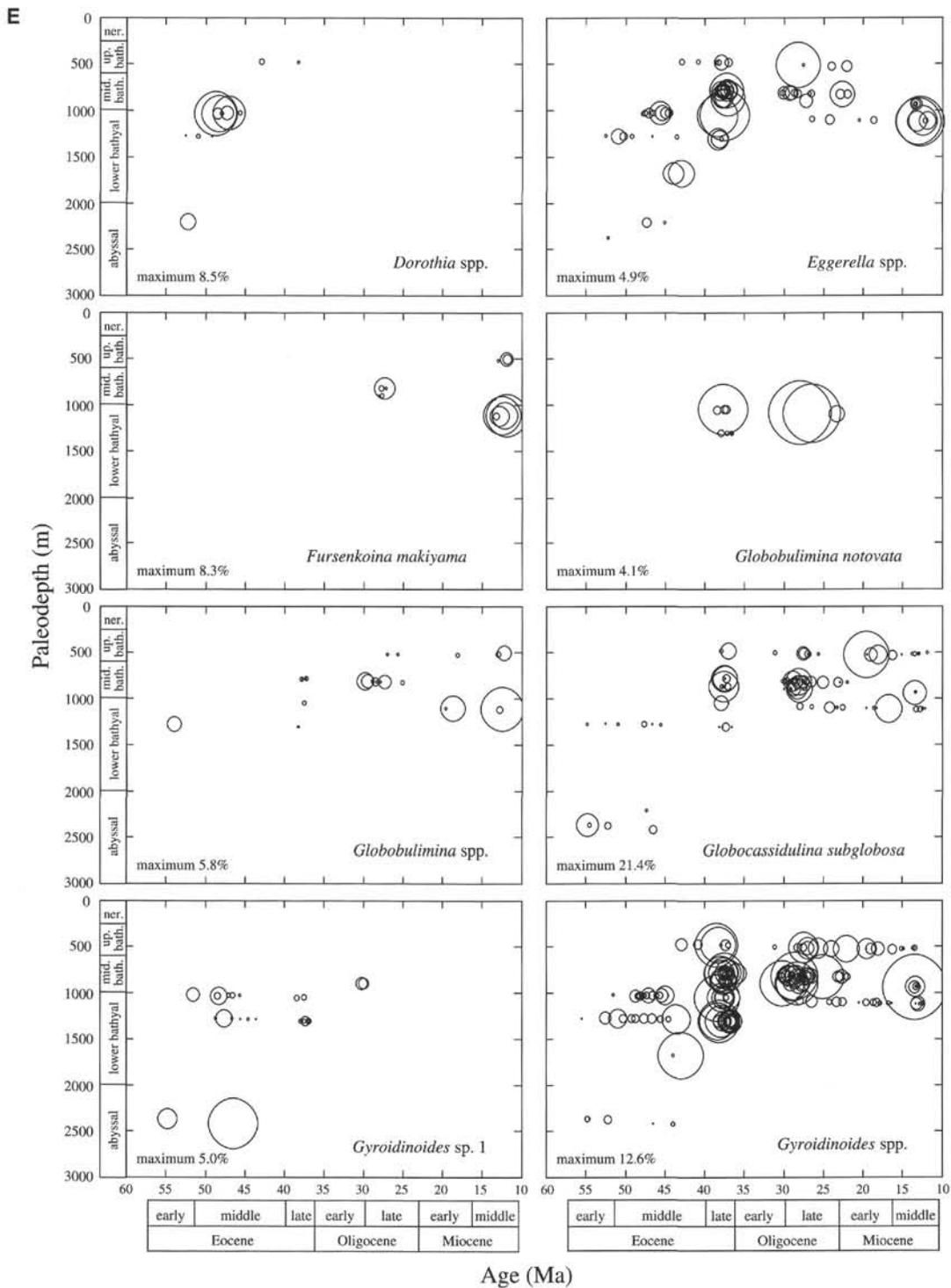


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of several sequences, which may be the slope equivalent of the shelf highstand systems tracts; and (4) more abundant *M. pompilioides* spanning several sequence boundaries. The sections immediately above sequence boundaries that contain predominantly in situ faunas with minor abundances of transported benthic foraminifers (<10% of the total fauna) may correlate to the lowstand systems tracts on the shelf. Minor abundances of transported benthic foraminifers reappear in the upper parts of several sequences, and may reflect minor shed-

ding of shelf sediments during highstand systems tracts progradation on the shelf.

### Transport and Sedimentation

Benthic foraminifers indicate that sedimentation was primarily in situ at our slope sites in the Eocene to early Miocene. All of the Eocene sections recovered yield in situ faunas, in agreement with

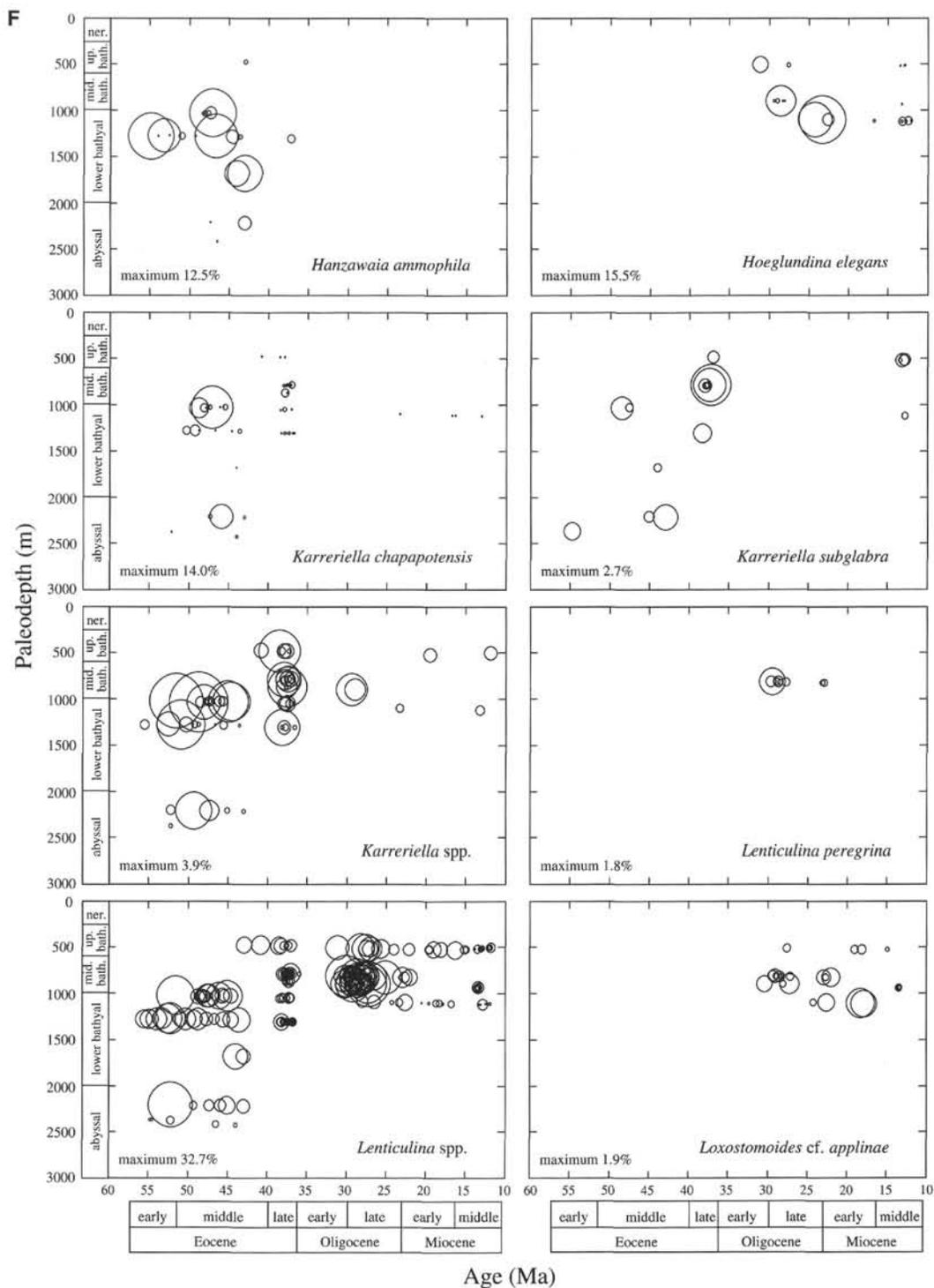


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previous studies at Sites 108, 390, 612, and 613 (Miller and Katz, 1987a). Transported biofacies are minimal in the Oligocene to lower middle Miocene sections at Sites 902, 903, 904, and 906, documenting for the first time that “quiescent” sedimentation continued in the region of our New Jersey slope sites well into the Neogene.

A dramatic change in the dominant mode of sedimentation occurred in the middle Miocene in our study area. At Sites 903 and 904, abundant downslope transport of shelf taxa (*B. gracilis*, *U. elongata*)

appears between Reflectors m3 (13.5 Ma) and m2 (12.5 Ma) (Fig. 15). *Buliminella gracilis* comprises nearly 100% of the faunas in the middle middle Miocene and *U. elongata* comprises nearly 100% of the faunas in the upper middle Miocene at Site 903 (Fig. 9). *Buliminella gracilis* comprises up to 50% of the upper middle to upper Miocene faunas at Site 904, but it is not as abundant as at the shallower Site 903 (Fig. 15), reflecting its greater distance from the sediment source. The extent of the downslope transport increases from

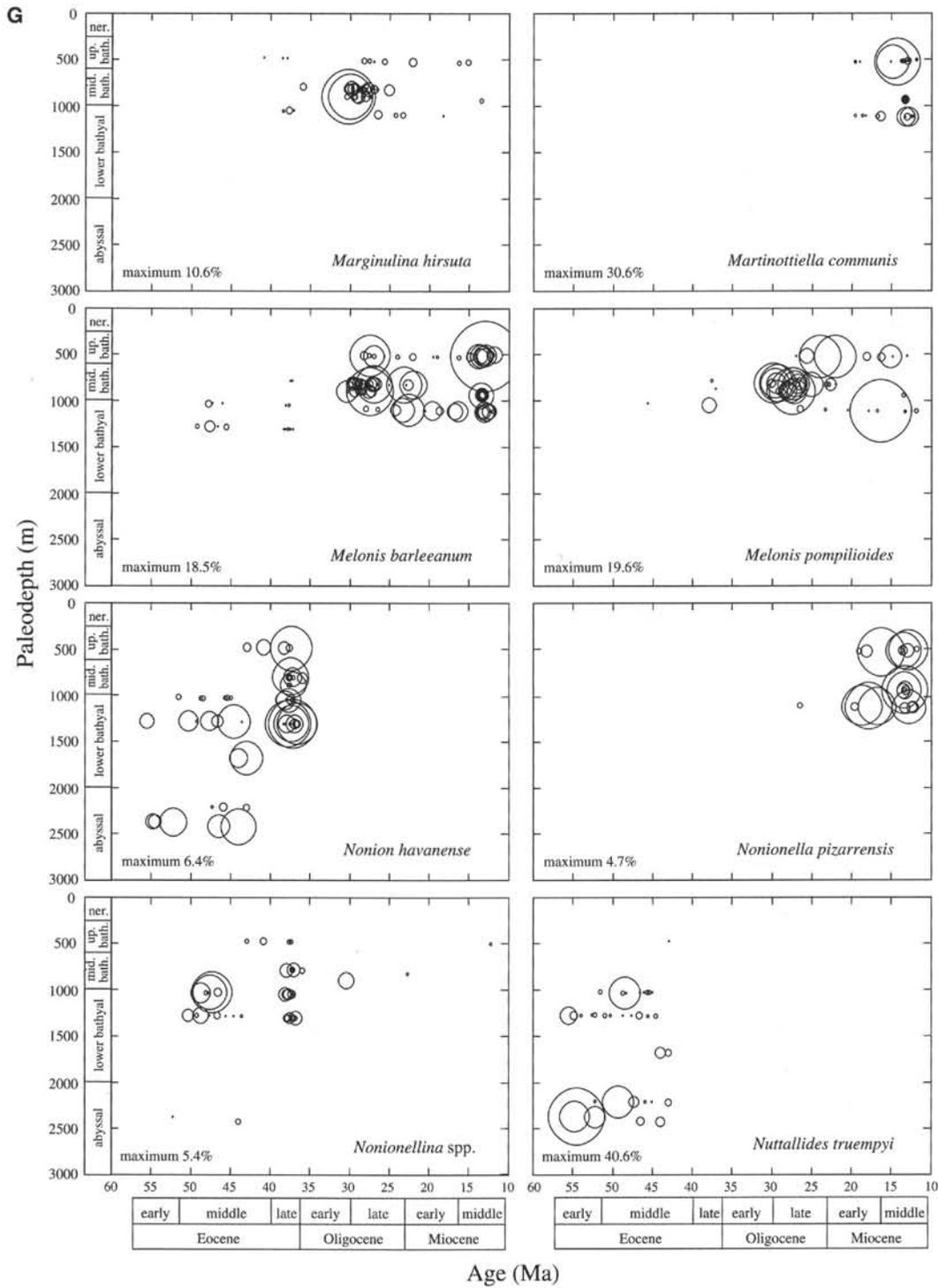


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<30% in the m2 sequence at Site 904 to >50% in the m1 (11.3–10.6 Ma) and younger sequences. The switch to dominant downslope transport at Site 904 occurred ~2 m.y. later than at Site 903 because Site 903 is more proximal to the source area. The corresponding upper middle to lower upper Miocene section at Site 902 contains no carbonate fossils, but does contain silts and clays with abundant plant and wood fragments accumulated at high rates; the input of this terrigenous debris also began at about Reflector m3 time (13.5 Ma) at

this site (Mountain, Miller, Blum, et al., 1994), concurrent with the time of canyon cutting at Site 906 (Mountain, Miller, Blum, et al., 1994; Mountain et al., this volume).

The coincident increase in transported neritic benthic foraminifers, the increase in terrigenous supply, and the first major Miocene slope canyon cutting event (Mountain, this volume) at 13.5 Ma are attributed to increased delivery of sediments to the slope from the shelf. This is indicated by seismic profiles that show that by about 13.5 Ma,

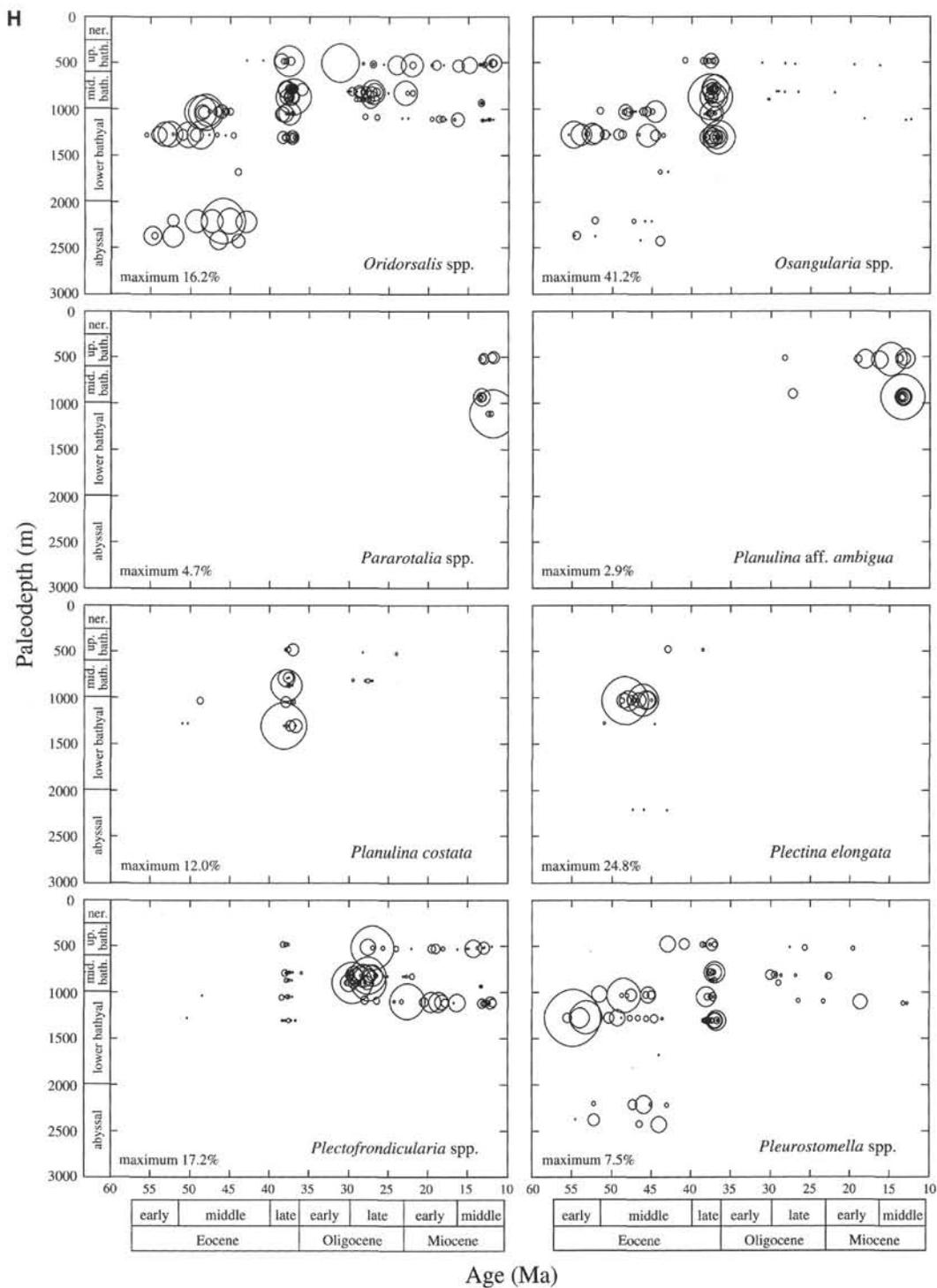


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clinoforms prograded from a location that is beneath the modern middle shelf to one that is beneath the modern outer shelf (Mountain, Miller, Blum, et al., 1994). Increased downslope transport accompanied the progradation of clinoforms from 90 km landward of Site 903 in the early Miocene (Reflectors m6–m5; see cross section in Mountain, Miller, Blum, et al., 1994) to ~60 km landward in the middle Miocene (Reflectors m4–m1). In addition, this was the time of major glacioeustatic lowerings (i.e., the Mi3 oxygen isotopic event of Mill-

er et al., 1991) and development of the permanent Antarctic Ice Sheet (Shackleton and Kennett, 1975), reducing the available accommodation space on the shelf and contributing to the increase in terrigenous flux to the slope. Transported neritic benthic foraminifers dominate the upper Miocene to the Pleistocene section (Christensen et al., this volume), reflecting the shedding of shelf sediments to the slope as clinoforms prograded to less than 20 km landward of Site 903 in the Pleistocene (Reflectors p6–p1). The benthic foraminiferal biofacies

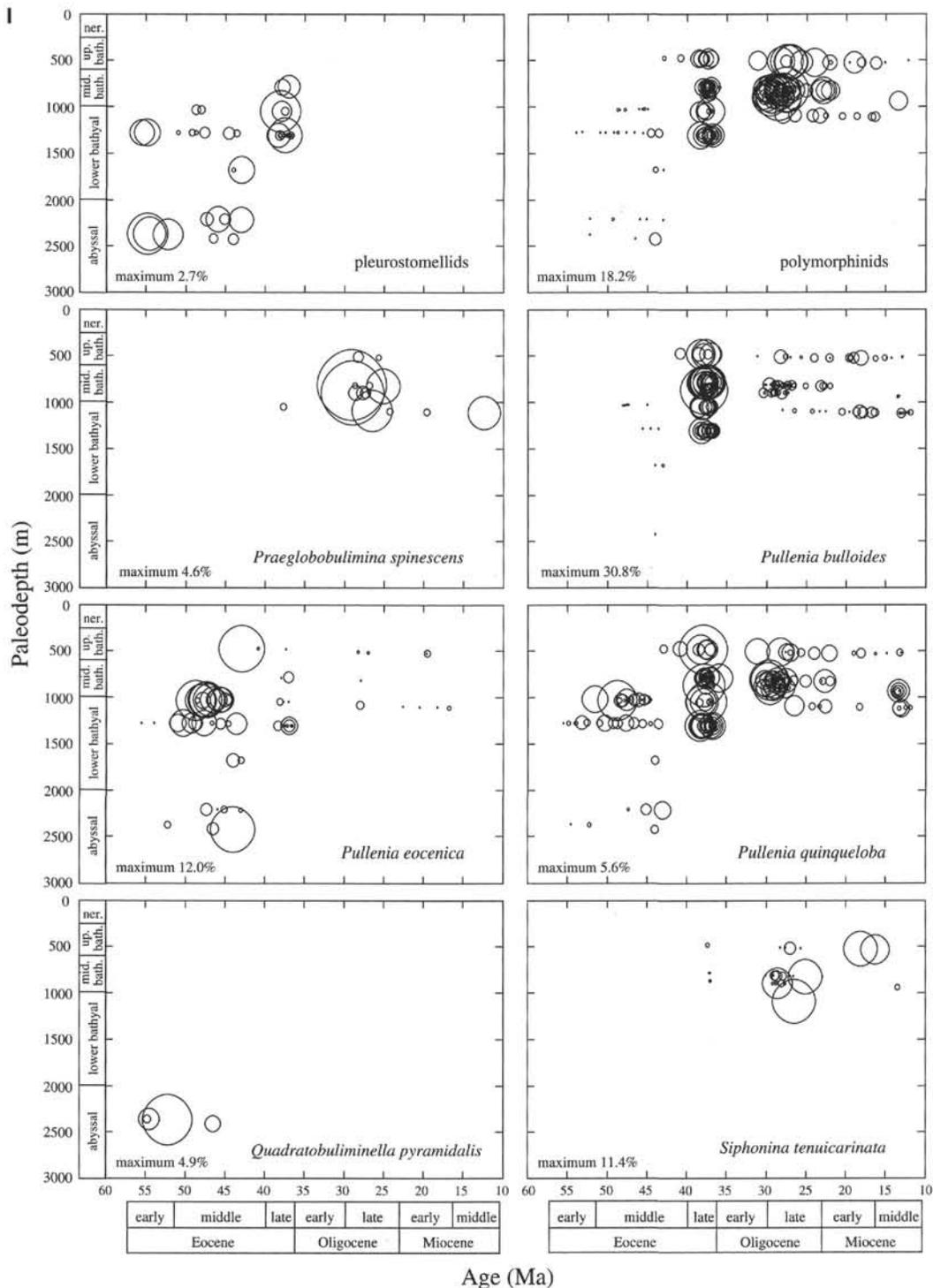


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changes at the sites studied here reflect the evolution from an Eocene carbonate-dominated ramp-type shelf and slope, to an Oligocene starved margin (both with in situ biofacies), to a prograding Miocene and younger siliciclastic-dominated margin (with progressive influence of allochthonous facies).

### SUMMARY AND CONCLUSIONS

Eocene to early middle Miocene benthic foraminifera on the New Jersey Transect comprise generally cosmopolitan deep-water faunas,

and biofacies changes primarily reflect global and regional paleoceanographic changes:

1. A bathyal biofacies shift occurred between the middle and late Eocene, correlating with a global bathyal-abyssal change associated with a late middle Eocene cooling event (e.g., Tjalsma and Lohmann, 1983; Miller et al., 1992; Thomas, 1992a).

2. Uniform late Eocene *B. alazanensis* biofacies on the New Jersey slope, in the Gulf of Mexico, and other Atlantic locations indicates that circum-Atlantic sites were ventilated by similar intermediate to upper deep-water masses at this time.

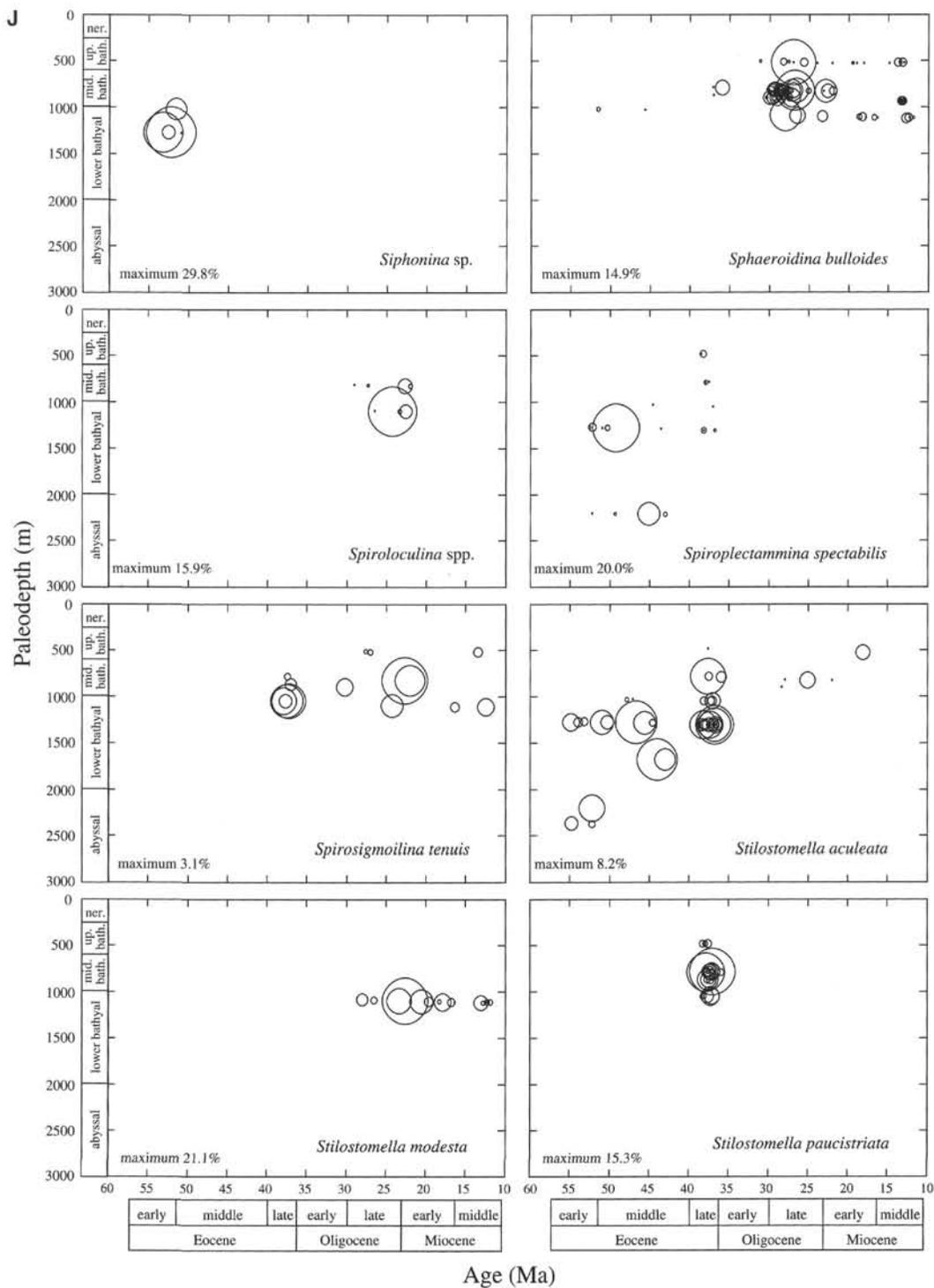


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3. A cosmopolitan *Lenticulina* spp. biofacies that dominated the Oligocene to middle Miocene bathyal zone was replaced by a *U. hispida*-*B. alazanensis*-dominated biofacies in the middle Miocene, correlating with a similar increase in *Uvigerina* spp. in the Gulf of Mexico. The faunal shift may be the regional expression of a global late early to early middle Miocene benthic foraminiferal event related to changes in deep-water sources and/or surface ocean productivity.

In addition to global and regional faunal changes, benthic foraminiferal biofacies changes at the Leg 150 slope sites exhibit varia-

tions related to sequence stratigraphy: (1) there is downslope-trans-ported shelf material above several sequence boundaries (may represent the slope expression of the shelf lowstand systems tracts) and in the upper parts of several sequences (within the slope equivalent of the shelf highstand systems tracts); (2) there is a preponderance of *U. proboscidea* in the lower parts of several sequences, which may represent in situ biofacies within the slope expression of the lowstand systems tracts; (3) *Stilostomella* spp. are more abundant in the upper parts of several sequences, which may be equivalent to the shelf highstand systems tracts; and (4) *M. pompilioides* is more abundant span-





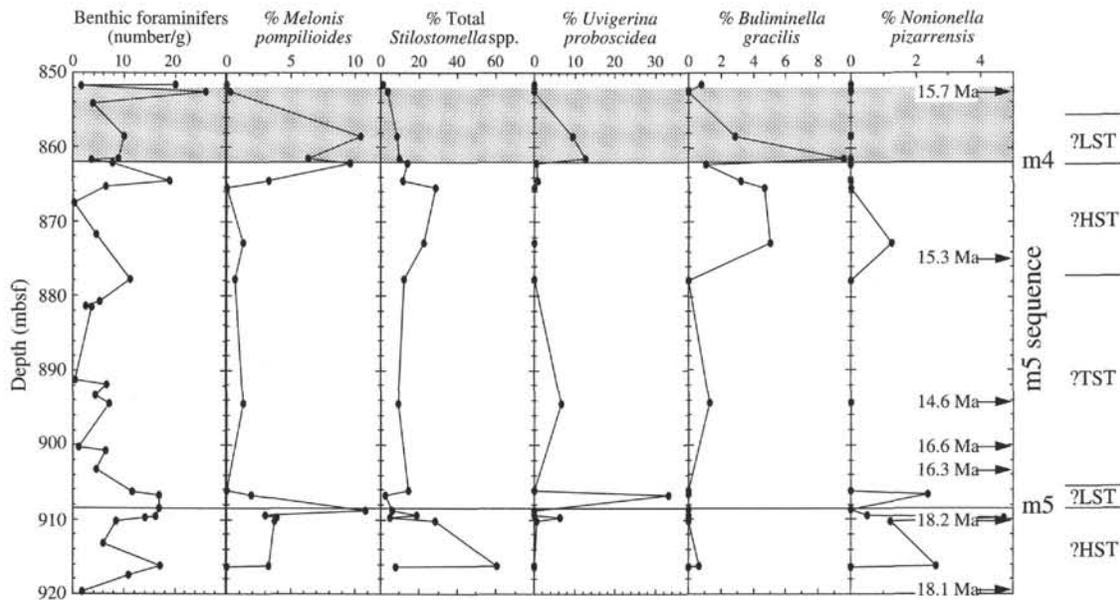


Figure 13. Detailed section across the m5 sequence at Site 903 showing the number of benthic foraminifers per gram and selected species percentages within a sequence stratigraphic framework. Shaded area represents the uncertainty in the placement of Reflector m4. Reflectors m4 (852–862 mbsf) and m5 (908.4 mbsf) correspond to sequence boundaries. LST = lowstand systems tracts; TST = transgressive systems tracts; and HST = highstand systems tracts. Ages are derived from Sr isotope stratigraphy (Miller et al., this volume).

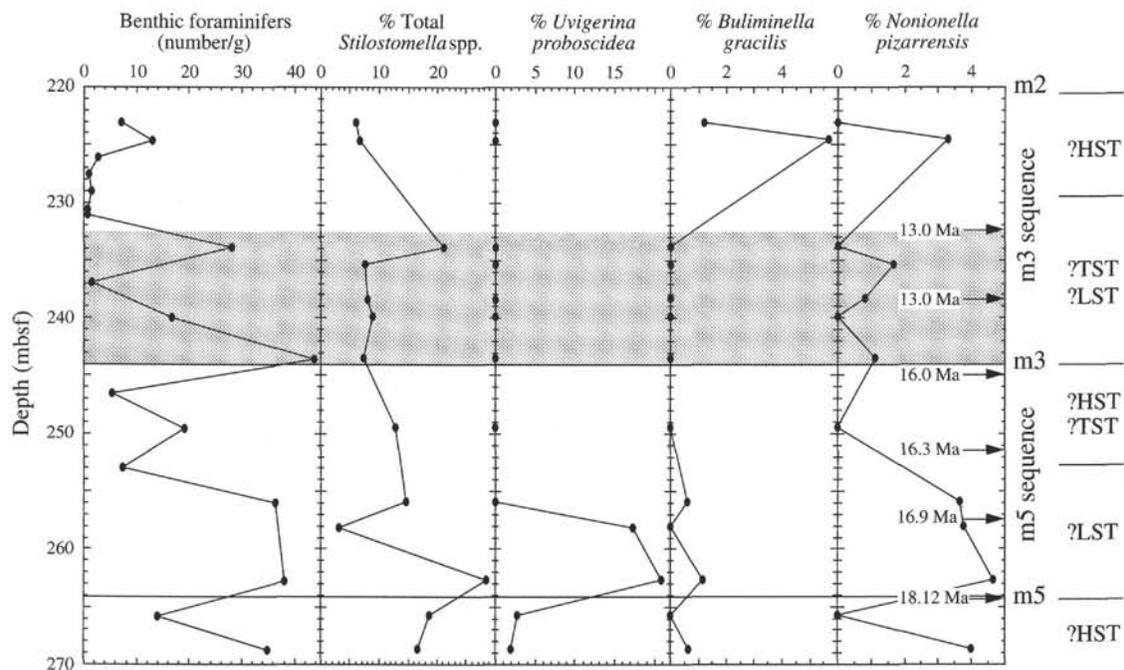


Figure 14. Detailed section across the m5 sequence at Site 904 showing the number of benthic foraminifers per gram and selected species percentages within a sequence stratigraphic framework. Shaded area represents the uncertainty in the placement of Reflector m3. Reflectors m2 (220 mbsf), m3 (232.5–244 mbsf), and m5 (264.2 mbsf) correspond to sequence boundaries. LST = lowstand systems tracts; TST = transgressive systems tracts; and HST = highstand systems tracts. Ages are derived from Sr isotope stratigraphy (Miller et al., this volume).

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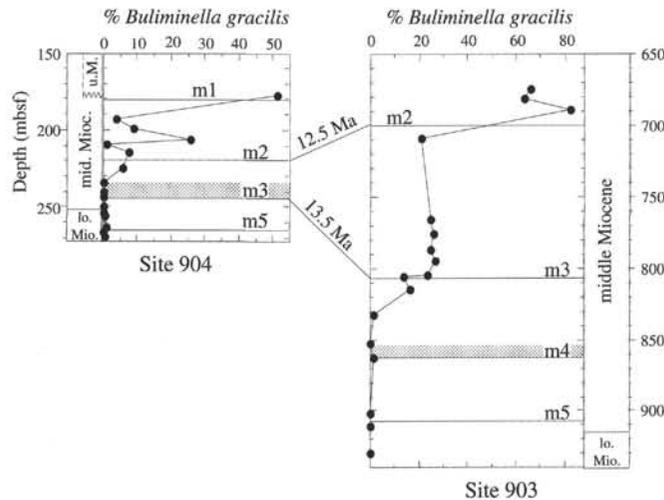


Figure 15. Percentage of *Buliminella gracilis* as a shelf transport proxy. At Sites 903 and 904, abundant downslope transport is indicated by the increase upsection of percentage *B. gracilis* between Reflectors m3 and m2.

## APPENDIX

### Taxonomic List for This Study

*Abyssamina quadrata* Schnitker and Tjalsma  
*Alabamina dissonata* (Cushman and Renz)  
*Alabamina wilcoxensis* Toulmin  
*Alabamina* sp.  
*Allomorphina paleocenica* Cushman  
*Ammodiscus* sp.  
*Amphicoryna ignota* Cushman and Siegfus  
*Angulogerina illingi* Cushman and Renz  
*Anomalinoidea aragonensis* (Nuttall)  
*Anomalinoidea capitatus* (Gumbel)  
*Anomalinoidea pseudogrosserugosus* Colom  
*Anomalinoidea semicibratus* (Beckmann)  
*Anomalinoidea spissiformis* (Cushman and Stainforth)  
*Anomalinoidea* sp. 904  
*Anomalinoidea* spp.  
*Aragonia aragonensis* (Nuttall)  
*Arenobulimina truncata* (Reuss)  
*Astacolus* spp.  
*Astrononion* sp.  
*Baculogypsina* sp.  
*Bigenerina* sp.  
*Bolivina advena* Cushman  
*Bolivina antegressa* Subbotina  
*Bolivina furcata* Cushman and Jarvis  
*Bolivina huneri* Howe  
*Bolivina* spp.  
*Bulimina aculeata* d'Orbigny  
*Bulimina alazanensis* Cushman

*Bulimina bradbury* Martin  
*Bulimina callahani* Galloway and Morrey  
*Bulimina glomarchallengeri* Tjalsma and Lohmann  
*Bulimina impendens* Parker and Bermúdez  
*Bulimina jacksonensis* Cushman  
*Bulimina macilenta* Cushman and Parker  
*Bulimina mexicana* Cushman  
*Bulimina semicostata* Nuttall  
*Bulimina* cf. *semicostata*  
*Bulimina subtruncana* Hagn  
*Bulimina trinitatis* Cushman and Jarvis  
*Bulimina tuxpamensis* Cole  
*Bulimina* sp. X  
*Bulimina* sp. A  
*Bulimina* spp.  
*Buliminella gracilis* Cushman  
*Buliminella grata* Parker and Bermúdez  
*Cassidulina crassa* d'Orbigny  
*Cassidulina havanensis* (Cushman and Bermúdez)  
*Cassidulina laevigata* d'Orbigny  
*Cassidulina* spp.  
*Cassidulinoidea* spp.  
*Ceratobulimina* sp.  
*Chilostomella* sp.  
*Chilostomella* sp.  
*Chrysalogonium* spp.  
*Cibicidoides bradyi* (Trauth)  
*Cibicidoides cookei* (Cushman and Garrett)  
*Cibicidoides crebbsi* (Hedberg)  
*Cibicidoides dickersoni* (Bermúdez)  
*Cibicidoides dohmi* Bermúdez

- Cibicidoides eocaenus* (Gümbel)  
*Cibicidoides eocaenus/praemundulus* intergrade  
*Cibicidoides grimsdalei* (Nuttall)  
*Cibicidoides grosseperforatus* van Morkhoven and Berggren  
*Cibicidoides haitiensis* (Coryell and Rivero)  
*Cibicidoides havanensis* (Cushman and Bermúdez)  
*Cibicidoides havanensis/praemundulus* intergrade  
*Cibicidoides laurisiae* (Mallory)  
*Cibicidoides micrus* Bermúdez  
*Cibicidoides mundulus* (Brady, Parker, and Jones)  
*Cibicidoides* aff. *mundulus*  
*Cibicidoides osangularis*  
*Cibicidoides pachyderma* (Rzehak)  
*Cibicidoides perlucidus* (Nuttall)  
*Cibicidoides popeliensis* Myatlyuk  
*Cibicidoides praemundulus* Berggren and Miller  
*Cibicidoides* aff. *praemundulus*  
*Cibicidoides?* *praemundulus*  
*Cibicidoides robertsonianus* (Brady)  
*Cibicidoides subhaidingerii* (Parr)  
*Cibicidoides subspiratus* (Nuttall)  
*Cibicidoides* sp. A  
*Cibicidoides* sp. C  
*Cibicidoides* sp. 13  
*Cibicidoides* spp.  
*Clinapertina complanata* Tjalsma and Lohmann  
*Clinapertina inflata* Tjalsma and Lohmann  
*Cyclammina* sp.  
*Dentalina* spp.  
*Dorothia trochoides* (Marsson)  
*Dorothia* spp.  
*Eggerella bradyi* (Cushman)  
*Eggerella* spp.  
*Ehrenbergina serrata* Reuss var. *gibbera* Galloway and Heminway  
*Favocassidulina* sp.  
*Fissurina* spp.  
*Fursenkoina makiyamai* (Chiji)  
*Fursenkoina* sp.  
*Gaudryina* spp.  
*Globobulimina notovata*  
*Globobulimina* spp.  
*Globocassidulina punctata* Berggren and Miller  
*Globocassidulina subglobosa* (Brady)  
*Gyroidinoides* sp. A  
*Gyroidinoides* sp. 1  
*Gyroidinoides* spp.  
*Hanzawaia ammophila* (Gümbel)  
*Hanzawaia mantaensis* (Galloway and Morrey)  
*Hanzawaia* spp.  
*Hoeglundina elegans* (d'Orbigny)  
*Hopkinsina* sp.  
*Karriella chapapotensis* (Cole)  
*Karriella chapapotensis* (Cole) var. *monumentensis* Mallory  
*Karriella cubensis* Cushman and Bermúdez  
*Karriella subglabra* (Guembel)  
*Karriella* sp. A  
*Karriella* spp.  
*Lagena* spp.  
*Laticarinina pauperata* (Parker and Jones)  
*Lenticulina peregrina* (Schwager)  
*Lenticulina* sp. A  
*Lenticulina* spp.  
Lenticulinids  
*Loxostomoides* cf. *applinae*  
*Loxostomoides* sp.  
*Loxostomum* spp.  
*Marginulina hirsuta* (d'Orbigny)  
*Marginulina* spp.  
Marginulinids  
*Martinottiella communis* (d'Orbigny)  
*Martinottiella* spp.  
*Massilina decorata* Cushman  
*Matanzia* sp.  
*Melonis barleeianum* (Williamson)
- Melonis pompilioides* (Fichtel and Moll)  
*Melonis* spp.  
*Nodosarella* spp.  
*Nonion havanense* Cushman and Bermúdez  
*Nonion* sp.  
*Nonionella pizarrensis*  
*Nonionella* spp.  
*Nonionellina* spp.  
*Nuttallides truempyi* (Nuttall)  
*Nuttallides* spp.  
*Nuttallinella florealis* (White)  
*Oolina* spp.  
*Oridorsalis* spp.  
*Orthomorphina* spp.  
*Osangularia* spp.  
*Pararotalia* sp.  
*Paratrochamminoides challengerii* Bronnimann and Whittaker  
*Planulina ambigua* (Franzenau)  
*Planulina* aff. *ambigua*  
*Planulina costata* (Hantken)  
*Planulina renzi* Cushman and Stainforth  
*Planulina* sp. A  
*Planulina* spp.  
*Planulina wuellerstorfi* (Schwager)  
*Plectina elongata* Cushman and Bermúdez  
*Plectina nuttalli* Cushman and Stainforth  
*Plectina* sp.  
*Plectofrondicularia kerni* Cook  
*Plectofrondicularia* sp. A  
*Plectofrondicularia* sp. Z  
*Plectofrondicularia* spp.  
*Pleurostomella* spp.  
Pleurostomellids  
Polymorphinids  
*Praeglobobulimina spinescens* Brady  
*Pseudonodosaria* spp.  
*Pullenia bulloides* (d'Orbigny)  
*Pullenia eocenica* Cushman and Siegfus  
*Pullenia quinqueloba* (Reuss)  
*Pullenia* spp.  
*Quadratobuliminella pyramidalis* de Klasz  
*Quinqueloculina* spp.  
*Rectuvigerina multicostata* (Cushman and Jarvis)  
*Rectuvigerina senni* (Cushman and Renz)  
*Rectuvigerina stonei* (Bermúdez)  
*Rectuvigerina vesca* (Finlay)  
*Saracenaria* spp.  
*Sigmoilopsis schlumbergeri* (Silvestri)  
*Sigmoilopsis* spp.  
*Siphonina* sp.  
*Siphonina tenuicarinata* Cushman  
*Siphotextularia* sp.  
*Sphaeroidina bulloides* d'Orbigny  
*Spiroloculina* spp.  
*Spiroplectammina spectabilis* (Grzybowski)  
*Spirosigmoilina tenuis* (Czjzek)  
*Spirosigmoilina* sp.  
*Stilostomella aculeata* (Cushman and Renz)  
*Stilostomella aculeata/subspinosa* intergrade  
*Stilostomella curvatura* (Cushman)  
*Stilostomella gracillima* (Cushman and Jarvis)  
*Stilostomella lepidula* (Schwager)  
*Stilostomella modesta* (Bermúdez)  
*Stilostomella nuttalli* (Cushman and Jarvis)  
*Stilostomella paucistriata* (Galloway and Morrey)  
*Stilostomella subspinosa* (Cushman)  
*Stilostomella* sp. C  
*Stilostomella* spp.  
*Textularia* spp.  
*Trifarina bradyi* Cushman  
*Trifarina danvillensis* (Howe and Wallace)  
*Trifarina* sp. X  
*Trifarina* sp. Y  
*Trifarina* sp. 2

*Tritaxia havanensis* (Cushman and Bermúdez)  
*Tritaxia paleocenica* Tjalsma and Lohmann  
*Tritaxia* spp.  
*Turrilina robertsi* (Howe and Ellis)  
*Uvigerina adelinensis* Palmer and Bermúdez  
*Uvigerina auberiana* d'Orbigny  
*Uvigerina aubertae* Boersma  
*Uvigerina carapitana* Hedberg  
*Uvigerina chirana* Cushman and Stone  
*Uvigerina cocoensis* Cushman  
*Uvigerina elongata* Cole  
*Uvigerina havanensis* Cushman and Bermúdez  
*Uvigerina hispida* Schwager

*Uvigerina mexicana* Nuttall  
*Uvigerina pigmea* d'Orbigny  
*Uvigerina proboscidea* Schwager  
*Uvigerina? proboscidea*  
*Uvigerina spinulosa* Hadley  
*Uvigerina* sp. A  
*Uvigerina* spp.  
*Vaginulina* sp.  
*Vaginulinopsis baggi* McLean  
*Vaginulinopsis* spp.  
*Virgulina* sp.  
*Vulvulina spinosa* Cushman  
*Vulvulina* spp.